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A new species of *Perinereis* Kinberg, 1865 (Annelida: Nereididae) from the Western Mediterranean Sea revealed by morphological and molecular approaches

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Abstract

This study uses morphological and molecular evidence to describe a new intertidal species of *Perinereis* Kinberg, 1865 (Annelida: Nereididae) from northeast Algeria. *Perinereis louizomarum* n. sp. belongs to Subgroup 1A, which is distinguished by a single bar-shaped paragnath on area VI of the proboscis and a dorsal ligule that is either slightly or not expanded in the posterior parapodia. The new species differs from *P. cultrifera* and similar congeners by having a bare area V and slightly enlarged posterior dorsal ligules. Furthermore, using newly generated and mined sequences from online databases, and mined sequences of *Perinereis* species from online, the analysis of cytochrome c oxidase subunit 1 (COI) mitochondrial gene sequences supported its status as new species. An identification key of the *Perinereis* species belonging to Subgroup 1A is also provided.

Keywords: Algeria; description; mtCOI; *Perinereis cultrifera*; Systematics.

Introduction

In terms of environmental and socio-economic importance, *Perinereis* Kinberg, 1865 is possibly the most important nereidid genus. Species such as *P. aibuhitensis* (Grube, 1878), *P. cultrifera* (Grube, 1840), and *P. linea* (Treadwell, 1936) are utilized in fishing and aquaculture industries (Lee *et al.*, 1992; Rouabah, 2003; Arias *et al.*, 2013), whereas some others, for instance, *P. gualpensis* (Jeldes, 1963) and *P. nuntia* (Savigny in Lamarck, 1818), are regarded as biomarkers in aquatic environmental contamination (Díaz-Jaramillo *et al.*, 2010; Rhee *et al.*, 2012). *Perinereis* species can also be found in a wide range of marine ecosystems, from shallow waters to abyssal zones (Wu *et al.*, 1985), although some dwell in estuarine or even semi-terrestrial environments (Glasby *et al.*, 2009; Villalobos-Guerrero *et al.*, 2021).

To facilitate practical identification, this heterogeneous and speciose nereidid genus has traditionally been divided into six informal morphological groups (1A, 1B, 2A, 2B, 3A, and 3B) (Hutchings *et al.*, 1991). In particular, the *Perinereis* Subgroup 1A (hereafter SG-1A) was proposed for species that share the presence of a single

bar-shaped paragnath on area VI and posterior chaetigers with dorsal ligules slightly or not expanded; twelve of the species are currently valid: *Perinereis arabica* (Mohammad, 1971) from Kuwait, *P. calmani* (Monro, 1926) from Australia and Macclesfield Bank (South China Sea), *P. cultrifera* (Grube, 1840) from the Tyrrhenian Sea (Italy), *P. dongalae* (Horst, 1924) from Indonesia, *P. falso-variegata* (Monro, 1933) from South Africa, *P. floridana* (Ehlers, 1868) from the Gulf of Mexico (USA), *P. helleri* (Grube, 1878) from the Philippines, *P. rullieri* (Pilato, 1974) from the Mediterranean Sea (Italy), *P. taorica* (Langerhans, 1881) from Madeira, *P. tenuisetis* (Fauvel, 1915) from the Mediterranean Sea (Italy), *P. villalobosi* (Rioja, 1947) from the Gulf of California, and also *P. capensis* (Kinberg, 1865) from South Africa, which was formerly overlooked by Hutchings *et al.* (1991).

Perinereis cultrifera is the most studied and culturally significant *Perinereis* SG-1A species in the Mediterranean Sea, particularly Algeria. It is widely exploited on the Northeast Algerian coast as bait in sport angling and recreational fishing for sea bream (*Dicentrarchus labrax*), sole (*Solea solea*), and other varieties of fishes, such as pageot (*Pagellus erythrinus*), small wolf (*Dicentrarchus*

labrax) and marbled (*Lithognathus mormyrus*) (Rouabah, 2003). However, Scaps *et al.* (2000) and Rouabah & Scaps (2003a) revealed that *P. cultrifera* might be a complex of species based on the morphological and biochemical divergence of specimens collected in Saint-Cloud, Annaba (Algeria) and from Saint-Aubin-sur-Mer, Normandy (English Channel). Similarly, Rouabah *et al.* (2009) also found that the spawning season, mode of reproduction, age of maturity, and biometric characteristics of specimens assigned to *P. cultrifera* differ according to the geographic location of the populations in question. Hence, it is likely that two (or more) species are involved under the same species name, *P. cultrifera*, in the Mediterranean Sea and the eastern North Atlantic.

Moreover, a total of eight species of *Perinereis* have been reported from the Mediterranean Sea: *Perinereis cultrifera* (Grube, 1840) from the Tyrrhenian Sea, Italy; *P. floridana* (Ehlers, 1868) from Florida, USA; *P. linea* (Treadwell, 1936) from Xiamen, China; *P. macropus* (Claparède, 1870) from the Tyrrhenian Sea, Italy; *P. marionii* (Audouin & Milne Edwards, 1833) from Bay of Biscay; *P. oliveirae* (Horst, 1889) from Portugal; *P. rullieri* (Pilato, 1974), and *P. tenuisetis* (Fauvel, 1915) from Sicily, Italy (Fauvel, 1915; Prevedelli *et al.*, 1990; Scaps *et al.*, 2000; Rouabah & Scaps 2003a,b; Rouabah & Rouabah 2007; Rouhi *et al.*, 2008; Arias *et al.*, 2013; Meghlaoui *et al.*, 2015; Gasmi *et al.*, 2016; Gillet, 2017; Bakalem *et al.*, 2020; Villalobos-Guerrero *et al.*, 2021; Rezzag-Mahcene *et al.*, 2022). Only four species (*P. cultrifera*, *P. macropus*, *P. rullieri*, and *P. tenuisetis*) were originally described from the Mediterranean Sea. Two other species have been considered alien in the Mediterranean Sea: *Perinereis linea* (Arias *et al.*, 2013) and *P. nuntia* (Zenetos *et al.*, 2010); however, the presence of the former species in the region was recently questioned (Villalobos-Guerrero *et al.*, 2021), whereas *P. nuntia* has been demonstrated as a complex of morphologically distinguishable species (Wilson & Glasby, 1993; Glasby & Hsieh, 2006), even between neighboring regions (Villalobos-Guerrero, 2019; Tosuji *et al.*, 2019). The records

of the other *Perinereis* species in the Mediterranean Sea need to be thoroughly assessed because it is likely that nominal species might be hidden in species complexes or correspond to new ones.

The combined use of morphology and DNA evidence is a powerful tool for disentangling nereidid species complexes or discovering previously unknown species (Glasby *et al.*, 2013; Sampértegui *et al.*, 2013; Kara *et al.*, 2020; Tosuji *et al.*, 2019; Drennan *et al.*, 2021; Wang *et al.*, 2021; Teixeira *et al.*, 2022). In particular, the mitochondrial cytochrome c oxidase I (COI) gene has been used to explain the interspecific relationships between nereidid species and their phylogeographic relationships; hence, used to analyze population structure, evolutionary relatedness, and delimitation of species (e.g., Park & Kim, 2007; 2017; Villalobos-Guerrero & Carrera-Parra, 2015; Kara *et al.*, 2018; Villalobos-Guerrero *et al.*, 2021; Villalobos-Guerrero *et al.*, 2022).

The current study describes a new species from the Algerian coast that belongs to the *Perinereis* SG-1A genus. Morphological comparisons were made between *Perinereis* species from the Mediterranean Sea and those from SG-1A. In addition, partial sequences of the new species' COI region were amplified to analyze genetic distances and compared to molecular data from other *Perinereis* species.

Material and Methods

Sampling sites

Nereidid specimens were collected from three stations on the northeastern coast of Algeria: El-Kala, Annaba, and Skikda (Fig. 1; Table 1). At each station, three replicates were sampled monthly during 2019 using a quadrat (25 cm × 25 cm) at 0.5 m depth on algae attached on hard bottoms at the intertidal and shallow sublittoral. The algae were scraped and deposited in containers to search for nereidid individuals. Several specimens were

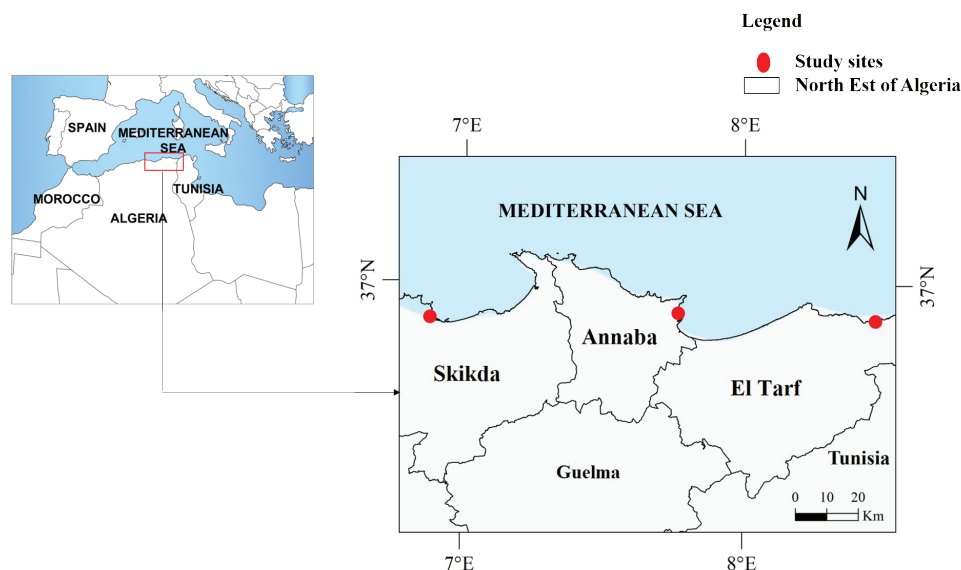


Fig. 1: Map of the eastern coasts of Algeria showing the three sampling sites of this study.

Table 1. Location and characterization of sampling sites.

Station	Locality	Coordinates	Biotopes
La Montagne	El-Kala	36.89815, 8.450911	Rocky shore, 0.5 m depth
Rezgui Rachid	Annaba	37.53547, 8.289361	Rocky and sandy shore, 0.5 m depth
Bikini	Skikda	36.87191, 6.900911	Rocky and sandy shore, 0.5 m depth

found fragmented, with either the anterior or posterior end missing. Consequently, we also carried out a second method to prevent mechanical fragmentation. A small amount of powdered alum made of Kalunite was sprinkled on the rocky ground, which is natural potassium and does not contain aluminum hydroxide. No scientific studies to date have demonstrated natural alum stone toxicity. Before fixation, the specimens were forced to evert the proboscis by exerting pressure gently on the prostomium, which eventually facilitated the observation of the paragnaths through the microscope. The specimens were fixed individually in 1.5 ml microcentrifuge tubes with a 4% formalin solution and then preserved in 70% ethanol. After identification, the annelids intended for the genetic study were stored individually in 1.5 ml tubes containing ethanol at 90% and placed at a temperature of 4°C. Finally, all specimens examined in this study, including the type material, were deposited at the *Muséum National d'Histoire Naturelle* (MNHN), Paris.

Morphological observation

Examination of the morphological features of the specimens, terminology, and standards to describe the species, and preparation and edition of figures are detailed elsewhere (Villalobos-Guerrero *et al.*, 2021 and references there cited). In addition, paired areas in the pharynx were indicated as 'a' for the left and 'b' for the right. Specimens were photographed using a digital camera (ZEISS AxioCam ICc1) and mounted on a stereomicroscope and compound microscope with a portable adaptor. Description of the species is based upon the morphology of the holotype unless otherwise stated, while variations are indicated separately for all the material. Anomalies found in one specimen are also described.

DNA extraction, PCR amplification and molecular analysis

Partial sequences of mitochondrial cytochrome c oxidase subunit I (COI) region were amplified to analyze the

genetic distances at the intraspecific level of *Perinereis* specimens collected in Algeria and to compare this data with other *Perinereis* species with sequences mined from BOLDSystems (BOLD) (Sujeevan & Hebert, 2007). DNA was extracted from tissue samples conserved in 90% ethanol using CTAB according to the protocol established by Winneppenninckx *et al.* (1993). The universal primer pair LoboF1 and LoboR1 (Lobo *et al.*, 2013) was used to amplify a fragment of the gene COI (Table 2). Polymerase Chain Reaction (PCR) amplifications were carried out using 12 µl of Taq DNA polymerase (Thermo Scientific), 14.88 µl of molecular biology grade water, 2.5 mM of MgCl₂, 0.2 mM of each dNTP, 0.2 µM of each primer (10 µM) and 1 µl of template DNA to make up a total reaction volume of 25 µl. Thermal cycling conditions were settled as follows: initial denaturation at 94°C for 5 min, followed by 5 cycles of 94 °C for 30 s, 45°C for 1 min 30 s, and 72°C for 1 min, then by 45 cycles of 94°C for 30 s, 54°C for 1 min 30 seconds, and 72°C for 1 min and final extension time at 72°C for 5 min. Amplicons were Sanger sequenced at the GENEWIZ®, just the forward primer (LoboF1). Agarose gels with BET were used to control PCR product size. The products were then electrophoresed, one direction was sequenced for eight products, and only one was sequenced in two directions. All sequences generated were deposited in GenBank under accession numbers OP459427–OP459435, OP585368 (Table 3).

The obtained sequences were checked manually with the Bioedit sequence editor (Hall, 1999). After checking individual 1, no difference was observed between the sequencing of the two strands. Both sequences (forward and reverse) were used to build a consensual sequence, which was compared with the other eight individuals. Sequences were aligned using ClustalW implemented in MEGA-X (Kumar *et al.*, 2018). Nucleotide-sequence divergences of the partial sequences of the COI region were used to calculate the intraspecific and interspecific pairwise genetic distances using the Kimura two-parameter (KP2) model (Kimura, 1980). This newly obtained data for Algerian specimens of *Perinereis* was compared with mtCOI sequences of other species mined from BOLD: *Perinereis aibuhitensis*, *P. brevicirris*, *P. cultrifera*, *P.*

Table 2. Primer pair used to amplify COI sequences of *Perinereis* specimens in this study.

Reference	Primer	Direction (5' – 3')
Lobo <i>et al.</i> (2013)	LoboF1	(F) KBTCHACAAAYCAYAARGAYATHGG
	LoboR1	(R) TAAACYTCWGGRTGWCCRAARAAYCA

Table 3. Species of *Perinereis* and outgroups used in this study and locality of sequences with respective GenBank accession numbers.

Species	GenBank COI	Location	n	Reference
<i>P. aibuhitensis</i> (Grube, 1878)	KC800611 KC800612 KC800613 KC800614	China	4	Deng, unpubl. data
	KY129885	China	1	Chen <i>et al.</i> , unpubl. data
	KF611806	Korea	1	Kim <i>et al.</i> (2015)
<i>P. brevicirris</i> (Grube, 1866)	JX966314	India	1	Sekar <i>et al.</i> (2016)
	JX503024 JX503025 JX503026	South Korea	3	Kim <i>et al.</i> , unpubl. data
	KC800628 KC800630	China	2	Deng, unpubl. data
	MN812983	Arcacha–France Portugal	1	Alves <i>et al.</i> , 2020
<i>P. cultrifera</i> (Grube, 1840)	KR916910 KR916911 KR916912	Portugal	3	Lobo <i>et al.</i> , 2016
	MN256545 MN256546	Unknown	2	Xing & Zhang, unpubl. data
<i>P. euinii</i> Park & Kim, 2017	KC800624 KC800625 KC800627	China	3	Deng, unpubl. data
	KY129883	China	1	Chenet <i>et al.</i> , unpubl. data
<i>P. louizomarum</i> n. sp.	OP459427 OP459428 OP459429 OP459430 OP459431 OP459432 OP459433 OP459434 OP459435	Algeria	9	This study
<i>P. suluana</i> (Horst, 1924)	JX420245 JX420246 JX420247 JX420251 JX420252 JX420253	Australia	6	Glasby <i>et al.</i> (2013)
<i>P. vallata</i> (Grube & Kröyer in Grube, 1858)	JX676143 JX676159 JX676180	India	3	Iyyapparajanarasimappallavan <i>et al.</i> , unpubl. data
	HQ705192 HQ705194 HQ705195	Chile	3	Sampertegui <i>et al.</i> (2013)
<i>P. wilsoni</i> Glasby & Hsieh, 2006	KC800623 KC800629 KC800631	China	3	Deng, unpubl. data
	KY129887 KY129888 KY129889	China	3	Chen <i>et al.</i> , unpubl. data
<i>Nereis pelagica</i> (Linnaeus, 1758)	HQ023615	Canada, Manitoba	1	Carr <i>et al.</i> , 2011
<i>Myrianida rubropunctata</i> (Grube, 1860)	GQ856203	Ferrol, Spain	1	Nygren & Pleijel (2010)

suluana, *P. wilsoni*, and *P. vallata*, including *Nereis pelagica* (Linnaeus, 1758) and *Myrianida rubropunctata* (Grube, 1860) (Syllidae) as outgroups (Table 3). These species were chosen because the number of BOLD available sequences is equal to or greater than that obtained in this study for *Perinereis* Algerian individuals. Based on the K2P model, the same sequences were used to calculate intraspecific genetic distances.

Results

Molecular analyses

A total of 581 bp of the partial mtCOI sequences was analyzed from nine *P. louizomarum* n. sp. individuals. The intraspecific pairwise distances between the COI sequences were relatively low (Table 4), with a mean intraspecific distance of 0.02%. The nine sequences represent three haplotypes. Six annelids shared the most abundant sequence. Neither insertion nor deletion was detected between the nine individual sequences. The interspecific pairwise distances based on six *Perinereis* species showed mean values between 0.173 and 0.276; however, for *P. louizomarum* n. sp., they varied from 0.262 to 0.325 (Table 5). According to the Neighbour-Joining tree construction, all the *Perinereis* species formed distinct clusters, except *P. brevicirris*, separated into three clades. In particular, *P. louizomarum* n. sp. clustered in a genetically different group with a high bootstrap value (Fig. 2). The morphological and molecular evidence obtained in this study allows us to confirm that the latter is a new species.

Morphology

A new *Perinereis* species was discovered among the rocky shores of Northeast Algeria. *Perinereis louizomarum* n. sp. has a single bar-shaped paragnath on area VI of the proboscis, and the proximal dorsal ligule is slightly expanded. The new species suits within the Subgroup 1A *sensu* Hutchings *et al.* (1991). *Perinereis louizomarum* n. sp. closely resembles the Mediterranean Sea species *P. cultrifera* and *P. rullieri*. However, morphological comparisons based on the descriptions, redescrptions, and illustrations of the type specimens (*vide* Grube, 1840; Pilato, 1974; Hutchings *et al.*, 1991; Park & Kim, 2017) reveal that *P. louizomarum* n. sp. can mainly be differentiated from both species by the counting of paragnaths on areas III and V, the form of distal dorsal ligules, and the location of dorsal cirri.

SYSTEMATICS

Family NEREIDIDAE de Blainville, 1818

Genus *Perinereis* Kinberg, 1865

Perinereis Kinberg, 1865: 175; 1910: 52.

Type species

Perinereis novaehollandiae Kinberg, 1865, by subsequent designation (*vide* Hartman, 1949). It is considered a junior synonym of *P. amblyodonta* Schmarda, 1861 (Ehlers, 1904; Hartman, 1959).

Table 4. Intraspecific pairwise distance of *Perinereis louizomarum* n. sp. calculated by Kimura-2-Parameters model.

	Ind. 1	Ind. 3	Ind. 4	Ind. 5	Ind. 6	Ind. 7	Ind. 8	Ind. 9
<i>Ind. 3</i>	0.0250							
<i>Ind. 4</i>	0.0163	0.0330						
<i>Ind. 5</i>	0.0162	0.0473	0.0127					
<i>Ind. 6</i>	0.0179	0.0538	0.0079	0.0046				
<i>Ind. 7</i>	0.0162	0.0554	0.0079	0.0108	0.0060			
<i>Ind. 8</i>	0.0163	0.0440	0.0047	0.0077	0.0031	0.0031		
<i>Ind. 9</i>	0.0130	0.0406	0.0111	0.0093	0.0077	0.0093	0.0062	
<i>Ind. 10</i>	0.0365	0.0238	0.0393	0.0517	0.0626	0.0641	0.0451	0.0484

Table 5. Interspecific pairwise genetic distances of *Perinereis* species (K2P distances).

Species	<i>P. aibuhitensis</i>	<i>P. brevicirris</i>	<i>P. cultrifera</i>	<i>P. suluana</i>	<i>P. vallata</i>	<i>P. wilsoni</i>
<i>P. brevicirris</i>	0.241					
<i>P. cultrifera</i>	0.274	0.276				
<i>P. suluana</i>	0.267	0.270	0.257			
<i>P. vallata</i>	0.222	0.222	0.251	0.254		
<i>P. wilsoni</i>	0.235	0.173	0.264	0.237	0.255	
<i>P. louizomarum</i> n. sp.	0.325	0.285	0.268	0.282	0.262	0.282

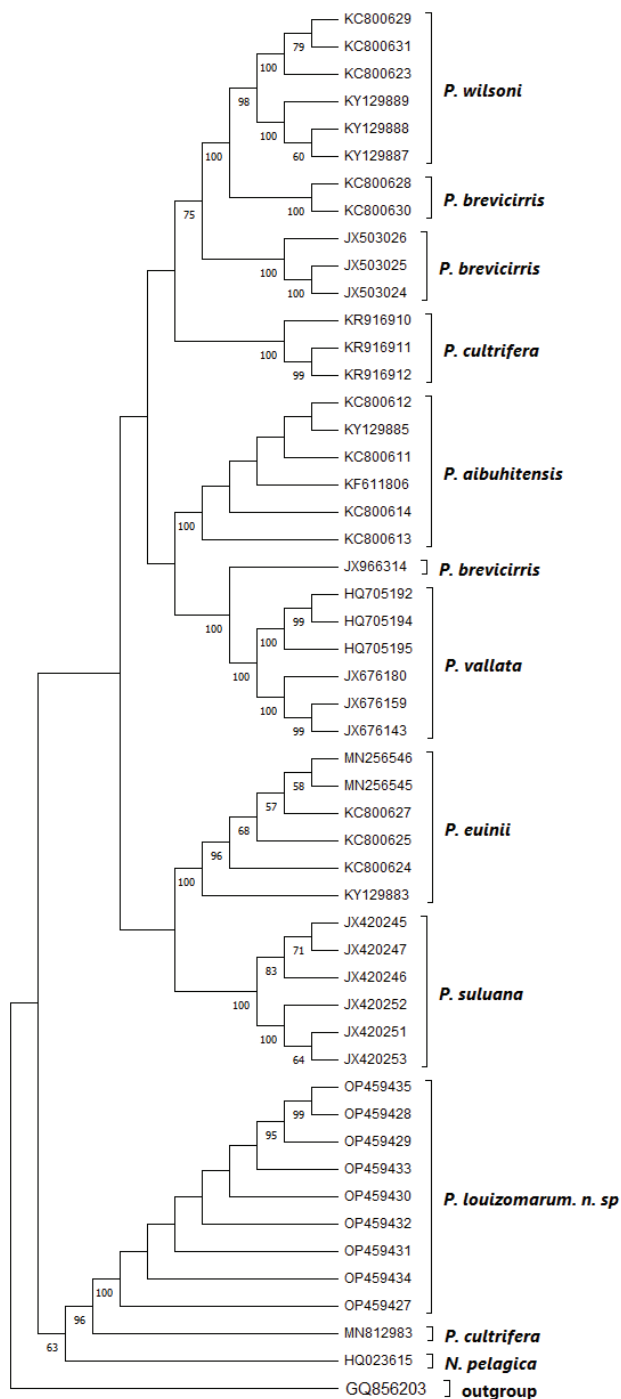


Fig. 2: Neighbor-Joining tree generated with MEGA-X based on COI sequence of nine species of *Perinereis*. The gene divergence was represented using Kimura's two-parameter (K2P) model.

Perinereis louizomarum n. sp.

urn:lsid:zoobank.org:act:A068FBA0-EB4D-4239-A98B-F4D1EF12ADBE

[Argelian name: *ver vert de roche*]

Figures 4, 5

Type material. Mediterranean Sea, Algeria. Holotype: MNHN-IA-TYPE 2065, El-Kala (36.89815° N, 8.450911° W), Annaba, Skikda, 22 May 2019, coll. H. Rezzag Mahcene, 0.5 m, substratum covered by algae,

rock, and sand, atokous female, in good condition.

Paratypes. Mediterranean Sea, Algeria. Six specimens (MNHN-IA-TYPE 2066), El-Kala, Algeria (36.89815° N, 8.450911° W), 02 May 2019, coll. H. Rezzag Mahcene, 0.5 m, among rocks covered by algae, atokous. Two specimens, one of them contains only the proboscis (MNHN-IA-TYPE 2067), El-Kala, 24 March 2019, 0.5 m, among rocks covered by algae, atokous, coll. H. Rezzag Mahcene. One specimen (MNHN-IA-TYPE 2068), Skikda (36.87191° N, 6.900911° W), 25 January 2020, coll. H. Rezzag Mahcene, 0.5 m, among rocks covered by algae, atokous.

Additional material. Mediterranean Sea, Algeria. One specimen (MNHN-IA-TYPE 2069), La Montagne Beach, El-Kala, Algeria (36.89815° N, 8.450911° W), 02 May 2019. Two specimens (MNHN-IA-TYPE 2070), Rezgui Rachid Beach, Annaba, Algeria (37.53547° N, 8.289361° W), 13 March 2019.

Diagnosis. *Perinereis* species belonging to subgroup 1A. Specimens with a crescent-shaped bar on area VI; areas VI–V–VI ridge pattern π -shaped; area V without paragnaths; areas VII–VIII with anterior band of paragnaths consisting of two rows; area III without laterally isolated paragnaths; posterodorsal tentacular cirri extending to chaetigers 4–6; distal dorsal ligule subequal in length throughout body; neuroacicular ligule markedly projected; falcigers with camerated shaft divided into four partitions.

Description. Holotype atokous female, complete, in good condition, 56 mm LT, 12 mm L10, 3.2 mm W10, with 58 chaetigers. Body color brownish (Fig. 3A), with brown pigmentation on dorsum of prostomium (except mid-posteriorly; Fig. 3B), inner part of palpophores (Fig. 3A), proximal half of palpostyles and antennae (Fig. 3A, B), entire cirrophores of tentacular cirri (Fig. 3A, B), pygidium, pygidial cirri and on dorsum of all segments, more intense posteriorly.

Prostomium campanulate, as long as wide (Fig. 3B); anterior end broad, distally complete; anterolateral gap between antenna and palpopore narrow, three-quarters as wide as basal diameter of antennae Palpopores sub-conical, thick, as long as wide, two-thirds of entire prostomium (Fig. 3A); sub-distal transverse groove distinct, deeply embedded. Palpostyles ovoid, large, one-third as wide as diameter of palpopore (Fig. 3A).

Antennae tapered, thick, extending forwards slightly beyond palpopore and posteriorly to distal third of length of prostomium; antennae separated with gap as wide as three-quarters of basal diameter of antennae (Fig. 3A, B).

Paired eyes black, arranged in trapezoid form, gap between both pairs twice as wide as diameter of posterior pair of eyes (Fig. 3A, B); anterior pair of eyes oval, subequal to basal diameter of antennae, with gap between both eyes 5.5 times their diameter, with lens barely distinct, purplish, covering 40% of eye; posterior pair of eyes rounded, slightly narrower than basal diameter of antennae, with lens distinct, purplish, placed centrally in eyes and covering 40% of it.

Tentacular belt 3 times wider than long, 1.5 times as long as chaetiger 1, with even anterior margin, dorsum

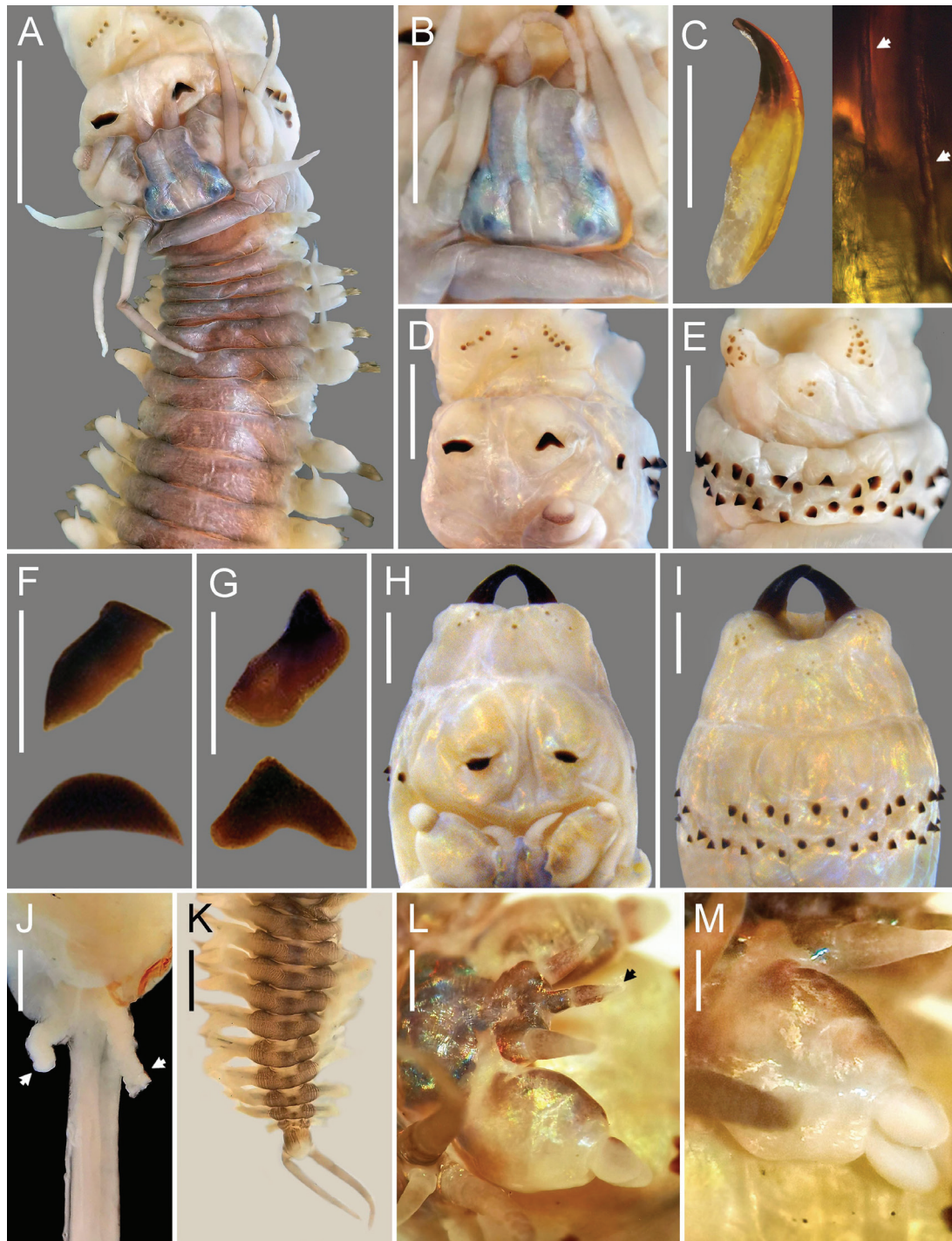


Fig. 3: *Perinereis louizomarum* n. sp. (A–G, K) Holotype (MNHN-IA-TYPE 2065) from El-Kala, Algeria, atokous: (A) Anterior end in dorsal view; (B) prostomium in dorsal view; (C) right jaw in dorsal view, inset of inner canals indicated by arrows; (D) proboscis in dorsal view; (E) proboscis in ventral view; bar-shaped paragnath of (F) left side and (G) right side of area VI, above in front-lateral view and below in frontal view; (K) pygidium in dorsal view. (H, I) Paratype (MNHN-IA-TYPE 2066) from El-Kala, Algeria, atokous: proboscis in (H) dorsal and (I) ventral views. (J) Paratype (MNHN-IA-TYPE 2066) from El-Kala, Algeria, atokous female: paired oesophageal caeca in dorsal view (indicated by arrows). (L, M) Paratype (MNHN-IA-TYPE 2068) from Skikda, Algeria, atokous: (L) Prostomium in lateral view with third anomalous antenna (indicated by arrow); (M) duplicate right palphophore and palpostyle. Scale bars: A: 3.5 mm; B: 1.7 mm; C: 5 mm; D, E, H, I, J: 1.2 mm; F, G, K: 8 mm; L: 1 mm; M: 0.8 mm.

without marked transverse wrinkles.

Tentacular cirri slender, smooth (Fig. 3A); postero-dorsal cirri extending posteriorly to chaetiger 2, 1.5 times as long as anterodorsal cirri; anterodorsal cirri extending posteriorly to chaetiger 1; posteroventral cirri extended over first third of prostomium; anteroventral cirri as long

as two-thirds of posteroventral cirri and as long as palphophore; cirrophores of anteroventral cirri ring-shaped, remaining cirrophores cylindrical, posterodorsal cirrophores 1.5 times as long as anterodorsal cirrophores, anteroventral cirrophores broadest, posteroventral cirrophores narrowest.

Proboscis everted, with maxillary and oral rings cylindrical, wider than long. Jaws edentulate, with distal half dark reddish, remaining yellow amber; 2 canals emerging from pulp cavity (Fig. 3C).

Proboscis with brownish, wear and small paragnaths on maxillary ring and dark-red and coarse paragnaths on oral ring; consisting of cones, except bars on area VI and areas VII–VIII (Fig. 3D–I). Area I: 2, longitudinal row of cones, distal one smaller. Areas IIa: 5 and IIb: 8, one or two irregular rows of uneven cones in oblique patch. Area III: 4, four slightly uneven cones in subcircular patch, without distinct isolated lateral groups. Areas IVa: 12 and IVb: 17, three irregular transverse rows of uneven cones in slightly curved patch. Area V: 0. Areas VIa: 1 and VIb: 1, crescent-shaped bars (Fig. 3F), right with concave base but asymmetrical and pointed (Fig. 3G). Areas VII–VIII: 38, two well-separated bands of coarse and uneven paragnaths, with anterior band consisting of two transverse nearly-aligned rows of cones and p-bars (Fig. 3E, I) (furrow rows with one p-bar, ridge rows with one cone), posterior band with two transverse rows of cones displaced from each other (paragnaths present only on ridge rows, with two cones on each region). Ridges of areas VI–VIII with π -shaped pattern (Fig. 3D, H). Gap between areas VI and VIII broad, as wide as palpophore (Fig. 3D, H). Paired oesophageal caeca present (Fig. 3J).

Parapodia without glandular patches on dorsum of segments (Fig. 4K). Notopodia consisting of dorsal cirrus, dorsal ligule (distal and proximal), notopodial prechaetal lobe and median ligule in biramous parapodia (Fig. 4A–E).

Dorsal cirri conical, thick, extending beyond distal region of dorsal ligule in anteriormost and anterior parapodia (Fig. 4A, B), shorter than that in middle and posterior parapodia (Fig. 4C–E), then subequal in posteriormost parapodia; dorsal cirri 3–3.5 times as long as proximal region of dorsal ligule in anteriormost parapodia (Fig. 4A), 2–2.5 times as long as that in anterior parapodia (Fig. 4B), subequal to or slightly longer than that in following parapodia (Fig. 4C–E); attached basally to dorsal ligule in anteriormost parapodia (Fig. 4A), one-third in anterior parapodia (Fig. 4B), medially in following parapodia (Fig. 4C–E).

Proximal region of dorsal ligule even throughout body; shorter than distal region of dorsal ligule in anteriormost and anterior parapodia (Fig. 4A, B), subequal to that in following parapodia (Fig. 4C–E); one glandular patch sub-oval in anterior parapodia (Fig. 4B), becoming distinctly smaller towards posterior end.

Distal region of dorsal ligule extending markedly beyond end of notoacicular throughout (Fig. 4B–E); thick, bluntly rounded and slightly longer than median ligule in anterior parapodia (Fig. 4B), longer than median ligule in following parapodia (Fig. 4C–E), bluntly triangular in middle parapodia (Fig. 4C), becoming digitiform towards posterior end (Fig. 4D, E); one massive glandular patch covering entirely distal dorsal ligule of anterior parapodia (Fig. 4B), much compact in middle and posterior parapodia.

Notopodial prechaetal lobe blunt, thick and short

(Fig. 4B) in anterior parapodia, becoming narrower and reduced to notoacicular papilla from parapodia 14 (Fig. 4C–E).

Median ligule thick, bluntly rounded in anterior parapodia (Fig. 4B), bluntly triangular in middle parapodia (Fig. 4C), becoming digitiform towards posterior end (Fig. 4D, E).

Neuropodia consisting of neuroacicular ligule with inferior lobe, ventral ligule, and ventral cirrus (Fig. 4A–E); superior and postchaetal lobes absent throughout.

Neuroacicular ligule shorter than ventral ligule in anteriormost parapodia (Fig. 4A), distinctly longer than that in anterior and middle parapodia (Fig. 4B, C), becoming subequal to or slightly shorter than that towards posterior end (Fig. 4D, E); 1.5–1.7 times as wide as ventral ligule in anteriormost and anterior parapodia, as wide as that in following parapodia.

Inferior lobe as small, blunt process in anteriormost and anterior parapodia, becoming narrower and reduced from parapodia 18.

Ventral ligule well-developed throughout, thick; digitiform and shorter than median ligule throughout, except bluntly rounded in anterior parapodia (Fig. 4B).

Ventral cirri digitiform and thick in anterior-most parapodia (Fig. 4A), becoming conical in following ones; as long as two-thirds of ventral ligule throughout.

Aciculae black, with basal end uncoloured. Notoacicular absent in first 2 parapodia (Fig. 4A). Neuroacicular extending beyond distal end of notoacicular throughout (Fig. 4B–E). Neuroacicular longer than median ligule in anterior parapodia (Fig. 4B), distinctly shorter than that in following parapodia (Fig. 4C–E).

Notochaetae all homogomph spinigers throughout; 15–16 spinigers present in anterior parapodia, 5–7 spinigers in following parapodia.

Upper neurochaetae consisting of homogomph spinigers and heterogomph falcigers, both present throughout; 8 spinigers present in anteriormost parapodia, 12 spinigers in anterior parapodia, 3–4 spinigers in following parapodia; 3–4 falcigers present throughout.

Lower neurochaetae consisting of heterogomph spinigers and heterogomph falcigers, both present throughout; 1–2 spinigers present in all parapodia; 24–26 falcigers in anteriormost and anterior parapodia, 5–6 falcigers in middle parapodia, 4–5 falcigers in posterior parapodia.

Blade of both homogomph (Fig. 4F) and heterogomph spinigers (Fig. 4G) finely serrated towards toothed edge, evenly spaced. Blade of heterogomph falcigers short (b/a ratio 0.75–1.1), convex, terminal tooth blunt with inconspicuous tendon, serrations present in about one-third to two-fifths (0.31–0.4) of total blade length (Fig. 4H–K); supracicular blades of similar size (Fig. 4H, J), subacicular ones becoming slightly shorter and thicker downwards in parapodia (Fig. 4I, K). Shaft of falcigers cambered, with cavity divided sub-distally into four longitudinal partitions (Fig. 4J, K).

Pygidium well developed, as long as last 3 chaetigers; anal cirri thick, short, equalling length of last 6 chaetigers, with small cirrophores (Fig. 3K).

Variation. Total length of complete specimens: 35–42

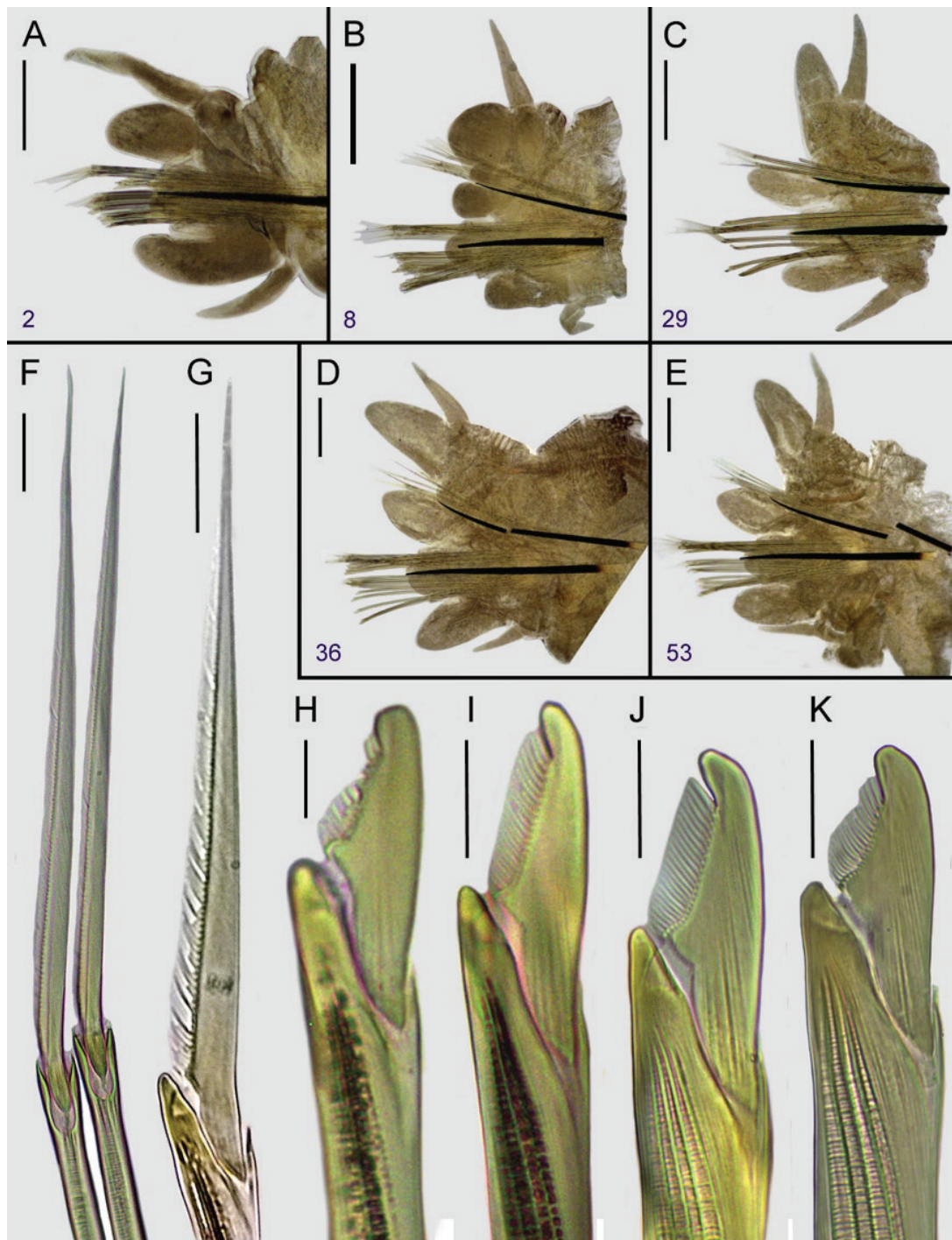


Fig. 4: *Perinereis louizomarum* n. sp. (A–E) Holotype (MNHN N IA type N: 2065) from El-Kala, Algeria, atokous: (A) parapodium of chaetiger 2; (B) parapodium of chaetiger 8; (C) parapodium of chaetiger 29; (D) parapodium of chaetiger 36; (E) parapodium of chaetiger 53; (F) homogomph spinigers from notochaetae (chaetiger 53); (G) heterogomph spiniger from subacicular neurochaetae (chaetiger 53); (H) heterogomph falciger from supracicular neurochaetae (chaetiger 2); (I) heterogomph falciger from subacicular neurochaetae (chaetiger 2); (J) heterogomph falciger from subacicular neurochaetae (chaetiger 8); (K) heterogomph falciger from subacicular neurochaetae (chaetiger 53). Scale bars: A: 0.5 mm; B: 0.6 mm; C, D: 0.4mm; E: 0.3 mm; F: 30 µm; G: 40 µm; H: 10 µm; I–K: 20µm.

mm; L15: 9–12 mm, W15: 4–5 mm, 37–62 chaetigers. Anterodorsal tentacular cirri extending back to chaetiger 3–4. Posterodorsal tentacular cirri extending back to chaetiger 4–6. Jaws smooth, sometimes with 1–2 (rarely 3) thick and blunt denticles. Number and arrangement of paragnaths: area I with 1–2; area II with 3–8; area III with 1–5; area IV with 6–19; area V without paragnaths, rarely

1–2; area VI with crescent-shaped bar, sometimes with concave base and pointed tip; area VII–VIII with 26–38, in two bands, anterior row with p-bars present, less distinct in coarser paragnaths, posterior row with cones on ridge rows only, albeit occasionally seem also present on furrow rows. Anal cirri as long as last 4–5 chaetigers.

Anatomical anomalies. One specimen (62 mm long,

3 mm wide, 62 chaetigers) with (1) three antennae of similar size, with the middle one thinner than laterals and bearing ceratophore (Fig. 2L); (2) two palpostyles on right palpophore (Fig. 3M); and (3) three tentacular cirri in the right side (Fig. 3M). Additionally, another specimen was found with area VII without paragnaths.

Habitat. Rocky shores covered by algae at 0.5 m depth, including *Corallina* sp., *Dictyopteris* sp., *Ulva lactuca* Linnaeus, and *Colpomenia sinuosa* (Mertens ex Roth) Derbès & Solier, and also found among the bivalve *Mytilus galloprovincialis* (Lamarck, 1819).

Etymology. The species' name combines the first author's parents' names, Louiza and Omar, to honor their devotion, love, and the warm life they made for her. The name is a noun in the genitive (ICZN, 1999).

Type locality. Annaba, Algeria, Mediterranean Sea.

Taxonomic remarks. *Perinereis louizomarum* n. sp. is a member of the *Perinereis* species SG-1A. Among the other 13 species within this subgroup (after Hutchings *et al.*, 1991), *P. louizomarum* n. sp. is similar to *P. cultrifera* from the Tyrrhenian Sea (Grube, 1840; Hutchings *et al.*, 1991; Park & Kim, 2017), *P. floridana* from the Gulf of Mexico (Ehlers, 1868; de León-González & Solís-Weiss, 1998) and *P. rullieri* from the Mediterranean Sea (Pilato, 1974) by having two well-distinguished bands of paragnaths on the areas VII–VIII, dorsal cirri of similar length to or slightly longer than distal dorsal ligule in anterior parapodia, and neuropodia without postchaetal lobe. Nevertheless, *P. louizomarum* n. sp. is distinguishable from these three species by the following features: (I) the lower number of denticles on jaws (normally 0, occasionally 2 or 3), in contrast to the higher number in *P. cultrifera* (4), *P. floridana* (9) and *P. rullieri* (4–5); (II) the absence of laterally isolated paragnaths on area III, in comparison to their presence in *P. cultrifera* and *P. rullieri* (rarely absent); (III) the lower number of paragnaths on areas III (1–5) and V (typically 0, rarely 1 or 2), in contrast to the higher number in *P. cultrifera* (area III: 5–12; V: 3, rarely 2, 4 or 5), *P. floridana* (area III: 16; V: 1, rarely 2) and *P. rullieri* (area III: 5–9, rarely higher; V: 3, rarely fewer or higher); (IV) the bluntly rounded distal dorsal ligule in anterior parapodia, in comparison to that conical or bluntly conical in *P. cultrifera*, *P. floridana* and *P. rullieri*; (V) the dorsal cirri located medially on dorsal ligule in posterior parapodia, in comparison to subdistally placed in *P. cultrifera* and *P. rullieri*; and (VI) the coarse paragnaths on areas VII–VIII, in comparison to those small and delicate in *P. cultrifera* and *P. rullieri*. Additionally, *P. louizomarum* n. sp. is different from *P. floridana* by having notopodial prechaetal lobes, at least in anterior parapodia, and neuropodia without superior lobe throughout the body, in contrast to their absence and presence in *P. floridana*, respectively.

Based on the biogeographic overlap, *P. tenuisetis* described from the Mediterranean Sea also warrants comparison with the new species. *Perinereis louizomarum* n. sp. can readably be distinguished from *P. tenuisetis* by the following diagnostic characteristics: in *P. louizomarum* n. sp., the aciculae are black, the dorsal cirri are placed medially on dorsal ligule in posterior parapodia, and neu-

rochaetae have heterogomph spinigers and falcigers; in contrast, *P. tenuisetis* has yellow light aciculae, dorsal cirri placed basally on dorsal ligule in the same parapodia, and subacicular neurochaetae with homogomph spinigers and falcigers.

Key to species of *Perinereis* Kinberg, 1865 belonging to the Subgroup 1A

This key includes all species now regarded as *Perinereis* Subgroup 1A *sensu* Hutchings *et al.* (1991). *Perinereis dongalae* originally described from Celebes, Indonesia by Horst (1924), is currently known only by the brief description and scarcely illustrated epitokous stage. Hylleberg *et al.* (1986) synonymized the species with *P. striolata* (Grube, 1878) without examination of the type material.

1. Area I with 7 or more paragnaths 2
- Area I with up to 4 paragnaths 3
2. Area I with 16 paragnaths; area V with *P. arabica* More than 8 paragnaths in two transverse hammas, 1971 rows (Kuwait)
- Area I with up to 8 paragnaths; area V with one paragnath *P. dongalae* (Horst, 1924) (Celebes, Indonesia)
3. Aciculae light yellow; dorsal cirri placed basally on dorsal ligule in posterior parapodia; neurochaetae with homogomph falcigers; subacicular neurochaetae with homogomph spinigers *P. tenuisetis* (Fauvel, 1915) (Sicily, Italy)
- Aciculae dark brown or black; dorsal cirri placed medially or subdistally on dorsal ligule in posterior parapodia; neurochaetae with heterogomph falcigers; subacicular neurochaetae with heterogomph spinigers 4
4. Areas VII–VIII with anterior band only, up to 13 paragnaths 5
- Areas VII–VIII divided into two well-separated bands (anterior and posterior), with more than 20 paragnaths 6
5. Ridges of area VI distally and sub-medially coalesced (areas VI–V–VI ridge pattern λ-shaped); area III without laterally isolated paragnaths; area V without paragnaths; areas VII–VIII with 10 or more paragnaths *P. calmani* (Monro, 1926) (Australia and Macclesfield Bank)

— Ridges of area VI distally separated from each other (areas VI–V–VI ridge pattern π -shaped); area III with laterally isolated paragnaths; area V with 1–3 paragnaths; areas VII–VIII with up to 6 paragnaths	<i>P. taorica</i> Langerhans, 1881 (Madeira)	13. Distal dorsal ligule bluntly conical in anterior parapodia; dorsal cirri located subdistally on dorsal ligule in posterior parapodia; jaws with 4–5 denticles; area III with up to 5–12 paragnaths, with laterally isolated cones; areas VII–VIII with small and delicate paragnaths	<i>P. cultrifera</i> (Grube, 1840) (Italy, Tyrrhenian Sea)
6. Area V without paragnaths	7		
— Area V with paragnaths	8	— Distal dorsal ligule bluntly rounded in anterior parapodia; dorsal cirri located medially on dorsal ligule in posterior parapodia; jaws with up to 3 denticles; area III with up to 5 paragnaths, without laterally isolated cones; areas VII–VIII with coarse paragnaths	<i>P. louizomarum</i> n. sp. (Algeria, Mediterranean Sea) <i>partim</i>
7. Neuropodial postchaetal lobe present throughout; dorsal cirri extending markedly beyond end of distal dorsal ligule (thrice ligule length) in posterior parapodia	<i>P. capensis</i> (Kinberg, 1865) (South Africa)		
— Neuropodial postchaetal lobe absent throughout; dorsal cirri extending barely beyond end of distal dorsal ligule in posterior parapodia	<i>P. louizomarum</i> n. sp. (Algeria, Mediterranean Sea) <i>partim</i>		
8. Dorsal cirri extending markedly beyond end of distal dorsal ligule (more than twice ligule length) in posterior parapodia	9		
— Dorsal cirri extending barely beyond end of distal dorsal ligule in posterior parapodia	12		
9. Area V with one paragnath	10		
— Area V with 3 or more paragnaths (rarely 2)	11		
10. Proximal dorsal ligule not enlarged in posterior parapodia; heterogomph falcigers with serrations present in about half of total blade length	<i>P. falsovariegata</i> Monro, 1933 (South Africa)		
— Proximal dorsal ligule enlarged in posterior parapodia; heterogomph falcigers with serrations present in about one-sixth of total blade length	<i>P. villalobosi</i> Rioja, 1947 (Gulf of California, Mexico)		
11. Areas VII–VIII with anterior band having two rows of paragnaths (one on furrows and one on ridges); area III with up to 9 paragnaths	<i>P. rullieri</i> Pilato, 1974 (Sicily, Italy)		
— Areas VII–VIII with anterior band having only furrow row of paragnaths; area III with 12–26 paragnaths	<i>P. helleri</i> (Grube, 1878) (Philippines)		
12. Notopodial prechaetal lobe absent throughout; jaws with 9 denticles; area III with 16 paragnaths	<i>P. floridana</i> (Ehlers, 1868) (Gulf of Mexico, USA)		
— Notopodial prechaetal lobe present in anterior parapodia; jaws with up to 5 denticles; area III with up to 12 paragnaths	13		

Discussion

Perinereis cultrifera was first described from Italy and has since been documented along the coasts of several regions (Fauvel, 1923; Day, 1967; Wu *et al.*, 1985). The species' global distribution has been questioned several times, most recently by Park & Kim (2017). A thorough redescription is still required to delimit its morphology; however, the original description (Grube, 1840) and a few comments and illustrations provided on the type material (Park & Kim, 2017) provide primary features that can be used to evaluate specimens from other localities. This was true for the Algerian specimens examined here. *P. louizomarum* n. sp. is discovered after a thorough comparison with the relevant literature.

Gravier & Dantan (1928) indicated that the reproduction of *P. cultrifera* was mainly of an atokous type in the Mediterranean Sea by the very few epitokous specimens found during their samplings. However, Durchon (1955, 1957, 1965) proposed that these different modes of reproduction, heteronereid and atokal, represent two distinct groups of *P. cultrifera* on the shores of the North Atlantic and the Mediterranean Sea. For instance, the species in the English Channel developed a markedly epitokal transformation, whereas in the Mediterranean, particularly in Algeria, the reproduction was carried out in the atokous state.

Later, the observations made by Scaps, Rouabah, and other colleagues (Scaps *et al.*, 2000; Rouabah & Scaps, 2003a, b; Rouabah & Rouabah, 2007) in further localities from the littorals of France, United Kingdom, Algeria and Tunis, confirmed that a complex of species might be referred to *P. cultrifera* not only due to the mode of reproduction but also to differences in the spawning season, age at maturity, morphology and biochemical characteristics. The authors concluded that three different populations could be found: (I) the epitokous form of the English Channel and Atlantic, characterized by having an elevated and a higher number of chaetigers (up to 120); (II) the atokous form of Algeria, in which the number segment is below 80; and (III) the epitokous form from Tunis with biometric features between the two above-described forms. *Perinereis louizomarum* n. sp. belongs to the atokous form of '*P. cultrifera*' (Durchon, 1957) since

no traces of epitokal modifications were found in our specimens, including a mature female, and the number of segments was below 70. Nevertheless, further surveys to collect the species should be carried out throughout the year to obtain individuals in different life stages since specimens representing both forms of reproduction have been documented to coexist in Algeria (Gravier & Dantan, 1928; Durchon, 1957).

Complementing morphological taxonomy with the molecular approach permits the direct comparison of species among a global reference library using fragments of specimens at any stage of the life cycle, therefore furnishing a universal master tool for the identification and discovery of cryptic forms (Costa & Carvalho, 2007). Several studies have examined variation in mitochondrial DNA sequences in a region of the COI gene as a dependable approach to distinguish polychaete species (e.g., Glover *et al.*, 2005; Bleidorn *et al.*, 2006; Rice *et al.*, 2008; Pleijel *et al.*, 2009; Barroso *et al.*, 2010; Nygren & Pleijel, 2011). For instance, species regarded as cosmopolitan were detected as species complexes of several cryptic species, such as the amphinomid *Eurythoe complanata* (Pallas, 1766) (Barroso *et al.*, 2010) or the phyllodocid *Eumida sanguinea* (Örsted, 1843), the latter comprising about ten species (Nygren & Pleijel, 2011).

In the present study, besides the morphological, reproductive, and biochemical differences settled in previous studies, *P. cultrifera* is confirmed as a species complex in the European seas but based on DNA sequence data. The mtCOI sequences available in GenBank for four specimens, called '*P. cultrifera*' from the South European Atlantic shelf (northern Portugal and northwest France) (Table 3), were compared with those retrieved here from the Western Mediterranean (Algeria). Interestingly, those sequences of *P. cultrifera* from the South European Atlantic shelf resulted in two distinct clades (Fig. 2), which suggests that two populations of '*P. cultrifera*' are present in the region. Marked genetic differences between these two populations with those specimens from Algeria were also found (Table 5). In addition to *P. cultrifera*, the sequences of the remaining six species also show distinct pairwise distances, which are relatively comparable with those documented to distinguish *Perinereis* species (Park & Kim, 2017; Villalobos-Guerrero *et al.*, 2021), supporting the establishment of *P. louizomarum* n. sp. as a new species. Based on the mtCOI sequences of the South European Atlantic populations, one of them represents the "*P. cultrifera*" Type I indicated by Scaps *et al.* (2000) and Rouabah & Rouabah (2007), and at least one likely represents an undescribed species. However, the material of both populations needs to be examined to understand and delimit their morphology, including the reproductive patterns of epitokal characters, which are considered relevant in nereidids to distinguish among closely related species (Read, 2007; Conde-Vela & Salazar-Vallejo 2015; Villalobos-Guerrero & Bakken 2018; Villalobos-Guerrero & Idris 2021).

On the other hand, Lobo *et al.* (2016) reported that polychaetes are suitable candidates for surveilling benthic ecosystems. An innovative and alternative approach

to develop a rapid evaluation of the ecological status of marine ecosystems is through genomic observations (Hajibabaei *et al.*, 2011; Avó *et al.*, 2017; Aylagas *et al.*, 2018). Studies on the molecular characterization of polychaete fauna along the Algerian waters are limited. Expanding the genetic studies of polychaetes in Algeria is recommended to see if environmental parameters impact them. Aylagas *et al.* (2014) suggested that the most abundant species encountered during monitoring programs should be sequenced with accurate taxonomic marking to assess ecological status reliably. Vijapure *et al.* (2019) reported that the polychaete DNA libraries could be expanded with sequences commonly found during ecological monitoring surveys.

Marine environments exposed to many anthropogenic pressures show a gradual and continuous loss of diversity (Halpern *et al.*, 2008). Erpenbeck *et al.* (2016) proposed regular monitoring of the ecological quality of marine and estuarine waters. It is often achieved by establishing species inventories that can be used as a benchmark against which future disturbances of diversity loss can be measured. Therefore, appropriate management strategies can be applied in areas where species richness is declining or where the loss of diversity is expected to be imminent (Thomsen & Willerslev, 2015). Aylagas *et al.* (2018), Lejzerowicz *et al.* (2015), and Bevilacqua & Terlizzi (2016), reported that biomonitoring mainly relies on morphological taxonomy, which is often criticized as labor-intensive, time-consuming and costly. To avoid these problems, Bik *et al.* (2012) and Sigamani *et al.* (2016) have proposed the integration of genetics into biological assessment methods in addition to classical taxonomy.

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References

- Alves, P.R., Halanych, K.M., Santos, C.S.G., 2020. The phylogeny of Nereididae (Annelida) based on mitochondrial genomes. *Zoologica Scripta*, 00, 1-13.
- Arias, A., Richter, A., Anadón, N., Glasby, C.J., 2013. Revealing polychaetes invasion patterns: Identification, reproduction and potential risks of the Korean ragworm, *Perinereis lineata* (Treadwell), in the Western Mediterranean. *Estuarine, Coastal and Shelf Science*, 131, 117-128.
- Audouin, J.V., Milne-Edwards, H., 1833. Classification des Annelides, et description de celles qui habitent les cotes de la France. *Annales des Sciences Naturelles, Paris, Serie 1*, 29, 195-269.
- Avó, A.P., Daniell, T.J., Neilson, R., Oliveira, S., Branco, J. et al., 2017. DNA barcoding and morphological identification of benthic nematodes assemblages of estuarine intertidal sediments: advances in molecular tools for biodiversity assessment. *Frontiers in Marine Science*, 4, 66.
- Aylagas, E., Borja, A., Rodríguez-Espeleta, N., 2014. Environmental status assessment using DNA metabarcoding: towards a genetics based marine biotic index (gAMBI). *PloS one*, 9 (3), e90529.
- Aylagas, E., Borja, A., Muxika, I., Rodríguez-Espeleta, N., 2018. Adapting metabarcoding-based benthic biomonitoring into routine marine ecological status assessment networks. *Ecological Indicators*, 95, 194-202.
- Bakalem, A., Gillet, P., Pezy, J.P., Dauvin, J.C., 2020. Inventory and the biogeographical affinities of Annelida Polychaeta in the Algerian coastline (Western Mediterranean). *Mediterranean Marine Science*, 21 (1), 157-182.
- Barroso, R., Klautau, M., Solé-Cava, A.M. Paiva, P.C., 2010. *Eurythoe complanata* (Polychaeta: Amphinomididae), the 'cosmopolitan' fireworm, consists of at least three cryptic species. *Marine Biology*, 157, 69-80.
- Bevilacqua, S., Terlizzi, A., 2016. Species surrogacy in environmental impact assessment and monitoring: extending the BestAgg approach to asymmetrical designs. *Marine Ecology Progress Series*, 547, 19-32.
- Bik, H.M., Porazinska, D.L., Creer, S., Caporaso, J.G., Knight, R. et al., 2012. Sequencing our way towards understanding global eukaryotic biodiversity. *Trends in ecology & evolution*, 27 (4), 233-243.
- Blainville, H.M.D de., 1818. *Mémoire sur la classe des Sétipodes, partie des Vers à sang rouge de M. Cuvier, et des Annélides de M. de Lamarck*. Bulletin des Sciences, par la Société Philomatique, Paris, 85 pp.
- Bleidorn, C., Kruse, I., Albrecht, S., Bartolomaeus, T., 2006. Mitochondrial sequence data expose the putative cosmopolitan polychaete *Scoloplos armiger* (Annelida, Orbiniidae) as a species complex. *BMC Evolutionary Biology*, 6 (1), 1-13.
- Carr, C.M., Hardy, S.M., Brown, T.M., Macdonald, T.A., Herbert, P.D.N., 2011. A Tri-Oceanic Perspective: DNA Barcoding Reveals Geographic Structure and Cryptic Diversity in Canadian Polychaetes. *PLoS ONE*, 6 (7), e22232.
- Claparède, E., 1870. Les Annélides Chétopodes du Golfe de Naples. Supplément. *Mémoires de la Société de physique et d'histoire naturelle de Genève*, 20 (2), 365-542.
- Conde-Vela, V.M., Salazar-Vallejo, S.I., 2015. Redescription of *Nereis oligohalina* (Rioja, 1946) and *N. garwoodi* González-Escalante & Salazar-Vallejo, 2003 and description of *N. confusa* sp. n. (Annelida, Nereididae). *ZooKeys*, (518), 15.
- Costa, F.O., Carvalho, G.R., 2007. The Barcode of Life Initiative: synopsis and prospective societal impacts of DNA barcoding of fish. *Genomics, Society and Policy*, 3 (2), 29-40.
- Day, J.H., 1967. A monograph on the Polychaeta of Southern Africa, Part 1. Errantia. *Trustees of the British Museum (Natural History)*, 656, 1-458.
- Díaz-Jaramillo, M., Ferreira, J.L., Amado, L.L., Ventura-Lima, J., Martins, A. et al., 2010. Biomonitoring of antioxidant and oxidative stress responses in *Perinereis gualpensis* (Polychaeta: Nereididae) in Chilean estuarine regions under different anthropogenic pressure. *Ecotoxicology and Environmental Safety*, 73, 515-523.
- Drennan, R., Wiklund, H., Rabone, M., Georgieva, M.N., Dahlgren, T.G. et al., 2021. *Neanthes goodayi* sp. nov. (Annelida, Nereididae), a remarkable new annelid species living inside deep-sea polymetallic nodules. *European Journal of Taxonomy*, 760 (1), 160-185.
- Durchon, M., 1955. Sur le polymorphisme présenté par quelques Néréidiens (Annélides Polychètes) au moment de la reproduction. *Bulletin de la Société d'histoire naturelle d'Afrique du Nord*, 46, 180-194.
- Durchon, M., 1957. Problèmes posés par le comportement des néréidiens au moment de leur reproduction. *Annals of Biology*, 33, 31-42.
- Durchon, M., 1965. Sur l'évolution phylogénétique et ontogénétique de l'épitoque chez les Néréidiens (Annélides Polychètes). *Zoologische Jahrbücher*, 1-12.
- Ehlers, E.H., 1868. *Die Borstenwürmer (Annelida Chaetopoda) nach systematischen und anatomischen Untersuchungen dargestellt*. Wilhelm Engelmann, Leipzig, 748 pp.
- Ehlers, E.H., 1904. Neuseelandische Anneliden. Abhandlungen der Königlichen Gesellschaft der Wissenschaften zu Göttingen. Mathematisch-Physikalische Klasse. *Neue Folge*, 3 (1), 1-80.
- Erpenbeck, D., Voigt, O., Al-Aidaros, A.M., Berumen, M.L., Büttner, G. et al., 2016. Molecular biodiversity of Red Sea demosponges. *Marine Pollution Bulletin*, 105 (2), 507-514.
- Fauvel, P., 1915. Polychètes pélagiques nouvelles des Campagnes de la Princesse-Alice. *Bulletin de l'Institut Océanographique*, 305, 1-11.
- Fauvel, P., 1923. *Polychètes errantes. Faune de France*, 5. Paul Lechevalier, Paris, 488 pp.
- Gasmi, H., Maamcha, O., Daas, T. Scaps, P., 2016. First record of *Perinereis macropus* and *Perinereis cultrifera* (Annelida, Polychaeta) from rocky shores east Algeria, (SW Mediterranean sea). *Journal of Entomology and Zoology Studies*, 4 (4), 160-166.
- Gillet, P., 2017. Liste des annélides polychètes du Maroc. *Bulletin de l'Institut Scientifique, Rabat, Section Science de la Vie*, 39, 19-26.
- Glasby, C.J., Hsieh, H.L., 2006. New species and new records of the *Perinereis nuntia* species group (Nereididae: Polychaeta) from Taiwan and other Indo-West Pacific shores. *Zoological Studies*, 45 (4), 553-577.
- Glasby, C.J., Timm, T., Muir, A.I., Gil, J., 2009. Catalogue of non-marine Polychaeta (Annelida) of the World. *Zootaxa*,

2070, 1-52.

- Glasby, C.J., Wei, N.W.V., Gibb, K.S., 2013. Cryptic species of Nereididae (Annelida: Polychaeta) on Australian coral reefs. *Invertebrate Systematics*, 27 (3), 245-264.
- Glover, A.G., Källström, B., Smith, C.R., Dahlgren, T.G., 2005. World-wide whale worms? A new species of *Osedax* from the shallow north Atlantic. *Proceedings of the Royal Society B: Biological Sciences*, 272 (1581), 2587-2592.
- Gravier, C., Dantan, J.L., 1928. Sur quelques points de la biologie des annélides Polychètes de la famille des néréidiens. *Compte Rendu Académie des Sciences*, Paris, 186, 1476.
- Grube, A.E., 1840. *Actinien, Echinodermen und Wurmen des Adriatischen und Mittelmeers nach Eigenen Sammlungen Beschrieben*. J.H. Bon, Königsberg 92 pp.
- Grube, A.E., 1858. *Annulata Örstediana: Enumeratio Annulatum quae in itinere der Indiam occidentalem et Americam centralem annis 1845-1848 suscept legit cl. A. S. Oersted, adjectis speciebus nonnullis a cl. H. Kröyer in itinere ad Americam meridionalem collectis. Videnskabelige Meddelelser fra Dansk Naturhistorisk Forening i Kjöbenhavn*, Nr. 1-4, 105-120.
- Grube, A.E., 1860. Beschreibung neuer oder wenig bekannter Anneliden. Fünfter Beitrag. *Archiv für Naturgeschichte*, Berlin, 26 (1), 71-118, plates III-V.
- Grube, A.E., 1866. Beschreibungen neuer von der Novara-Expedition mitgebrachter Anneliden und einer neuen Landplanarie. *Verhandlungen der kaiserlich-königlichen zoologisch-botanischen Gesellschaft in Wien*, 16, 173-184.
- Grube, A.E., 1878. *Annulata Semperiana. Beiträge zur Kenntnis der Annelidenfauna der Philippinen nach den von Herrn Prof. Semper mitgebrachten Sammlungen*. Mémoires de l'Académie Impériale des Sciences de St.-Petersbourg, Russia, 300 pp.
- Hajibabaei, M., Shokralla, S., Zhou, X., Singer, G.A., Baird, D.J., 2011. Environmental barcoding: a next-generation sequencing approach for biomonitoring applications using river benthos. *PLoS ONE*, 6 (4), e17497.
- Hall, T.A., 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic acids symposium series*, 4, 95-98.
- Halpern, B.S., Walbridge, S., Selkoe, K.A., Kappel, C.V., Micheli, F. et al., 2008. A global map of human impact on marine ecosystems. *Science*, 319 (5865), 948-952.
- Hartman, O., 1949. The Marine Annelids erected by Kinberg with notes on some others types in the Swedish State Museum. *Arkives för Zoologi*, 42 (1), 1-137.
- Hartman, O., 1959. Catalogue of the Polychaetous Annelids of the World. Parts 1 & 2. *Occasional Papers of the Allan Hancock Foundation*, 23, 1-628.
- Horst, R., 1889. Contributions towards the knowledge of the Annelida Polychaeta. *Notes from the Leyden Museum*, 11, 161-186, plates 7-8.
- Horst, R., 1924. Polychaeta errantia of the Siboga-Expedition. Part III. Nereidae and Hesionidae. *Siboga-Expedition Leyden 99*, Monograph 24, 145-198.
- Hutchings, P.A., Reid, A., Wilson, R.S., 1991. *Perinereis* (Polychaeta, Nereididae) from Australia, with redescrptions of six additional species. *Records of the Australian Museum*, 43, 241-274.
- Hylleberg, J., Nateewathana, A., Bussarawit, S., 1986. Polychaetes of Thailand. Nereidae (Part 1); *Perinereis* and *Pseudonereis* with notes on species of commercial value. *Phuket Marine Biological Center Research Bulletin*, 43, 1-22.
- ICZN 1999. *International Code of Zoological Nomenclature*. The International Trust for Zoological Nomenclature, Fourth Edition, London, UK, 306 pp.
- Jeldes, F., 1963. *Un nuevo nereido de agua dulce para Chile. Gayana, Zoologia*, 9, 4-10.
- Kara, J., Macdonald, A.H., Simon, C.A., 2018. Integrative taxonomic methods reveal an incorrect synonymisation of the South African *Pseudonereis podocirra* (Schmarda) as the widespread *Pseudonereis variegata* (Grube) from Chile. *Invertebrate Systematics*, 32 (6), 1282-1297.
- Kara, J., Santos, C.S., Macdonald, A.H., Simon, C.A., 2020. Resolving the taxonomic identities and genetic structure of two cryptic *Platynereis* Kinberg species from South Africa. *Invertebrate Systematics*, 34 (6), 618-636.
- Kim, H., Jung, G., Lee, Y.C., Pae, S.J., Kim, C.G. et al., 2015. The complete mitochondrial genome of the marine polychaete: *Perinereis aibuhitensis* (Phyllodocida, Nereididae). *Mitochondrial DNA*, 26 (6), 869-870.
- Kimura, M., 1980. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *Journal of molecular evolution*, 16, 111-120.
- Kinberg, J.G.H., 1865. *Annulata nova. Nereidum dispositio nova. Öfversigt af Kongelige Vetenskaps-Akademiens Förhandlingar*, 22, 167-179.
- Kumar, S., Stecher, G., Li, M., Knyaz, C., Tamura, K., 2018. MEGA X: molecular evolutionary genetics analysis across computing platforms. *Molecular biology and evolution*, 35 (6), 1547.
- Lamarck, J.B.M., 1818. *Histoire naturelle des Animaux sans Vertèbres, présentant les caractères généraux et particuliers de ces animaux, leur distribution, leurs classes, leurs familles, leurs genres, et la citation des principales espèces qui s'y rapportent; precedes d'une Introduction offrant la détermination des caracteres essentiels de l'Animal, sa distinction du vegetal et desautres corps naturels, enfin, l'Exposition des Principes fondamentaux de la Zoologie*. Deterville, Paris, 612 pp.
- Lamarck, J.B.M., 1819. *Histoire naturelle des animaux sans vertèbres*. Verdière, Paris, 343 pp.
- Langerhans, P., 1881. *Ueber einige canarische Anneliden*. Nova Acta der Ksl Leopold-Carol.-Deutschen Akademie der Naturforscher, Halle, 142 pp.
- Lee, J.H., Je, J.G., Choi, J.W., 1992. Taxonomical review of *Perinereis aibuhitensis* Grube, 1878 (Nereidae: Polychaeta) in Korea. *The Korean Journal of Zoology*, 8, 1-10.
- Lejzerowicz, F., Esling, P., Pillet, L., Wilding, T.A., Black, K.D. et al., 2015. High-throughput sequencing and morphology perform equally well for benthic monitoring of marine ecosystems. *Scientific reports*, 5 (1), 1-10.
- León González, J.A. de, Solís-Weiss, V., 1998. The genus *Perinereis* (Polychaeta: Nereididae) from Mexican littoral waters, including the description of three new species and the redescrptions of *P. anderssoni* and *P. elenacasoae*. *Proceedings of the Biological Society of Washington*, 111, 674-693.
- Linnaeus, C., 1758. *Systema Naturae per regna tria naturae*,

- secundum classes, ordines, genera, species, cum characteribus, differentiis, synonymis, locis*. Editio decima, reformata (10th revised edition), 824 pp.
- Lobo, J., Costa, P.M., Teixeira, M.A., Ferreira, M.S., Costa, M.H. *et al.*, 2013. Enhanced primers for amplification of DNA barcodes from a broad range of marine metazoans. *BMC Ecology*, 13 (1), 34.
- Lobo, J., Teixeira, M.A., Borges, L.M., Ferreira, M.S., Hollatz, C. *et al.*, 2016. Starting a DNA barcode reference library for shallow water polychaetes from the southern European Atlantic coast. *Molecular Ecology Resources*, 16 (1), 298-313.
- Meghlaoui, Z., Daas, T., Snani, M., Daas-Maamcha, O., Scaps, P., 2015. Annual variations of the presence of Nereididae (Annelida: Polychaeta) from intertidal rocky shores along the east coast of Algeria. *Check List*, 11 (6), 1808-1808.
- Mohammad, M.B.M., 1971. Intertidal polychaetes from Kuwait, Arabian Gulf, with descriptions of three new species. *Zoological Journal of the Linnean Society*, 163, 285-303.
- Monro, C.C.A., 1926. Polychaeta of the 'Alert' Expedition. Families Hesionidae and Nereidae. *Zoological Journal of the Linnean Society*, 36, 311-323.
- Monro, C.C.A., 1933. The Polychaeta Errantia collected by Dr. Crossland at Colón, in the Panama Region, and the Galapagos Islands during the expedition of the S.Y. "St. George." *Proceedings of the Zoological Society of London*, 103 (1), 1-96.
- Nygren, A., Pleijel, F., 2010. Redescription of *Imajimaea draculai*—a rare syllid polychaete associated with the sea pen *Funiculina quadrangularis*. *Journal of the Marine Biological Association of the United Kingdom*, 90 (7), 1441-1448.
- Nygren, A., Pleijel, F., 2011. From one to ten in a single stroke—resolving the European *Eumida sanguinea* (Phyllodocidae, Annelida) species complex. *Molecular phylogenetics and evolution*, 58 (1), 132-141.
- Örsted, A.S., 1843. *Annulorum danicorum conspectus. Librariae Wahlianae, Hafniae*, 52 pp.
- Pallas, P.S., 1766. *Miscellanea zoologica. Quibus novae imprimis atque obscurae animalium species describuntur et observationibus iconibusque illustrantur*. Petrum van Cleef. Hagí Comitum., xii +, 224 pp.
- Park T.S., Kim, W., 2007. A taxonomic study on *Perinereis nuntia* species group (Polychaeta: Nereididae) of Korea. *Korean Journal of Systematic Zoology*, 23 (1), 75-85.
- Park, T., Kim, W., 2017. Description of a new species for Asian populations of the "Cosmopolitan" *Perinereis cultrifera* (Annelida: Nereididae). *Zoological Science*, 34, 252-260.
- Pilato, G., 1974. *Perinereis rullieri*, nuova specie di nereidi (Annelida, Polychaeta) delle coste Siciliane. *Animalia*, Catania, 1 (1-3), 25-37.
- Pleijel, F., Rouse, G., Nygren, A., 2009. Five colour morphs and three new species of *Gyptis* (Hesionidae, Annelida) under a jetty in Edithburgh, South Australia. *Zoologica Scripta*, 38 (1), 89-99.
- Prevedelli, D., Castelli, A., Abbiati, M., Zunarelli Vandini, R., 1990. Distribution and life cycle of *Perinereis rullieri* Pilato (Polychaeta, Nereididae), a Mediterranean Endemism. *Rapport Commission internationale de la Mer Méditerranée*, 32, 1.
- Read, G.B., 2007. Taxonomy of sympatric New Zealand species of *Platynereis*, with description of three new species additional to *P. australis* (Schmarda) (Annelida: Polychaeta: Nereididae). *Zootaxa*, 1558 (1), 1-28.
- Rezzag Mahcene, H., Denis, F., Meziane, T., Maamcha, O., Daas, T., 2022. General structure of Nereids of the East Coast of Algeria. *Aquaculture, Aquarium, Conservation & Legislation*, 15, 282-293.
- Rhee J.S, Won, E.J, Kim, R.O, Choi, B.S, Choi, I.Y. *et al.*, 2012. The polychaete, *Perinereis nuntia* ESTs and its use to uncover potential biomarker genes for molecular ecotoxicological studies. *Environmental Research*, 112, 48-57.
- Rice, S.A., Karl, S., Rice, K.A., 2008. The *Polydora cornuta* complex (Annelida: Polychaeta) contains populations that are reproductively isolated and genetically distinct. *Invertebrate Biology*, 127 (1), 45-64.
- Rioja, E., 1947. Estudios Anelido16gicos XIX. Observaciones sobre algunos nereidos de las costas de Mexico. *Annales Instituto de Biologia, Mexico*, 18, 527-535.
- Rouabah, A., 2003. *Comparaison au plan biochimiques et génétique de population de l'annélide polychète Perenereis cultrifera du littoral français et Algerien : précision des liens de parenté*. Ph.D. Thesis. University of Mentouri, Constantine, 48 pp.
- Rouabah, A., Rouabah, L., 2007. Biodiversité et complexe d'espèces *Perinereis cultrifera*: un exemple de spéciation. *Sciences & Technologie. C, Biotechnologies*, 25, 79-87.
- Rouabah, A., Rouabah, L., Tahar, A., Scaps, P., 2009. Variation in paragraph number of the different morphs of the polychaete *Perinereis cultrifera* in relation to geographical location and type of reproduction. *Academic Journal of Entomology*, 1 (2), 19-26.
- Rouabah, A., Scaps, P., 2003a. Two-dimensional electrophoresis analysis of proteins from epitokous forms of the polychaete *Perinereis cultrifera* from English Channel and the Mediterranean Sea. *Cahiers de Biologie Marine*, 44, 227-236.
- Rouabah, A., Scaps, P., 2003b. Life cycle and population dynamics of the polychaete *Perinereis cultrifera* from the Algerian Mediterranean Coast. *Marine Ecology*, 24 (2), 85-99.
- Rouhi, A., Sif, J., Gillet, P., Deutsch, B., 2008. Reproduction and population dynamics of *Perinereis cultrifera* (Polychaeta: Nereididae) of the Atlantic coast, El Jadida, Morocco. *Cahiers de Biologie Marine*, 49, 150-160.
- Sampertegui, S., Rozbaczylo Narvaez, N., 2013. Morphological and molecular characterization of *Perinereis gualpensis* (Polychaeta: Nereididae) and its phylogenetic relationships with other species of the genus off the Chilean coast, Southeast Pacific. *Cahiers de Biologie Marine*, 54, 27-40.
- Savigny, J.C., 1818. Annelida. pp. 1-612. In: *Histoire Naturelle des Animaux sans Vertèbres*. De Lamarck J.B. (Ed). Détreville, Paris.
- Scaps, P., Rouabah, A., leprêtre, A., 2000. Morphological and biochemical evidence that *Perinereis cultrifera* (Polychaeta: Nereididae) is a complex of species. *Journal of the Marine Biological Association of the United Kingdom*, 80, 735-736.
- Schmarda, L., 1861. Neue wirbellose Thiere beobachtet und gesammelt art einer Reise urn die Erde 1853-1857. 1. Neue Turbellarien, Rotatorien und Anneliden. *Wilhelm Engelmann, Leipzig*, 1 (2), 1-164.
- Sekar, V., Rajasekaran, R., Prasannakumar, C., Sankar, R., Srid-

- har, R. *et al.*, 2016. Morphological and COI sequence-based characterisation of marine polychaete species from Great Nicobar Island, India. pp. 99-111. In: *DNA Barcoding in Marine Perspectives*. Trivedi, S., Ansari, A., Ghosh, S., Rehman, H. (Eds). Springer, Cham.
- Sigamani, S., Perumal, M., Thivakaran, G.A., Thangavel, B., Kandasamy, K., 2016. DNA barcoding of macrofauna act as a tool for assessing marine ecosystem. *Marine Pollution Bulletin*, 152, 107891.
- Sujeewan, R., Hebert, P.A.U.L., 2007. BOLD: the barcode of life data system. *Molecular ecology notes*, 7 (3), 355-364.
- Teixeira, M.A., Langeneck, J., Vieira, P.E., Hernández, J.C., Sampieri, B.R. *et al.*, 2022. Reappraisal of the hyperdiverse *Platynereis dumerilii* (Annelida: Nereididae) species complex in the Northern Atlantic, with the description of two new species. *Invertebrate Systematics*, 36 (11), 1017-1061.
- Thomsen, P.F., Willerslev, E., 2015. Environmental DNA—An emerging tool in conservation for monitoring past and present biodiversity. *Biological conservation*, 183, 4–18.
- Tosuji, H., Bastrop, R., Götting, M., Park, T., Hong, J. S. *et al.*, 2019. Worldwide molecular phylogeny of common estuarine polychaetes of the genus *Hediste* (Annelida: Nereididae), with special reference to interspecific common haplotypes found in southern Japan. *Marine Biodiversity*, 49, 1385-1402.
- Treadwell, A.L., 1936. Polychaetous annelids from Amoy, China. *Proceedings of the United States National Museum*, 83 (2984), 261-279.
- Vijapure, T., Sukumaran, S., Manohar, C.S., 2019. Molecular characterization and phylogenetics of Indian polychaete fauna: scope for implementation in ecological monitoring. *Aquatic Ecology*, 53, 665-677.
- Villalobos-Guerrero, T.F., 2019. Redescription of two overlooked species of the *Perinereis nuntia* complex and morphological delimitation of *P. nuntia* (Savigny in Lamarck, 1818) from the Red Sea (Annelida, Nereididae). *Zoosystema*, 41 (1), 465-496.
- Villalobos-Guerrero, T.F., Carrera-Parra, L.F., 2015. Redescription of *Alitta succinea* (Leuckart, 1847) and reinstatement of *A. acutifolia* (Ehlers, 1901) n. comb. based upon morphological and molecular data (Polychaeta: Nereididae). *Zootaxa*, 3919 (1), 157-178.
- Villalobos-Guerrero, T.F., Bakken, T., 2018. Revision of the *Alitta virens* species complex (Annelida: Nereididae) from the North Pacific Ocean. *Zootaxa*, 4483 (2), 201-257.
- Villalobos-Guerrero, T.F., Idris, I., 2021. Reproductive morphology and redescription of some *Neanthes* Kinberg, 1865 (Annelida: Nereididae) species from the southeastern Asian seas, with comparative synoptic tables of accepted species. *The European Zoological Journal*, 88 (1), 556-594.
- Villalobos-Guerrero, T.F., Park, T., Idris, I., 2021. Review of some *Perinereis* Kinberg, 1865 (Annelida: Nereididae) species of the Group 2 *sensu* Hutchings, Reid & Wilson, 1991 from the Eastern and Southeastern Asian seas. *Journal of the Marine Biological Association of the United Kingdom*, 101 (2), 279-307.
- Villalobos-Guerrero, T.F., Kara, J., Simon, C., Idris, I., 2022. Systematic review of *Neanthes* Kinberg, 1865 (Annelida: Errantia: Nereididae) from southern Africa, including a preliminary molecular phylogeny of the genus. *Marine Biodiversity*, 52 (2), 21.
- Wang, Y., Cheng, H., Wang, C., 2021. A new eyeless species of *Nicon* (Annelida: Nereididae) from the deep Northwest Pacific Ocean. *Acta Oceanologica Sinica*, 40 (12), 20-26.
- Wilson, R.S., Glasby, C.J., 1993. A revision of the *Perinereis* species group (Polychaeta: Nereididae). *Records of the Australian Museum*, 45, 253-277.
- Winnepeninckx, B., Backeljau, T., De Wachter, R., 1993. Extraction of high molecular weight DNA from molluscs. *Trends Genetics*, 9, 407.
- Wu, B.L., Sun, R., Yang, D.J., 1985. *The Nereidae (polychaetous annelids) of the Chinese coast*. Springer-Verlag China Ocean Press Beijing, Berlin, 234 pp.
- Zenetos, A., Gofas, S., Verlaque, M., Çinar, M.E., Garcia Raso, *et al.*, 2010. Alien species in the Mediterranean Sea by 2010. A contribution to the application of European Union's Marine Strategy Framework Directive (MSFD). Part I. Spatial distribution. *Mediterranean Marine Science*, 11 (2), 381-493.