

# THE ECHINODERM NEWSLETTER

Number 18.

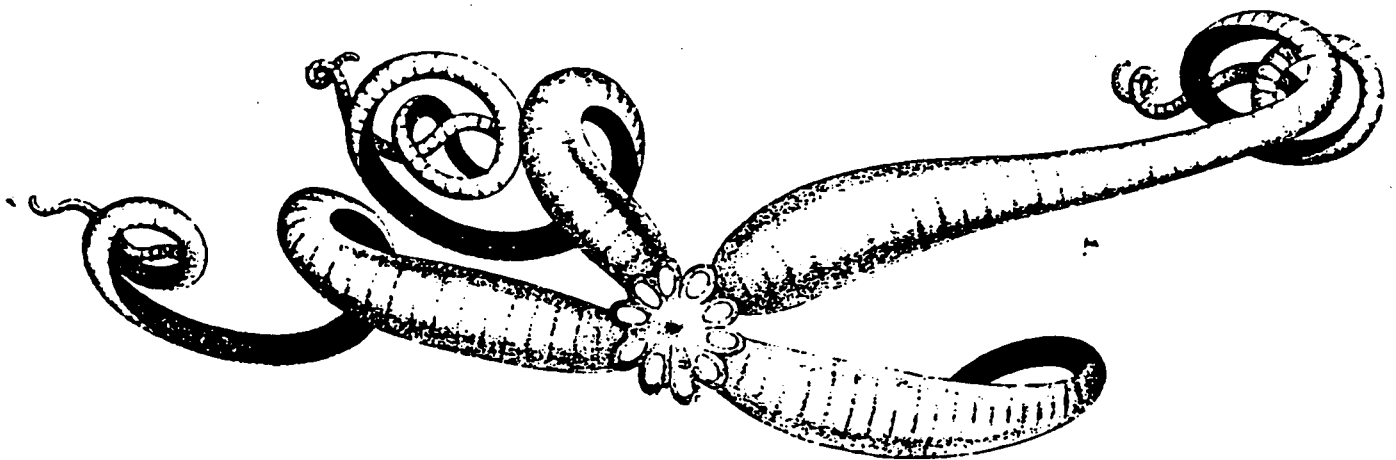
1993

Editor: John Lawrence  
Department of Biology  
University of South Florida  
Tampa, Florida 33620, U.S.A.

Distributed by: David Pawson  
Department of Invertebrate Zoology  
Smithsonian Institution  
Washington, D.C. 20560, U.S.A.

The newsletter contains information concerning meetings and conferences, publications of interest to echinoderm biologists, titles of theses on echinoderms, and research interests and addresses of echinoderm biologists. Individuals who desire to receive the newsletter should send their name, address, and research interests to the editor.

The newsletter is not intended to be a part of the scientific literature and should to be cited, abstracted, or reprinted as a published document.



Koehler  
1904

## Table of Contents

<b>Echinoderm specialists</b>	
<b>Addresses</b> .....	1
<b>Telephone - FAX - e-mail numbers</b> .....	39
<b>Requests and Information</b> .....	45
<b>Books</b> .....	48
<b>Recent publications on echinoderms</b> .....	59
<b>Theses</b>	
<b>Belgium</b> .....	92
<b>Canada</b> .....	92
<b>Chile</b> .....	92
<b>Colombia</b> .....	93
<b>France</b> .....	93
<b>Germany</b> .....	94
<b>Ghana</b> .....	94
<b>Ireland</b> .....	94
<b>Jamaica</b> .....	94
<b>Norway</b> .....	94
<b>Pakistan</b> .....	95
<b>Puerto Rico</b> .....	95
<b>Republic of China</b> .....	95
<b>Russia</b> .....	95
<b>Sweden</b> .....	95
<b>United Kingdom</b> .....	95
<b>United States</b> .....	96
<b>Papers presented at meetings</b>	
<b>International Coral Reef Symposium</b> .....	98
<b>Zoological Society of Japan</b> .....	99
<b>American Society of Zoologists</b> .....	102
<b>North American Friends of Echinoderms</b> .....	104
<b>Geological Society of America</b> .....	108
<b>Ailsa's Section</b> .....	111
<b>Echinoderm biologists (part 3)</b>	
<b>Alexander Emmanuel Rodolphe Agassiz</b> .....	131
<b>(J.M. Anderson, C. Birkeland, C. Conand,</b>	
<b>A. Farmanfarmanian, L. Fenaux, J.C. Ferguson,</b>	
<b>L. Fischelson, N.D. Holland, M. Jangoux,</b>	
<b>C. Johnson, D. Levitan, G. Manchenko,</b>	
<b>K.J. McNamara, R. Mooi, V.W. Pentreath,</b>	
<b>M. Propp, R. Scheibling, M. Sibuet, J. Woodley)</b> ...	135
<b>Obituary: James Eric Smith</b> .....	149

ADAMS, NIKKI. DEPARTMENT OF ZOOLOGY, 5751 MURRAY HALL, ROOM 100, UNIVERSITY OF MAINE, ORONO, ME 04469-0146, USA

AGATSUMA, YUKIO. HOKKAIDO CENTRAL FISHERIES, EXPERIMENTAL STATION, HAMANAKA 238, YOICHI HOKKAIDO, JAPAN

ALBUQUERQUE, MARIA. DEPT DE CIENCIAS BIOLOGICA, UNIVERSIDAD SANTA URSULA, RUA FERNANDO FERRARI, 75 BOTAFOL, RIO DE JANEIRO, BRAZIL

ALI, MOHAMED SAID M. DEPARTMENT OF GEOLOGY, FACULTY OF SCIENCE, EL MINIA UNIVERSITY, EL MINIA, EGYPT

ALLEN, JOHN A. UNIVERSITY MARINE BIOLOGICAL STATION, MILLPORT, ISLE OF CUMBRAE, SCOTLAND, UK

ALVA, VICTOR. BIOLOGIE MARINE (CP.160), UNIVERSITE LIBRE DE BRUXELLES, 50, AV.F.D. ROOSEVELT, BRUXELLES, BELGIUM

ALVAREZ, LEONARDO R. COLORADO #79, NAPOLES, MEXICO D.F. 03810, MEXICO

ALVAREZ, MARTINEZ DE. DEPARTAMENTO DE ZOOLOGIA, UNIVERSIDAD DE LA LAGUNA, TENERIFE, CANARY ISLANDS

ANDACHT, TRACY. DUKE UNIVERSITY LAB., PIVERS ISLAND, BEAUFORT, NC 28516, USA.

ANDERSON, EDWIN J. GEOLOGY DEPARTMENT, TEMPLE UNIVERSITY, PHILADELPHIA, PA 19122, USA

ANDERSON, JOHN M. 110 ROAT ST., ITHACA, NY 14850, USA.

ANDERSON, ROLAND. PUGET SOUND BIOLOGIST, THE SEATTLE AQUARIUM, PIER 59, SEATTLE, WA 908101, USA

ANDRADE, HECTOR. UNIVERSIDAD DE VALPARAISO, INSTITUTO DE OCEANOLOGIA, CASILLA 13-D, VINA DEL MAR, CHILE

ARANGUIZ, CHITA B.G. DEPTO. BIOLOGIA MARINA, UNIVERSIDAD CATOLICA DEL NORTE, CASILLA 117, COQUIMBO, CHILE

ARENDT, YURII A. PALEONTOLOGICAL INSTITUTE, RUSSIAN ACADEMY OF SCIENCES, PROF SOYUSNAYA STR. 123, 117647 MOSCOW, RUSSIA

ARNAUD, PATRICK M. CENTRE D'OCEANOLOGIE DE MARSEILLE,  
STATION MARINE D'ENDOUME, 13007 MARSEILLE, FRANCE

ARONSON, RICHARD B., C/O DEPT. INVERTEBRATE ZOOLOGY, MAIL  
STOP 163, SMITHSONIAN INSTITUTION, WASHINGTON, DC 20560, USA

ARTECHE, INAKI. DEPARTAMENTO DE BIOLOGIA (ZOOLOGIA), FACULTAD DE  
CIENCIAS, APARTADO 644, BALBOA, SPAIN

AUSICH, WILLIAM L. DEPT. OF GEOLOGICAL SCIENCES, 155 SOUTH OVAL  
MALL, THE OHIO STATE UNIVERSITY, COLUMBUS, OH 43210, USA

AUSTIN, WILLIAM. KHOYATAN MARINE LABORATORY, 4635 ALDER GLEN  
ROAD, RR1, COWICHAN BAY, BRITISH COLUMBIA, VOR INO CANADA

BAKER, ALAN. NATIONAL MUSEUM OF NEW ZEALAND, TE PAPA  
TONGAREWA, PO BOX 467, WELLINGTON, NEW ZEALAND

BALL, BRENDAN JOHN. MARTIN RYAN MARINE SCIENCE INSTITUTE,  
ZOOLOGY DEPARTMENT, UNIVERSITY COLLEGE GALWAY, GALWAY,  
IRELAND

BALL, MARY. DEPT. OF ZOOLOGY, AUBURN UNIVERSITY, AUBURN, AL  
36849, USA

BALSER, ELIZABETH. DEPT OF BIOLOGICAL SCIENCES, 132 LONG HALL,  
CLEMSON UNIVERSITY, CLEMSON, SC 29634-1903, USA

BARANOVA, Z.I. ZOOLOGICAL INSTITUTE, RUSSIAN ACADEMY OF  
SCIENCES, ST. PETERSBURG 199164, RUSSIA

BARKER, MICHAEL. PORTOBELLO MARINE LABORATORY, PO BOX 8,  
PORTOBELLO, DUNEDIN, NEW ZEALAND

BARON, CLIFFORD. DEPT. OF ZOOLOGY, UNIVERSITY OF CALIFORNIA,  
BERKELEY, CA 94720, USA

BARTSCH, ILSE. BIOLOGISCHE ANSTALT HELGOLAND, NORTHESTR. 31,  
2000 HAMBURG 51, GERMANY

BASCH, LARRY. INSTITUTE OF MARINE SCIENCES, 272 APPLIED  
SCIENCES, UNIVERSITY OF CALIF SANTA CRUZ, SANTA CRUZ, CA 95064,  
USA

BASKAR, B.K. CENTRAL MARINE FISHERIES R.I., E.R.G. ROAD,  
ERNACULAM, COCHIN-682031, INDIA

BAUMILLER, TOMASZ. DEPT. OF GEOPHYSICAL SCIENCES, UNIVERSITY OF CHICAGO, 5734 S. ELLIS AVE., CHICAGO, IL 60637, USA

BAY-SCHMITH, ENRIQUE. INSTITUTO DE BIOLOGIA CELULAR, UNIVERSIDAD DE CONCEPCION, CONCEPCION, CHILE

BAZHIN, ALEXANDER. 18 NABEREZHNYAYA, KAMCHATNIRO, PETROPAVLOVSK-KAMCHATSKY 68302, RUSSIA

BEAMON, JOE. WEST GEORGIA COLLEGE, CARROLLTON, GA 30118, USA

BEAVER, HAROLD. DEPT. OF GEOLOGY, BAYLOR UNIVERSITY, WACO, TX 76798, USA

BELYAEV, GEORGE M. P.P. SHIRSHOV INSTITUTE OF OCEANOLOGY, KRASIKOVA STR. 23, MOSCOW 117218, RUSSIA

BELL, BRUCE. 2601 NW EXPRESSWAY, SUITE 333W, OKLAHOMA CITY, OK 73112, USA

BENNETT, LAURA. BODEGA MARINE LABORATORY, BAY FLAT ROAD, BODEGA BAY, CA 94923, USA

BERENTS, PENELOPE. INVERTEBRATE ZOOLOGY, THE AUSTRALIAN MUSEUM, COLLEGE ST., SYDNEY, NSW, AUSTRALIA

BERGER, JACQUES. UNIVERSITY OF TORONTO, DEPARTMENT OF ZOOLOGY, TORONTO, ONTARIO M5S 1A1, CANADA

BILLETT, DAVID. INSTITUTE OCEANOGRAPHIC SCIENCE, DEACON LABORATORY, BROOK ROAD, WORMLEY, GODALMING, SURREY GU8 SUB, ENGLAND, U.K.

BIRENHEIDE, RUDIGER. C/O TATSUO MOTOKAWA; BIOL. LAB, TOKYO INSTITUTE OF TECHNOLOGY, OOKAYAMA, MEGURO-KU, TOKYO 152, JAPAN

BIRKELAND, CHARLES. MARINE LABORATORY, UNIVERSITY OF GUAM STATION, MANGILAO, GU 96923, USA

BLACK, W. ROBERT. DEPARTMENT OF ZOOLOGY, THE UNIV. OF WEST AUSTRALIA, NEDLANDS, WESTERN AUSTRALIA, 6009, AUSTRALIA

BLAKE, DANIEL. DEPT. GEOLOGY 245 NHB, UNIVERSITY OF ILLINOIS, 1301 W. GREEN ST., URBANA, IL 61801, USA

BOCKELIE, JOHAN F. NORSK HYDRO EXPLORATION, PO BOX 200, N-1321,  
STABEKK, NORWAY

BOXZAROWSKI, ANDREZEJ. SILESIAN UNIVERSITY, DEPT. OF EARTH  
SCIENCE, MIELCZARSKI8EGO STR. 60, 41-200 SOSNOWIEC, POLAND

BOOTH, BILLY B. JR. MOTE MARINE LABORATORY, 1600 CITY ISLAND  
PARK, SARASOTA, FL 33577, USA

BORZONE, C.A. CENTRO DE BIOLOGIA MARINHA, UNIVERSIDADE  
FEDERALE PARANA, 83200 PARANAGUA, PARANA, BRAZIL

BOSCH, ISIDRO. DEPARTMENT OF BIOLOGY, STATE UNIVERSITY OF NEW  
YORK, GENESEO, NY 14454, USA

BOTTJER, DAVID. DEPT. OF GEOLOGICAL SCIENCES, UNIVERSITY OF  
SOUTHERN CALIFORNIA, LOS ANGELES, CA 90089, USA

BOUDOURESQUE, C.F. FACULTE DES SCIENCES DE LUMINY,  
LABORATOIRE D'ECOLOGIE DU BENTHOS, 13288 MARSEILLE CEDEX 9,  
FRANCE

BOUGOIN, ALLAIN. CENTRE UNIVER. DE SHIPPAGAN, UNIVERSITE DE  
MONCTON, SHIPPAGAN, NEW BRUNSWICK, EOB 2PO CANADA

BOULAND, CATHERINE. DEPARTEMENT DE BIOLOGIE, UNIVERSITE  
LAVAL, QUEBEC, QC G1K 7P4, CANADA

BRANDT, DANITA, DEPT. OF GEOGRAPHY & GEOLOGY, EASTERN  
MICHIGAN UNIVERSITY, YPOSILANTI, MI 48197, USA

BRANSTRATOR, JON W. DEPARTMENT OF GEOLOGY, EARLHAM  
COLLEGE, RICHMOND, IN 47374, USA

BRAY, RICHARD. 176 WEST MAIN ST., PROT JERVIS, NY 12771, USA

BREIMER, ALBERT. RIJKSMUSEUM VAN GEOL MINERALOG,  
GARENMARKT 1B, 2311 PG LEIDEN, NETHERLANDS

BRETON, GERARD. MUSEUM D'HISTOIRE NATURELLE, PLACE DU VIEUX  
MARCHE, 76600 LE HAVRE, FRANCE

BRETT, CARLETON. DEPT. OF GEOLOGICAL SCIENCES, UNIVERSITY OF  
ROCHESTER, ROCHESTER, NY 14627, USA

BRITO, IGNACIO M. INSTITUTO DE GEOCIENCIAS, U.F.R.J.-C.C.M.N., ILHA O FUNDAO, 21910 RIO DE JANEIRO, BRAZIL

BRITTO, MAURO. DEPT. DE ZOOLOGIA-SALA 356, UNIVERSIDADE FEDERAL DO PARANA, CXPOSTAL 3034, CURITIBA, CEP 80.000 BRAZIL

BROADHEAD, THOMAS. UNIVERSITY OF TENNESSEE, DEPT. OF GEOLOGICAL SCIENCE, KNOXVILLE, TN 37996, USA

BROWER, JAMES. DEPARTMENT OF GEOLOGY, SYRACUSE UNIVERSITY, SYRACUSE, NY 13244-1070, USA

BRUNEL, PIERRE. DEPART. DE SCIENCES BIOLOGIQUES, UNIVERSITE DE MONTREAL, CASSIER POSTAL 6128, SUCCURSALE A., MONTREAL, QUEBEC H3C 3J7, CANADA

BUTTRON, BLANCA E. INSTITUTO DE GEOLOGIA, UNAM, CD. UNIVERSITARIA, DELEG, COYOACAN, 04510 MEXICO, D.F., MEXICO

BUREAU, FABRICE. LABORATOIRE BIOLOGIE MARINE 73, AVENUE MARSTRIAU (BATIMENT 4), 7000 MONS, BELGIUM

BURKE, ROBERT. DEPARTMENT OF BIOLOGY, UNIVERSITY OF VICTORIA, VICTORIA, BRITISH COLOMBIA, V8W 2F2, CANADA

BURKE, TOM. DEPT. OF THE INTERIOR, MARINE MINERALS, B.L.M. 753, 18TH & C STREETS NW, WASHINGTON, DC 20242, USA

BURTON, MARGARET. DEPARTMENT OF BIOLOGY, MEMORIAL UNIVERSITY OF NEWFLD., ST JOHN'S, NEWFOUNDLAND A1B 3X9, CANADA

BUSSARAWIT, SOMCHAI. PHUKET MARINE BIOLOGICAL CENTER, PO BOX 60, PHUKET 83000, THAILAND

BUSTOS, EDUARDO. INSTITUTO DE FOMENTO PESQUERO, AVE. DIEGO PORTALES 1450, PUERTO MONTT, CHILE

BYRNE, MARIA. DEPARTMENT OF EMBRYOLOGY AND HISTOLOGY, 5-13 UNIVERSITY OF SYDNEY, SYDNEY N.S.W. 2006, AUSTRALIA

CAFFI, MARGARITA G. DEPTO DE ZOOLOGIA, UNIVERSIDADE DE CONCEPCION, CASILLA 1367, CONCEPCION, CHILE

CALDWELL, JOHN W. DEPT. ENVRION ENGINEERING SCI, UNIVERSITY OF FLORIDA, 111 BLACK HALL, GAINESVILLE, FL 32611, USA

CAMARGO, TANIA MARIA. INSTITUTO OCEANOGRAFICO, UNIVERSIDADE DE SAO PAULO, SAO PAULO, BRAZIL

CAMERON, R. ANDREW. DIVISION OF BIOLOGY 156-29, CALIFORNIA INSTITUTE OF TECH, PASADENA, CA 91125, USA

CAMPBELL, ANDREW. SCHOOL OF BIOLOGICAL SCIENCES, QUEEN MARY & WESTFIELD COLLEGE, UNIVERSITY OF LONDON, MILE END ROAD, LONDON E1 4NS, ENGLAND

CAMPBELL, DAVID B. BIOLOGY DEPARTMENT, RIDER COLLEGE, 2083 LAWRENCEVILLE ROAD, LAWRENCEVILLE, NJ 08648, USA

CANNON, L.R.G. QUEENSLAND MUSEUM, PO BOX 3300, SOUTH BRISBANE, QUEENSLAND 4101, AUSTRALIA

CARCAMO, ALFONSO G. UNIVERSIDAD DE CHILE-OSORNO, CASILLA 933, OSORNO, CHILE

CAREY, ANDREW G. JR. SCHOOL OF OCEANOGRAPHY, OREGON STATE UNIVERSITY, CORVALLIS, OR 97331, USA

CARNEVALI, M.D.C. DIPARTIMENTO DI BIOLOGIA, UNIV. DEGLI STUDI DI MILANO, VIA CELORIA 26, 20133 MILANO, ITALY

CARNEY, ROBERT. COASTAL RESOURCES LABORATORY, LOUISIANA STATE UNIVERSITY, BATON ROUGE, LA 70803, USA

CARPENTER, ROBERT C. DEPT. OF BIOLOGY, CALIFORNIA STATE UNIVERSITY, NORTHRIDGE, CA 91330, USA

CASTILLO, JUAN CARLOS. ECOLOGIA MARINA, FAC. C BIOLÓGICAS, U. CATOLICA DE CHILE, CASILLA 114-D, SANTIAGO, CHILE

CASTRO, LILY ROMINA. C/O CICESE, PO BOX 434843, SAN DIEGO, CA 92143-4843, USA

CHAET, A.B. DEPT. OF CELL AND MOLEC. BIOL., UNIVERSITY OF WEST FLORIDA, PENSACOLA, FL 32514, USA

CHANG, CLIFFORD. CHEMISTRY DEPARTMENT, UNIVERSITY OF WEST FLORIDA, PENSACOLA, FL 32504, USA

CHAO, SHYH-MIN. DEPARTMENT OF BIOLOGY, TUNG HAI UNIVERSITY, TAICHUNG, TAIWAN, REPUBLIC OF CHINA



CHEN, CHANG-PO. INSTITUTE OF ZOOLOGY, ACADEMIA SINICA,  
NANKAN, TAIPEH, TAIWAN, 11529 REPUBLIC OF CHINA

CHIA, FU-SHIANG. DEPARTMENT OF ZOOLOGY, UNIVERSITY OF  
ALBERTA, EDMONTON, ALBERTA, T6G 2E9 CANADA

CHITA, BERNADA G. A. ECOLOGIA MARINA PONT, UNIVERSIDAD  
CATOLICA DE CHILE, CASILLA 114-D, SANTIAGO, CHILE

CHIU, SEIN TUCK. DEPARTMENT OF BIOLOGY, HONG KONG BAPTIST  
COLLEGE, 224 WATERLOO ROAD, KOWLOON, HONG KONG

CHRISTENSEN, AAGE MOELLER. MARINBIOLOGISK LAB. KOBENHAVNS,  
UNIVERSITET STRANDPROMENDADEN, 30000 HELSINGER, DENMARK

CLARK, AILSA MCGOWN. GYLLYNGDUNE, SOUTH ROAD, WIVELSFIELD  
GREEN, HAYSWARDS HEATH, SUSSEX, ENGLAND RH17 7QS, UK

COBB, JAMES L.S. GATTY MARINE LAB, ST. ANDREWS, FIFE, SCOTLAND,  
U.K.

CODOCEO, MARIA. SECCION HIDROBIOLOGIA, MUSEO NACIONAL HIST.  
NATURAL, CASILLA 787, SANTIAGO, CHILE

COLON-JONES, ELIZABETH. DIV. MARINE BIOLOGY FISHERIES, RSMAS,  
UNIVERSITY OF MIAMI, 4600 RICKENBACKER CAUSEWAY, MIAMI, FL  
33149, USA

CONAND, CHANTAL. LABORATOIRE BIOLOGIE MARINE, UNIVERSITE DE  
LA REUNION, 15 AVENUE RENE CASSIN, 97489 SAINT-DENIS CEDEX, LA  
REUNION, FRANCE

CONCEPCION, MARCOS. DEPARTAMENTO DE ECOLOGIA, FACULTAD DE  
BIOLOGIA, UNIVERSIDAD DE MURCIA, MURCIA, SPAIN

CONLU, PRUDENCIA. COLLEGE OF FISHERIES, UNIVERSITY OF  
PHILIPPINES IN THE VISAYAS, MIAGAO, ILOILO, THE PHILIPPINES

CONSTABLE, ANDREW. FACULTY ENVIRONMENTAL SCIENCES, GRIFFITH  
UNIVERSITY, NATHAN, QUEENSLAND, AUSTRALIA 4111

COSTA, HENRIQUE R. DPTO BIOLOGIA MARINA, UFRJ, B1 A CCS-1  
FUNDAO, RIO DE JANEIRO, 21910 BRAZIL

COSTELLOE, JOHN. ZOOLOGY DEPARTMENT, UNIVERSITY COLLEGE,  
GALWAY, IRELAND

COUILLARD, PIERRE. DEPT. DE SCIENCES BIOLOGIQUES, UNIVERSITE DE MONTREAL, CP6128 SUCC A, MONTREAL, H3C 3J7 CANADA

COULON, P. LAB. DE BIOLOGIE MARINE CP 160, UNIVERSITE LIBRE DE BRUXELLES, 50 AV F.D. ROOSEVELT, B-1050 BRUXELLES, BELGIUM

COWDEN, CINDY. 123 ZOOLOGY RESEARCH, 1117 W JOHNSON ST., MADISON, WI 53706, USA

COX, ROBERT. DEPART. OF GEOLOGICAL SCIENCE, UNIVERSITY OF MICHIGAN, ANN ARBOR, MI 48109, USA

CRUMP, ROBIN. ORIELTON FIELD CENTRE, PEMBROKE SA7 15E2, DYFED, PEMBROKE, ENGLAND

CUENCA, CATHERINE. MUSEUM HISTOIRE DE NANTES, 12 RUE VOLTAIRE, 44000 NANTES, FRANCE

CUTRESS, BERTHA. DEPT. OF MARINE SCIENCES, UNIVERSITY OF PUERTO RICO, PO BOX 5000, MAYAGUEZ, PR 00709, USA

DAFNI, JACOB. EILAT COLLEGE, PO BOX 1301, EILAT 88112, ISRAEL

DAHORA, MARIA MP SILVA. MUSEU NACIONAL DA QUIN BV/UFRJ, QUINTA DE BOA VISTA, SAO CHRISTOVAO, RIO DE JANEIRO, 20942 BRAZIL

D'ANDREA, ANTHONY F. MARINE SCIENCE RESEARCH CENTER, SUNY AT STONY BROOK, STONY BROOK, NY 11794, USA

DAVID, BRUNO. CENTRE DE SCIENCES DE LA TERRE, UNIVERSITE DE BOURGOGNE, 6, BD. GABRIEL, 21000 DIJON, FRANCE

DAYTON, PAUL K. III. DEPT. OF ZOOLOGY, PKO A001, SCRIPPS INSTITUTION OF OCEANOGRAPHY, LA JOLLA, CA 92093, USA

DEARBORN, JOHN H. DEPARTMENT OF ZOOLOGY, 5751 MURRAY HALL, UNIVERSITY OF MAINE, ORONO, ME 04469-5751, USA

DECELIS, K. ALEJANDRO. NATIONAL MUSEUM, EXECUTIVE BUILDING, RIZAL PARK, ERMITA, MANILA, PHILIPPINES

DEGREEF, YVES. LABORATOIRE BIOLOGIE MARINE, C.P. 160, UNIVERSITE LIBRE DE BRUXELLES, AVE. F.D. ROOSEVELT 50, B 1050 BRUXELLES, BELGIUM

DELAVALT, ROBERT. LAB BIOL CELLULAIRE & ANIMALE, UNIVERSITE D'ORLEANS, 45045 ORLEANS CEDEX, FRANCE

DELMAS, P. LAB. DE ZOOL. MARINE, CERAM UNIVERSITE D'AIX-MARSEILLE III, AVE ESCADRILLE NORM-NIEMEN, (CP 342), F-13397, MARSEILLE CEDEX 13, FRANCE

DEMARGNE, MICHAEL. RUE DE L'ANCIENNE MAIRIE 38730, VIRIEU SUR BOURBRE, FRANCE

DE MOURA-BRITTO, MAURO. INSTITUTO DE TERRAS, CARTOFRAFIA E FLORESTAS, RUE DESEMBARGADOR, MOTTA, 3384-80610 CURITIBA, BRAZIL

DE RIDDER, CHANTAL. LABORATOIRE DE BIOLOGIE MARINE C.P. 160, UNIVERSITE LIBRE DE BRUXELLES, AVE. F.D. ROOSEVELT, B-1050 BRUXELLES, BELGIUM

DEUTZMANN, HELMUT. STUECKERSTR. 4, 4000 DUESSELDORF, GERMANY

DIEHL, WALTER, DEPT. OF BIOLOGICAL SCIENCE, MISSISSIPPI STATE UNIVERSITY, PO DRAWER BY, MISSISSIPPI STATE, MS 39762, USA

DIX, TREVOR G. FISHERIES RESEARCH LABORATORY, TAROONA, TASMANIA 7006, AUSTRALIA

DOBSON, WILLIAM E. DEPT. OF BIOLOGY, APPALACHIAN STATE UNIVERSITY, BOONE, NC 28608, USA

DOI, TERUO. LAB ECOLOGY, FAC SCIENCE, KYUSHU UNIVERSITY 33, FUKUOKA, JAPAN

DOLMATOV, IGOR YU. LABORATORY OF COMPARATIVE CYTOLOGY, INSTITUTE OF MARINE BIOLOGY, VLADIVOSTOK 690032, RUSSIA

DONOVAN, STEPHEN. DEPARTMENT OF GEOLOGY, UNIVERSITY OF THE WEST INDIES, MONA, KINGSTON 7, JAMAICA

DOTAN, AARON. DEPARTMENT OF ZOOLOGY, TEL AVIV UNIVERSITY, RAMAT-AVIV, 69978 ISRAEL

DOTY, J.E. 1224 SOUTH FOURTH STREET, ALHAMBRA, CA 91801, USA

DOWNEY, MAUREEN, E. 443 SUTHERLAND ROAD, FRIDAY HARBOR, WA 98250, USA

DROZDOV, ANATOLY L. INSTITUTE OF MARINE BIOLOGY, FAR EAST  
BRANCH, ACADEMY OF SCIENCES OF USSR, VLADIVOSTOK 690032, RUSSIA

DUBE, FRANCOIS. DEPT. OF OCEANOGRAPHY, UNIV. DU QUEBEC A  
RIMOUSKI, RIMOUSKI, QUEBEC, G5L3A1 CANADA

EBERT, THOMAS. A. DEPARTMENT OF BIOLOGY, SAN DIEGO STATE  
UNIVERSITY, SAN DIEGO, CA 92183, USA

ECKELBARGER, KEVIN. DARLING MARINE CENTER, UNIVERSITY OF  
MAINE, 25 CLARK'S COVE ROAD, WALPOLE, ME 04573, USA

ECKERT, JAMES D. 16 WEST HAMPTON RD., ST. CATHERINES, ONTARIO  
L2T 3E5, CANADA

ECKHAUT, IGOR. UNIVERSITE DE MONS, LAB. DE BIOLOGIE MARINE, 19  
AV. MAISTRIAU, 7000 MONS, BELGIUM

EERNISSE, DOUGLAS J. MUSEUM OF ZOOLOGY, UNIVERSITY OF  
MICHIGAN, ANN ARBOR, MI 48109, USA

ELLERS, OLAF. DEPARTMENT OF ZOOLOGY, UNIVERSITY OF  
CALIFORNIA, DAVIS, CA 95616, USA

EMERSON, CAROLYN J. BIOLOGY DEPARTMENT, MEMORIAL UNIV. OF  
NEWFOUNDLAND, ST. JOHN'S, NEWFOUNDLAND, A1B 3X9 CANADA

EMLET, RICHARD B. OREGON INST. OF MARINE BIOLOGY, UNIVERSITY  
OF OREGON, CHARLESTON, OR 97420, USA

EMSON, ROLAND. DIVISION OF LIFE SCIENCES, KINGS COLLEGE  
LONDON, KENSINGTON CAMPUS, CAMPDEN HILL RD, LONDON W87AH,  
ENGLAND, UK

ENDELMAN, LEONID G. PALEONTOLOGICAL INSTITUTE, RUSSIAN  
ACADEMY OF SCIENCES, PROFSOYUSNAYA UL. 123, 117647 MOSCOW,  
RUSSIA

ENGSTROM, NORMAN. COLLEGE OF LIBERAL ARTS & SCIENCES,  
NORTHERN ILLINOIS UNIVERSITY, DEKALB, IL 60115, USA

ESCOUBET, P. MARINELAND-AQUARIUMS, 306 AVENUE MOZART, 06600  
ANTIBES, FRANCE

ETTENSON, FRANK R. DEPT. OF GEOLOGICAL SCIENCES, UNIVERSITY OF  
KENTUCKY, LEXINGTON, KY 40504, USA

EVDOKIMOV, VLADIMIR V. PACIFIC RESEARCH INSTITUTE OF FISHERIES  
AND OCEANOGRAPHY, LABORATORY OF PHYSIOLOGY, 690600  
VLADIVOSTOK, RUSSIA

EYLERS, JOHN P. N.C. MUSEUM OF LIFE & SCIENCE, 433 MURRAY AVE.,  
PO BOX 15190, DURHAM, NC 27704, USA

FANKBONER, PETER V. DEPT. OF BIOLOGICAL SCIENCES, SIMON FRASER  
UNIVERSITY, BURNABY, B.C. V5A 1S6, CANADA

FARMANFARMAIAN, ALLAHVERDI. NELSON BIOLOGY LAB, RUTGERS  
UNIVERSITY, PO BOX 1059, PISCATAWAY, NJ 08856, USA

FAY, ROBERT O. OKLAHOMA GEOLOGICAL SURVEY, UNIVERSITY OF  
OKLAHOMA, 830 S. UVAL, NORMAN, OK 73019, USA

FECHTER, HUBERT. ZOOLOGISCHE STAATSSAMMLUNG,  
MUNCHHAUSENSTR. 21, D-8000 MUENCHEN 60, GERMANY

FEDER, HOWARD. INSTITUTE OF MARINE SCIENCE, UNIVERSITY OF  
ALASKA, FAIRBANKS, AK 99701, USA

FELL, JULIAN. 1545 WELLS PLACE, ERRINGTON, BRITISH COLUMBIA, VOR  
1VO CANADA

FENAUX, LUCIENNE. STATION ZOOLOGIQUE, (CEROV) BP 28, 06230  
VILLEGRANCHE SUR MER, FRANCE

FERAL, JEAN-PIERRE. OBSERVATOIRE OCEANOGRAPHIQUE, 66650  
BANYULS-SUR MER, FRANCE

FERGUSON, JOHN C. DEPARTMENT OF BIOLOGY, ECKERD COLLEGE,  
BOX 12560, ST. PETERSBURG, FL 33733, USA

FERRAND, JEAN-GUY. LAB BIOL CELLULAIRE & ANIMALE, UER  
SCIENCES, UNIVERSITE D'ORLEANS, F 45046 ORLEANS CEDEX, FRANCE

FIERRO, JUAN F.T. MUSEO DE HISTORIA NATURAL DE CONCEPCION,  
CASILLA 1054, CONCEPCION, CHILE

FISHELSON, LEV. DEPT. OF ZOOLOGY, TEL-AVIV UNIVERSITY, 155 HERZL  
ST., TEL-AVIV, ISRAEL

FLAMMANG, PATRICK. UNIVERSITE DE MONS, LABORATOIRE DE  
BIOLOGIE MARINE, (BAT. IV) 19, AV. MAISTRIAU, B-7000 MONS, BELGIUM

FLEMING, TIMOTHY P. 2025 OROFINO GULCH, HELENA, MT 59601, USA

FONTAINE, ARTHUR R. DEPT. OF BIOLOGY, UNIVERSITY OF VICTORIA,  
VICTORIA, B.C., V8W 2Y2, CANADA

FOSTER, MERRILL W. DEPT. OF GEOLOGICAL SCIENCES, BRADLEY  
UNIVERSITY, PEORIA, IL 61625, USA

FOSTER, ROBERT J. BHP PETROLEUM, GPO BOX 1911 R, MELBOURNE,  
VICTORIA 3000, AUSTRALIA

FOUDA, MOUSTAFA. DEPT. OF ZOOLOGY, AL-ASHAR UNIVERSITY, NASR-  
CITY, CAIRO, EGYPT

FOURNIER, DANIEL. MUSEUM D'HISTOIRE NATURELLE, 1 RUE  
DOLOMIEU, F-3800 GRENOBLE, FRANCE

FOX, DAVID J. ZOOLOGY DEPARTMENT, UNIVERSITY OF TENNESSEE,  
KNOXVILLE, TN 37996-0810, USA

FRANK, PETER. INVERTEBRATE ZOOLOGY DIVISION, NATIONAL  
MUSEUM OF NATURAL SCI, OTTAWA, ONTARIO, K1A 0M8 CANADA

FRANKEL, EDGAR. GEOLOGY DEPARTMENT, UNIVERSITY OF  
TECHNOLOGY, PO BOX 123, BROADWAY 2007, SYDNEY, AUSTRALIA

FRANKLIN, SUE. ZOOLOGY DEPARTMENT A08, SYDNEY UNIVERSITY,  
SYDNEY, NSW 2006, AUSTRALIA

FRANZ, DAVID R. BIOLOGY DEPT., BROOKLYN COLLEGE, CNY,  
BROOKLYN, NY 11210, USA

FRANZEN-BENGTSON, CHRISTINA. PALEONTOLOGISKA INSTITUTIONEN,  
BOX 558, S-751 22 UPPSALA, SWEDEN

FREST, TERENCE. DEPARTMENT OF GEOLOGY, UNIVERSITY OF IOWA,  
IOWA CITY, IA 52242, USA

FUJITA, TOSHIHIKO. TOHOKU NAT. FISH. RESEARCH INT, HACHINOHE,  
SHIMO-MEKURAKUBO 25-259, HACHINOHE 031, JAPAN

GAGE, JOHN D. DUNSTAFFNAGE MARINE LAB., PO BOX 3, OBAN,  
ARGYLL, SCOTLAND PA34 4AD, UK

- GAGNON, JEAN-MARC. OCEANOGRAPHIE CHIMIQUE, INSTITUT MAURICE-LAMONTAGNE, BOITE POSTALE 1000, MONT-JOLI (QC), G5H 3Z4 CANADA
- GALLEMI, JAUME. MUSEU DE GEOLOGIA, PARC DE LA CIUTADELLA S/N, 08003 BARCELONA, SPAIN
- GEBRUK, ANDREW V. INSTITUTE OF OCEANOGRAPHY, USSR ACADEMY OF SCIENCES, KRASSIKOVA STR., 23, MOSCOW 117 218, RUSSIA
- GENTIL, FRANCK. STATION BIOLOGIQUE, PLACE G, TEISSIER, F-29211 ROSCOFF, FRANCE
- GEORGE, SOPHIE. HARBOR BRANCH OCEANOGR. INST., 5600 OLD DIXIE HIGHWAY, FORT PIERCE, FL 34946, USA
- GERONIMO, ABELARDO L. NO 2 E SANTOS AVE (MEERALCO RD) BAMBANG, PASIG, METRO MANILA, PHILIPPINES
- GHYOOT, MARIANNE. LABORATOIRE DE BIOLOGIE MARINE, UNIVERSITE LIBRE DE BRUXELLES, 50 AV., F.D. ROOSEVELT, 1050 BRUXELLES, BELGIUM
- GIORDANO DE FREITAS, SOLANGE. MUSLEU NACIONAL QUINTA DA BOA VISTA/UFRJ, DEPARTAMENTAO DE INVERTEBRADOS, SAO CRISTOVAO, RJ CEP 20942, BRAZIL
- GIBSON, MICHAEL. DEPT. GEOSCIENCES AND PHYSICS, UNIVERSITY OF TENNESSEE, 222 EPS BUILDING, MARTIN, TN 38238, USA
- GIUDICE, GIOVANNI. DEPARTIMENTO DI VILOGIA, CELLULARE E DELLO SVILUPPO, VIA ARCHIRAFI 20-90123, PALERMO, ITALY
- GLUCHOWSKI, EDWARD. DEPARTMENT OF EARTH SCIENCES, SILESIAN UNIVERSITY, LAB. PALEONTOL. & STRATIGRAPHY, BEDZINSKA STR. 60, 41-200 SOSNOWIEC, POLAND
- GLYNN, PETER W. RSMAS, UNIVERSITY OF MIAMI, 4600 RICKENBACKER CAUSEWAY, MIAMI, FL 33149, USA
- GOLDBERG, AUTHUR S. DIVISION OF NATURAL SCIENCES, SOUTHAMPTON COLLEGE, SOUTHAMPTON, NY 11968, USA
- GOLDEN, JULIA. DEPT. OF GEOLOGY, 121 TROWBRIDGE HALL, UNIVERSITY OF IOWA, IOWA CITY, IA 52242-1379, USA

GOLDSCHMID, ALFRED. ZOOLOGISCHES INSTITUT, UNIVERSITÄT  
SALZBURG, AKADEMIESTR 26, A-5020 SALZBURG, AUSTRIA

GOODING, RICHARD U. 19 HIGHGATE PARK, ST. MICHAEL, BARBADOS,  
WEST INDIES

GORDON, CARLA M. UNIVERSITY OF THE WEST INDIES, MONA,  
KINGSTON 7, JAMAICA

GRABOWSKI, GAIL. KEWALO MARINE LABORATORY, UNIVERSITY OF  
HAWAII, 41 AHUI ST., HONOLULU, HI 96813, USA

GRANGE, KEN R. N2 OCEANOGRAPHIC INSTITUTE, PO BOX 14901,  
KILBIRNIE, WELLINGTON, NEW ZEALAND

GRAS, HERIBERT. ZOOLOGICAL INSTITUTE II, UNIV. KOLN, WEYERTAL  
119, 5000 KILN 41, GERMANY

GREENSTEIN, BENJAMIN J. DEPARTMENT OF GEOLOGY, SMITH  
COLLEGE, NORTHAMPTON, MA 01063, USA

GRUZOV, E.N. ZOOLOGICAL INSTITUTE, ACADEMY OF SCIENCES,  
LENINGRAD 199164, RUSSIA

GUENSBURG, THOMAS E. PHYSICAL SCIENCE DIVISION, ROCK VALLEY  
COLLEGE, 3301 N. MULFORD ROAD, ROCKFORD, IL 61101, USA

GUILLE, ALAIN. LABORATOIRE ARAGO, 66650 BANYULS-SUR-MER,  
FRANCE

GUILLOU, MONIQUE. LAB. D'OCEANOGRAPHIE BIOLOGIQUE, URA CNRS  
D1513, 6 AVENUE LE GORGEU, BP 452, 29.275 BREST CEDEX, FRANCE

GUISADO ARANGUIZ, CHITA B. INSTITUTO DE ZOOLOGIA, UNIVERSIDAD  
AUSTRAL DE CHILE, CASILLA 567, VALDIVIA, CHILE

GUTIERREZ, JUAN E. INSTITUTO INVEST. OCEANOLOGIAS,  
UNIVERSIDAD DE CHILE, CASILLA 1240, ANTOFAGASTA, CHILE

GUTT, JULIAN. ALFRED-WEGENER-INSTITUTE FÜR POLAR UND  
MEERESFORSCHUNG, COLUMBUS-STRASSE, D-2850 BREMERHAVEN,  
GERMANY

HAEDRICH, RICHARD L. NFLD. INST. FOR COLD OCEAN SCI, MEMORIAL  
UNIVERSITY, ST. JOHN'S, NEWFOUNDLAND A1B 3X7, CANADA



HAMADA, SPENCER. BIOLOGY DEPARTMENT, WEST GEORGIA COLLEGE,  
CARROLLTON, GA 30118, USA

HAMMOND, LAURENCE STUART. VICTORIA INSTI MARINE SCIENCES, 14  
PARLIAMENT PLACE, MELBOURNE, VICTORIA, AUSTRALIA

HARMELIN, JEAN-GEORGES. STATION MARINE D'ENDOUME, 13007  
MARSEILLE, FRANCE

HARRIOTT, VICKI. ZOOLOGY DEPARTMENT, UNIVERSITY OF  
QUEENSLAND, ST. LUCIA, 4067, BRISBANE, AUSTRALIA

HARRIS, LARRY G. ZOOLOGY DEPARTMENT, UNIVERSITY OF NEW  
HAMPSHIRE, DURHAM, NH 03824, USA

HARROLD, CHRISTOPHER, RESEARCH DIVISION, MONTEREY BAY  
AQUARIUM, 886 CANNERY ROW, MONTEREY, CA 93940, USA

HARTMANN, G. ZOOLOGISCHES INSTITUT, ZOOLOGISCHES MUSEUM,  
MARTIN LUTHER KING PLATZ 3, 2 HAMBURG 13, GERMANY

HARTSOCK, FRANKLIN B. ALASKA FISHERIES SCIENCE CENTER, KODIAK  
INVESTIGATIONS, PO BOX 1638, KODIAK, AK 99615-1638, USA

HAUDE, REIMUND. INSTITUTE MUSEUM GEOL PALAONT, GOLDSCHMIDT  
STR. 3, D-3400 GOTTINGEN, GERMANY

HAY, MARK. INSTITUTE OF MARINE SCIENCE, 3407 ARENDELL STREET,  
MOREHEAD CITY, NC 28557, USA

HAYASHJI, HJIROSHI. SUGASHIMA MARINE BIOL. LAB., NAGOYA  
UNIVERSITY, SUGASHIMA, TOBA, MIE 517, JAPAN

HECKER, R.T. PALEONTOLOGICAL INSTITUTE, ACADEMY OF SCIENCE  
USSR, LENINSKY PROSPECT 33, MOSCOW V-71, RUSSIA

HEDDLE, DUNCAN. DEPARTMENT OF ZOOLOGY, UNIVERSITY OF  
ABERDEEN, TILLYDRONE AVE., ABERDEEN AB9 2TN, SCOTLAND

HELLER, JAMES A. DEPT. OF GEOLOGICAL SCIENCES, THE UNIVERSITY  
OF TENNESSEE, KNOXVILLE, TN 37921, USA

HENDLER, GORDON. LIFE SCIENCES, L.A. COUNTY MUSEUM NAT. HIST.,  
900 EXPOSITION BLVD., LOS ANGELES, CA 90007, USA

HERRERA, JOAN. DEPARTMENT OF ZOOLOGY, UNIVERSITY OF FLORIDA, GAINESVILLE, FL 32611, USA

HERRING, P.J. INSTTT. OCEANOGRAPHIC SCIENCES, BROOK ROAD, WORMLEY, GODALMING, SURREY GU8 5UB, ENGLAND, UK

HESS, HANS K. IM GERSTENACKER 8, CH-4102 BINNGEN, SWITZERLAND

HICKEY, ANNE. BIOLOGY DEPARTMENT, UNIV. OF THE WEST INDIES, CAVE HILL, BARBADOS

HIDAKA, MICHIO. DEPARTMENT OF BIOLOGY, UNIVERSITY OF RYUKYUS, SHURI, NAHA, OKINAWA 903, JAPAN

HIGHSMITH, RAYMOND. INSTITUTE OF MARINE SCIENCES, UNIVERSITY OF ALASKA, FAIRBANKS, AK 99775-1080, USA

HILL, ROBERT B. DEPARTMENT OF ZOOLOGY, UNIVERSITY OF RHODE ISLAND, KINGSTON, RI 02881, USA

HIMMELMAN, JOHN H. DEPARTMENT DE BIOLOGIE, UNIVERSITE LAVAL, QUEBEC G1K 7P4, CANADA

HINES, GENE. DEPARTMENT OF BIOLOGY, UNIVERSITY OF ALABAMA AT BIRMINGHAM, UNV STATION, BIRMINGHAM, AL 35294, USA

HODGSON, ALAN N. DEPT. OF ZOOLOGY AND ENTOMOLOGY, RHODES UNIVERSITY, PO BOX 94, GRAHAMSTOWN 6140, SOUTH AFRICA

HOEBAUS, ERHARD. ZOOLOGISCHES INSTITUT DER, UNIVERSITAT WIEN, DR KARL LUEGER RING 1, A 1010 VIENNA, AUSTRIA

HOGGINS, DIANE DAVENPORT. 49 VIEW ST., SANDY BAY, HOBART, TASMANIA, 7005 AUSTRALIA

HOLTERHOFF, PETER. DEPARTMENT OF GEOLOGY, UNIVERSITY OF CINCINNATI, CINCINNATI, OH 45221, USA

HOPKINS, THOMAS S. DEPARTMENT OF BIOLOGICAL SCIENCES, UNIVERSITY OF ALABAMA, BOX 870344, TUSCALOOSA, AL 35487-0344, USA

HOROWITZ, ALAN STANLEY. DEPARTMENT OF GEOLOGY, INDIANA UNIVERSITY, 1005 EAST STREET, BLOOMINGTON, IN 47401, USA

HOSHI, MOTONORI. DEPARTMENT OF LIFE SCIENCE, TOKYO INST. TECHNOLOGY, NAGATSUTA, MIDORI-KU YOKOHAMA 227, JAPAN

HOSHIAI, T. NAT. INST. POLAR RESEARCH 9-10, KAGA 1-CHOME,  
ITABASHI-KU, TOKYO 173, JAPAN

HOTCHKISS, FREDERICK, H.C. 26 SHERRY ROAD, HARVARD, MA 01451,  
USA

HUDA, ISHRAT ARA. ZOOLOGICAL SURVEY DEPARTMENT, NISHTAR  
ROAD, KARACHI, PAKISTAN

IMAOKA, TOHRU. KATATA 28094-3, SHIRAHAMA, NISHIMURO,  
WAKAYAMA, 649-22 JAPAN

IRIMURA, SEIICHI. 1-49-1 OKUSAWA, SETAGAYA-KU, TOKYO, JAPAN 158

ISAEVA, VALERIA V. INSTITUTE OF MARINE BIOLOGY, 690032  
VLADIVOSTOK, RUSSIA

IWATA, K.S. DEPARTMENT OF BIOLOGY, OKAYAMA UNIVERSITY,  
TSUSHIMA NAKA 3 CHOME, OKAYAMA 700, JAPAN

JABLONSKI, DAVID. DEPT. GEOPHYSICAL SCIENCES, UNIVERSITY OF  
CHICAGO, CHICAGO, IL 60637, USA

JAECKLE, WILLIAM B. SMITHSONIAN ENVIRONMENTAL RESEARCH  
CENTER, PO BOX 28, EDGEWATER, MD 21037, USA

JAMES, D.B. RESEARCH CENTER OF CMFRI, 29 COMMANDER-IN-CHIEF  
ROAD, MADRAS 600105, INDIA

JANGOUX, MICHEL. LAB. DE BIOLOGIE MARINE, C.P. 160, UNIVERSITE  
LIBRE DE BRUXELLES, 50 AVENUE F.D. ROOSEVELT, B-1050 BRUXELLES,  
BELGIUM

JANIES, DANIEL. DEPARTMENT OF ZOOLOGY, UNIVERSITY OF FLORIDA,  
GAINESVILLE, FL 32611, USA

JARAMILLO, EDUARDO. INSTITUTO DE ZOOLOGIA, UNIVESIDAD  
AUSTRAL DE CHILE, CASILLA 567, VALDIVIA, CHILE

JEAL, FRANK. ZOOLOGY DEPARTMENT, TRINITY COLLEGE, DUBLIN 2  
IRELAND

JEFFERIES, R.P.S. DEPARTMENT OF PALEONTOLOGY, BRITISH MUSEUM,  
NATURAL HISTORY, CROMWELL ROAD, LONDON SW7 5BD, ENGLAND

JELLETT, JOANNE. 47 WAKE UP HILL ROAD, RR#1, CHESTER BASIN,  
NOVA SCOTIA, CANADA

JENKINS, ANNE. DEPT. OF MARINE BIOLOGY, UNIVERSITY OF  
LIVERPOOL, PORT ERIN, ISLE OF MAN, ENGLAND

JENKINS, KENNETH. DEPARTMENT OF BIOLOGY, CALIFORNIA STATE  
UNIVERSITY, LOS ANGELES, CA 90840, USA

JENSEN, MARGIT. ZOOLOGICAL MUSEUM, UNIVERSITETSPARKEN 15, DK-  
2100 COPENHAGEN 0, DENMARK

JESIONEK-SZYMANSKA, WANDA. ZAKLAD PALEOBIOLOGII PAN, AL.  
ZWIRKI I WIGURY 93, 02-089 WARSZAWA, POLAND

JONES, IRA. DEPARTMENT OF BIOLOGY, CALIFORNIA STATE  
UNIVERSITY, LONG BEACH, CA 90840, USA

JORDON, A. JOHN. 5688 51ST STREET, DELTA, B.C., V4K 3T7 CANADA

JOST, PETER M. KANT OBERFORSTAMT. ZURICH, KASPAR-ESCHER-  
HAURS, CH-8090 ZURICH, SWITZERLAND

JYO, TAKI. HOKKAIDO ABASHIRI FISHERIES, EXPERIMENTAL STATION,  
ABASHIRI, HOKKAIDO 099-31, JAPAN

KACZMARSKA, GRAZYNA. MOCCYNASIYO 5/25, 85-016 BYDGOSZCY,  
POLAND

KAMMER, THOMAS W. DEPT. OF GEOLOGY, WEST VIRGINIA UNIVERSITY,  
PO BOX 6300, MORGANTOWN, WV 26506-6300, USA

KANATANI, HARUO. DEPT DEVELOPMENTAL BIOLOGY, NAT. INST. FOR  
BASIC BIOLOGY, NYODAJI-CHO, OKAZAKI, 444, JAPAN

KAO, M.H. DEPARTMENT OF BIOLOGY, MEMORIAL UNIV. OF  
NEWFOUNDLAND, ST. JOHN'S, NEWFOUNDLAND, A1B 3X9 CANADA

KASYANOV, VLADIMIR L. INSTITUTE OF MARINE BIOLOGY,  
VLADIVOSTOK 690022. RUSSIA

KATSURA, SHIGERU. SCHOOL OF DENTISTRY, ORAL ANATOMY,  
TOKUSHIMA UNIVERSITY, TOKUSHIMA, JAPAN

KAWAMURA, KAZUHIRO. HOKKAIDO CENTRAL FISHERIES,  
EXPERIMENTAL STATION, YOICHI, HOKKAIDO, JAPAN

- KEEGAN, BRENDAN F. DEPARTMENT OF ZOOLOGY, UNIVERSITY COLLEGE, GALWAY, EIRE
- KELLER, BRIAN D. SMITHSONIAN TROP RESEARCH INST, UNIT 0948, APO AA, MIAMI, FL 34002-0948, USA
- KELLY, STUART M. 7224 17TH AVENUE, TAKOMA PARK, MD 20912, USA
- KELSO, DONALD. DEPARTMENT OF BIOLOGY, GEORGE MASON UNIVERSITY, 4400 UNIVERSITY DRIVE, FAIRFAX, VA 22030, USA
- KENDALL, B. PO BOX 17032, VIPINGO, VIA MOMBASA, KENYA
- KEOUGH, M.J. ZOOLOGY DEPARTMENT, UNIVERSITY OF MELBOURNE, ADELAIDE, S.A., AUSTRALIA
- KESLING, R.V. MUSEUM OF PALEONTOLOGY, UNIVERSITY OF MICHIGAN, ANN ARBOR, MI 48109, USA
- KIKUCHI, TAJI. AMAKUSA MARINE BIOLOGICAL LAB, KYUSHU UNIVERSITY, TOMIOKA, REIHOKU CHO, AMAKUSA, KUMAMOTO 863-25, JAPAN
- KLIKUSHIN, VLADIMIR. PETERSBURG PASLEONTOLOGICAL LAB, 26 LINE 9 (2), 199026 S. PETERSBURG, RUSSIA
- KLINGER, THOMAS SCOTT. DEPARTMENT OF BIOLOGY AND ALLIED HEALTH SCIENCES, BLOOMSBURG UNIVERSITY, BLOOMSBURG, PA 17815, USA
- KNOTT, K. EMILY. DEPT. OF BIOLOGICAL SCIENCES, UNIVERSITY OF ALABAMA, PO BOX 870344, TUSCALOOSA, AL 35487, USA
- KOBAYASHI, NAOMASA. BIOLOGICAL LABORATORY, DOSHISHA UNIVERSITY, KAMIKYO-KU, KYOTO 602, JAPAN
- KOGO, ICHIZO. TAGAWA PRIMARY SCHOOL, 9-37 TAGAWA 2 CHOME, YODOGAWA-KU, OSAKA 532, JAPAN
- KOJIMA, MANABU. DEPARTMENT OF BIOLOGY, TOYAMA UNIVERSITY, GOFUKU 3190, TOYAMA SHI/KEN 930, JAPAN
- KOLATA, DENNIS R. ILLINOIS STATE GEOLOGICAL SURV, NATURAL RESOURCES BUILDING, 615 E. PEABODY, CHAMPAIGN, IL 61820, USA

KOMATSU, MIEKO. DEPARTMENT OF BIOLOGY, FACULTY OF SCIENCE,  
TOYAMA UNIVERSITY, TOYAMA 930, JAPAN

KOSTER, MARION. ABTEILUNG MEERESZOOLOGIE, INSTITUT FUR  
MEERESKUNDE, DUSTERNBROOKER WEG, 2300 KIEL 1, GERMANY

KRISHNAN, MARY BAI. MARINE BIOLOGICAL STATION, ZOOLOGICAL  
SURVEY OF INDIA, 12, LEITHCASTLE STREET, MADRAS 28, INDIA

KRISHNARAJAH, PADMINI. UNIVERSITY OF JAFFNA, DEPARTMENT OF  
ZOOLOGY, JAFFNA, SRI LANKA

KRISTAN-TOLLMAN, EDITH M. K. SCHEIBENBERGSTRASSE 53, A-1180  
WIEN, AUSTRIA

KROPACH, CHAIM. IOLR, PO BOX 8030, HAIFA, ISRAEL

KUSHLINA, VERONIKA B. PALEONTOLOGICAL INSTITUTE,  
PROFISOJUZNAYA UL. 124, 1171868 GSP-7 V-321 MOSCOW, RUSSIA

LABARBARA, MICHAEL. DEPARTMENT OF ANATOMY, THE UNIVERSITY  
OF CHICAGO, 1025 EAST 57TH STREET, CHICAGO, IL 60637, USA

LAHAYE, MARIE-CHRISTINE. LABORATOIRE DE BIOLOGIE MARINE, C.P.  
160, UNIVERSITE LIBRE DE BRUXELLES, 50 MR F.D. ROOSEVELT, 1050  
BRUXELLES, BELGIUM

LAMBERT, PHILIP. BIOLOGY SECTION, ROYAL BRITISH COLUMBIA  
MUSEUM, 675 BELLEVILLE STREET, VICTORIA, B.C. CANADA

LANE, GARY N. GEOLOGY DEPARTMENT, INDIANA UNIVERSITY,  
BLOOMINGTON, IN 47405, USA

LARES, MICHAEL. DEPARTMENT OF BIOLOGY, UNIVERSITY OF SOUTH  
FLORIDA, TAMPA, FL 33620, USA

LARRAIN, ALBERTO P. DEPARTAMENTO DE BIOLOGIA, UNIVERSIDAD DE  
CONCEPCION, CONCEPCION, CHILE

LAUDON, LOWELL, GEOLOGY DEPARTMENT, UNIVERSITY OF WISCONSIN,  
MADISON, WI 53706, USA

LAVERACK, M.S. GATTY MARINE LAB, UNIVERSITY OF ST. ANDREWS,  
FIFE, SCOTLAND KY168L3, UK

LAWRENCE, JOHN M. DEPARTMENT OF BIOLOGY, UNIVERSITY OF SOUTH FLORIDA, TAMPA, FL 33620, USA

LAYOUS, YVES. 29 RUE PIERRE NICOLE, 75005 PARIS, FRANCE

LECLAIR, ELIZABETH. DEPT. GEOPHYSICAL SCIENCES, 5734 S. ELLIS AVE., CHICAGO, IL 60637, USA

LECLERC, MICHAEL. BIOLOGY DEPT., UER SCIENCES, ORLEANS UNIVERSITY, 45045 ORLEANS, FRANCE

LEELING, BEATRIX. ZOOLOGISCHES INSTITUT & MUSEUM, UNIVERSITAT HAMBURG, MARTIN LUTHER KING PLATZ 3, HAMBURG 13, GERMANY

LEIBSON, NINA L. INSTITUTE OF MARINE BIOLOGY, VLADIVOSTOK 690041, RUSSIA

LE MENN, JEAN. LAB. DE P & S DE PALEOZOIC, UNIV. DE BRATAGNE OCCIDENTALE, 6, AV LE GORGEU, 29275 BREST, FRANCE

LESSIOS, HARIS. SMITHSONIAN TROPICAL RESEARCH INSTITUTE, UNIT 0948, APO, MIAMI, FL 34002-0948, USA

LEVERONE, JAY. MOTE MARINE LABORATORY, 1600 CITY ISLAND PARK, SARASOTA, FL 34236, USA

LEVIN, VALERY S. KAMCHATKA DEPARTMENT OF MARINE, BIOLOGY AND BIOTECHNOLOGY, PO BOX 197, PETROPAVLOVSK-KAMCHATSKY 863000, RUSSIA

LEVINGS, C.D. PACIFIC ENVIRONMENT INSTITUTE, 4160 MARINE DRIVE, WEST VANCOUVER, B.C. V7V1N6, CANADA

LEVITAN, DONALD. DEPARTMENT OF ZOOLOGY, UNIVERSITY OF CALIFORNIA, DAVIS, CA 95616-8755, USA

LEVORSON, C.O. BOX 1, RICEVILLE, IA 50466, USA

LIAO, YULIN. ACADEMIA SINICA, INSTITUTE OF OCEANOLOGY, 7 NAN-HAI ROAD, QINGDAO, CHINA

LITVINOVA, NINA M. USSR ACADEMY OF SCIENCES, INSTITUTE OF OCEANOLOGY, 23 KRASIKOVA, MOSCOW V-218, RUSSIA 117218

LOPEZ-IBOR, ALICIA. CATERA DE ZOOLOGIA INVERTEBRADOS, UNIVERSIDAD COMPLUTENSE, MADRID 3, SPAIN

LOVELY, ERIC. DEPARTMENT OF ZOOLOGY, UNIVERSITY OF NEW HAMPSHIRE, DURHAM, NH 03824, USA

LUBCHENCO, JANE. DEPARTMENT OF ZOOLOGY, CORDLEY 3029, OREGON STATE UNIVERSITY, CORVALLIS, OR 97331-2914, USA

LUCAS, JOHN. SCHOOL OF BIOLOGICAL SCIENCES, JAMES COOK UNIVERSITY, TOWNSVILLE, QUEENSLAND 4811, AUSTRALIA

LUTZEN, JORGEN. INST OF COMPARATIVE ANATOMY, UNIVERSITY OF COPENHAGEN, UNIVERSITETSPARKEN 15, 2100 COPENHAGEN, DENMARK

MACZYNSKA, STEFANIA-ST. POLISH ACADEMY OF SCIENCES, MUSEUM OF THE EARTH, 20-26 AL. NA, SKARPIE, 00-488 WARSAW, POLAND

MADSEN, F. JENSENIUS. UNIV. ZOOLOGISKE MUSEUM, UNIVERSITETSPARKEN 15, DK-2100 COPENHAGEN, DENMARK

MAGNIEZ, PIERRE. LAB BIOL. INVERTEBRES MARINES, MUSEUM D'HISTOIRE NATURELLE, 55 RUE DE BUFFON, PARIS 75005, FRANCE

MAHARAVO, JEAN. CENTRE NATIONAL DE RECHERCHES D'OCEANOGRAPHIQUES, B.P. 68, (207) NOSY-BE, MADAGASCAR

MANCHENKO, GENNADY P. INSTITUTE OF MARINE BIOLOGY, VLADIVOSTOK 690022, RUSSIA

MARCOPOULOU-DIACANTONI, ANA. UNIVERSITY OF ATHENS, SUBFAC. SCIENCE OF EARTH, 93 MARATHONODROMOU STR., 15125 MAROUSSI, ATHENS, GREECE

MARCOS, CONCEPCION. DEPARTAMENTO AL ECOLOGIA, FACULDAD DE BIOLOGIA, UNIVERSIDAD DE MURCIA, APTDO 4.021 MURCIA, SPAIN

MARKOV, ALEXANDER V. PALEONTOLOGICAL INSTITUTE, PROFSOJUZNAYA UL. 123, 117868 MOSCOW, RUSSIA

MARQUES, V.M.A.M. DEPT ZOOLOGIA & ANTROPLIOGIA, FACULDADE CIENCIAS DE LISBOA, RUA DA ECOLA POLITECHNICA, 1200, LISBON, PORTUGAL

MARSH, LOISETTE M. WESTERN AUSTRALIAN MUSEUM, FRANCIS STREET, PERTH, WESTERN AUSTRALIA, 6000, AUSTRALIA

MARSHALL, CHARLES. DEPARTMENT OF EARTH AND PLANETARY SCIENCES, UNIVERSITY OF CALIFORNIA, LOS ANGELES, CA 90024, USA



MARTIN, R. ERIK. APPLIED BIOLOGY, INC. , PO BOX 974, JENSEN BEACH,  
FL 34958-0974, USA

MARTIN, RICHARD B. DIVISION OF FISHERIES, CSIRO MARINE  
LABORATORIES, GPP BOX 1538, HOBART, TASMANIA 7001, AUSTRALIA

MARTIN, RICHARD. CSIRO DIVISION LABORATORIES, GPP BOX 1538,  
HOBART, TASMANIA 7001, AUSTRALIA

MASACARENHAS, BERNARDO DE A. MUSEU NACIONAL, QUINTA DA BOA  
VISTA S/NO, SAO CRISTOVAO, RIO DE JANEIRO, BRAZIL

MASSIN, CLAUDE. INST ROYAL SCIENCES NATURELLES, DE BELGIQUE, 29  
RUE VAUTIER, 1040 BRUXELLES, BELGIUM

MATURO, FRANK JR. ZOOLOGY DEPARTMENT, UNIVERSITY OF  
FLORIDA, GAINESVILLE, FL 32611, USA

MAYTIA, LIC SUSANA. MUSEO NACIONAL DE HISTORIA NAT, CC399,  
MONTEVIDEO, URUGUAY

MCKENZIE, DOUGLAS. SCOTTISH MARINE BIOLOGICAL ASS, PO BOX 3,  
OBAN, ARGYLL, SCOTLAND PA34 4AD, UK

MCKNIGHT, DONALD. N.Z. OCEANOGRAPHIC INSTITUTE, PO BOX 14901,  
KILBIRNIE, WELLINGTON, NEW ZEALAND

MCMURRAY, SHEONA. GLASGOW POLYTECHNIC, COWCADDENS ROAD,  
GLASGOW G4 9BA, SCOTLAND, UK

MCNAMARA, KENNETH. WESTERN AUSTRALIAN MUSEUM, FRANCIS  
STREET PERTH 6000, WESTERN AUSTRALIA, AUSTRALIA

MCCARTHY, DANIEL. DEPARTMENT OF BIOLOGY, FLORIDA STATE  
UNIVERSITY, TALLAHASSEE, FL 32304, USA

MCCLINTOCK, JAMES B. DEPT. OF BIOLOGY, UNIVERSITY OF ALABAMA  
AT BIRMINGHAM, UAB STATION, BIRMINGHAM, AL 35294, USA

MCCONNAUGHEY, BAYARD H. 1653 FAIRMONT BLVD., EUGENE, OR  
97403, USA

MCEDWARD, LARRY. DEPARTMENT OF ZOOLOGY, 223 BARTRAM HALL,  
UNIVERSITY OF FLORIDA, GAINESVILLE, FL 32611, USA

MCINTOSH, GEORGE CLAY. ROCHESTER MUSEUM & SCIENCE CTR, 657 EAST AVE., PO BOX 1480, ROCHESTER, NY 14620, USA

MCKEMIE, CHARLES E. 1113 ZEBULON ROAD, GRIFFIN, GA 30223, USA

MCKINNEY, MICHAEL. DEPT. GEOLOGY, UNIVERSITY OF TENNESSEE, KNOXVILLE, TN 37966, USA

MEDEIROS-BERGEN, DOT. DEPARTMENT OF ZOOLOGY, UNIVERSITY OF NEW HAMPSHIRE, SPAULDING LIFE SCIENCE, DURIHAM, NH 03824, USA

MEIJER, LARURENT. CNRS, STATION BIOLOGIQUE, 29682 ROSCOFF CEDEX, FRANCE

MEIN, BIRGIT. ZOOL. INST. UND ZOOLOGISCHES, MUSEUM DER UNIVERSITÄT HAMBURG, MARTIN LUTHER KING PLATZ 3, 2000 HAMBURG 13, GERMANY

MENGE, BRUCE A. DEPARTMENT OF ZOOLOGY, OREGON STATE UNIVERSITY, CORVALLIS, OR 97331, USA

MESSING, CHARLES, NOVA UNIV OCEANOGRAPHIC CENTER, 8000 N. OCEAN DRIVE, DANIA, FL 33004, USA

MEYER, CHRISTIAN A. GEOLOGICAL INSTITUTE, BALTZERSTRASSE 1, CH-3012 BERNE, SWITZERLAND

MEYER, DAVID L. DEPARTMENT OF GEOLOGY, UNIVERSITY OF CINCINNATI, CINCINNATI, OH 45221, USA

MILLER, JOHN E. 2012 KENT ST. NE, PALM BAY, FL 32907, USA

MINTA, LEIGH W. OFFICE OF ACADEMIC PROGRAMS, WA 859, CALIFORNIA STATE UNIVERSITY, HAYWARD, CA 94542-3011, USA

MIRONOV, ALEXANDER. INSTITUTE OF OCEANOLOGY, ACADEMY OF SCIENCES, KRASIKOVA 23, MOSCOW 117218, RUSSIA

MITROVIC, JOVANKA P. FACULTY OF MINING & GEOLOGY, KAMENICKA 6, PO BOX 227, 11000 BEOGRAD, YUGOSLAVIA

MLADENOV, PHILIP V. DEPARTMENT OF MARINE SCIENCE, UNIVERSITY OF OTAGO, PO BOX 56, DUNEDIN, NEW ZEALAND

MOCRETSOVA, NINA D. PACIFIC RESEARCH INSTITUTE OF FISHERIES AND OCEANOGRAPHY 4, SHEVCHENKO ALLEY, 690600 VLADIVOSTOK, RUSSIA

MOOI, RICHARD. INVERTEBRATE ZOOLOGY, CALIFORNIA ACADEMY OF SCIENCES, GOLDEN GATE PARK, SAN FRANCISCO, CA 94118, USA

MOORE, ANDREW. GATTY MARINE LAB, EAST SANDS, ST. ANDREWS, FIFE, SCOTLAND, UK

MORISHITA, AKIRA. DEPARTMENT OF EARTH SCIENCES, NAGOYA UNIVERSITY, CHIKUSA, NAGOYA, JAPAN

MORRILL, JOHN. DIVISION OF NATURAL SCIENCES, NEW COLLEGE-USF, 5700 N. TRAIL, SARASOTA, FL 34243, USA

MOTOKAWA, TATSUO. FACULTY OF SCIENCE; BIOL. LAB., TOKYO INSTITUTE OF TECHNOLOGY, OKAYAMA, MEGURO-KU, TOKYO 152, JAPAN

MU, EN-ZHI. NANJING INSTITUTE OF GEOLOGY AND PALEONTOLOGY, ACADEMIA SINICA NANJING, PEOPLE'S REPUBLIC OF CHINA

MU, A T. INSTITUTE OF GEOLOGY AND PALEONTOLOGY, ACADEMIA SINICA NANJING, PEOPLE'S REPUBLIC OF CHINA

MUANR-BERNAT, JAIME. PLAZA MERCEDARIOS 4, C-07002 PALMA DE MALLORCA, BALEARIC ISLANDS, SPAIN

MUKAI, HIROSHI. AKKESHI MARINE BIOLOGICAL STA., HOKKAIDO UNIVERSITY AKKESHI, HOKKAIDO 088-11 JAPAN

MUNAR, JAIME. DEPT DE GEOLOGIA, UNIV PALMA DE MALLORCA, CARRETERA DE VALLDEMOSA KN7.5, MALLORCA, ISLAS BALAERES, SPAIN

MUNK, ERIC. ALASKS FISHERIES SCIENCE CENTR, KODIAK INVESTIGATIONS, PO BOX 1638, KODIAK, AK 99615-1638, USA

NAGAHAMA, YOSHITAKA. LAB OF REPRODUCTIVE BIOLOGY, NATIONAL INSTITUTE BASIC BIOL, OKAZAKI, 444 JAPAN

NAIDENKO, TAMARA K. INSTITUTE OF MARINE BIOLOGY, 6990032 VLADIVOSTOK, RUSSIA

NAKANO, EIZO. BIOSCIENCE RESEARCH, NIJIGACKA 1-7, MEITO, NAGOYA 465, JAPAN

NATEEWATNANA, ANUWAT. PHUKET MARINE BIOL. CENTER, PO BOX 60, PHUKET 83000, THAILAND

NAUEN, CORNELIA. INSTITUT FUR MEERESKUNDE, DUSTERNBROOKER WEG 20, 2300 KIEL 1, GERMANY

NEBELSICK, JAMES H. INSTITUTE AND MUSEUM OF GEOLOGY AND PALAEOLOGY, UNIVERSITY OF TUBINGEN, SIGWARSTSTR. 10, D-7400 TUBINGEN 1, GERMANY

NEILL, BRUCE. DEPARTMENT OF BIOLOGY, MONTANA STATE UNIVERSITY, BOZEMAN, MT 49715, USA

NEMOTO, SKIN-ICHI. TATEYAMA MARINE LABORATORY, OCHANOMIZU UNIVERSITY, KOH-YATSU, UMI-NO-HOSHI, TATEYAMA, CHIBA 294-03, JAPAN

NIESEN, THOMAS. DEPT. MARINE BIOLOGY, DIV. BIOL, SAN FRANCISCO STATE UNIVERSITY, 1600 HOLLOWAY AVE., SAN FRANCISCO, CA 94132, USA

NESTLER, H. GEOLOG. - PALEONTOL. INSTITUT, UNIVERSITAT GRIEFSWALD, FREIDRICH-LUDWIG-JAHR STR 17A, D-0-2200 GRIEFSWALD, GERMANY

NEUMANN, CHRISTIAN. FREIE UNIVERSITAT BERLIN, INSTITUT FUR PALAONTOLOGIE, SCHWENDENERSTRASSE 8, 1000 BERLIN 33, GERMANY

NICHOLS, DAVID. HATHGERLY LABORATORIES, UNIVERSITY OF EXETER, PRINCE OF WALES RD., EXETER EX4 4PS, ENGLAND

NISHIHIRA, MORITAKA. TOHOKU UNIVERSITY, FACULTY OF SCIENCE, BIOLOGICAL INSTITUTE, SENDAI, 980 JAPAN

NOJIMA, S. AITSU MARINE BIOLOGICAL LAB., KUMAMOTO UNIVERSITY, AITSU, MATSUSHIMA-CHO, AMAKUSA, KUMAMOTO-KEN 861-61, JAPAN

NORRIS, DANIEL R. MARINE LABORATORY, UNIVERSITY OF GUAM, UOG STATION, MANGILAO, GM 96923, USA

O'CONNOR, BRENDAN D.S. ZOOLOGY DEPARTMENT, UNIVERSITY COLLEGE, GALWAY, IRELAND

- O'HARA, TIMOTHY D. DEPT. OF INVERTEBRATE ZOOLOGY, MUSEUM OF VICTORIA, MELBOURNE 3000, VICTORIA, AUSTRALIA
- O'LOUGHLIN, P. MARK. 256 QUEENS PARADE, CLIFTON HILL 3068, AUSTRALIA
- O'NEILL, PATRICIA. DEPARTMENT OF ZOOLOGY, UNIVERSITY OF WEST AUSTRALIA, WESTERN AUSTRALIA 6009, AUSTRALIA
- OGURO, CHITARU. DEPARTMENT OF BIOLOGY, TOYAMA UNIVERSITY, TOYAMA 930, JAPAN
- OHTA, SUGURU. OCEAN RESEARCH INSTITUTE, UNIVERSITY OF TOKYO, MINAMIDAI 1-15-1, NAKANO, TOKYO 164, JAPAN
- OJEDA, F. PATRICIO. DEPARTAMENTO DE ECOLOGIA, UNIVERSIDADE CATOLICA, CASILLA 114-D, SANTIAGO, CHILE
- OJI, TATSUO. GEOLOGICAL INSTITUTE, UNIVERSITY OF TOKYO, HONGO, TOKYO 113, JAPAN
- OKADA, MINORU. SCIENCE EDUCATION INSTITUTE, OSAKA PREF, KARITA 4-CHOME, SUMIYOSHI-KU, OSAKA 558, JAPAN
- OKAMOTO, KYOKO. GEOLOGICAL INSTITUTE, UNIVERSITY OF TOKYO, HONGO, TOKYO 113, JAPAN
- OKAZAKI, KAYO. DEPARTMENT OF BIOLOGY, TOKYO METROPOLITAN UNIVERSITY, FUKAZAWA 2-1-1, SETAGAYA-KU, TOKYO 158, JAPAN
- OLAVE, SERGIO. INSTITUTO DE FOMENTO PESQUERO, CASILLA 78, CASTRO, CHILE
- OLVER, JANE. LANCE'S COTTAGE, PARKGATE ROAD, NEWDIGATE, SURREY RH5 5DY, ENGLAND
- ORLER, PARICIA MABEL. CADIC CC.92 USHUAIA, TIERRA DEL FUEGO, ARGENTINA
- OTSU, INES. INST INVESTIGACIONES OCEANOLOG, UNIVERSIDAD DE CHILE, CASILLA 1240 ANTOFAGASTA, CHILE
- OUDEJANS, R.C.H.M. LAB CHEMICAL ANIMAL PHYSIOLOGY, THE STATE UNIVERSITY UTRECHT, PADUALAAN 8, 3508 TB UTRECHT, NETHERLANDS

- PABIAN, ROGER K. CONSERVATION & SURVEY DIVISION, UNIVERSITY OF NEBRASKA, LINCOLN, NE 68588, USA
- PAGETT, RICHARD MICHAEL. HUNTERS BROOK HOUSE, HOGGS LANE, PURTON WILSHIRE, ENGLAND SN5 9HQ, UK
- PAINE, ROBERT T. DEPARTMENT OF ZOOLOGY, UNIVERSITY OF WASHINGTON, SEATTLE, WA 98195, USA
- PALLER, MARC S. 301 NOTT ST M, SUITE 100, SCHENECTADY, NY 12305, USA
- PAPE-LINDSTROM, PAMELA. UNIVERSITY OF SOUTH CAROLINA, BIOLOGY DEPARTMENT, COLUMBIA, SC 29210, USA
- PARSLEY, RONALD. DEPARTMENT OF GEOLOGY, TULANE UNIVERSITY, NEW ORLEANS, LA 70118, USA
- PASSAMENTE, JOSEPH. MARINE SCIENCE INSTITUTE, UNIVERSITY OF PHILIPPINES, PO BOX 1, DILIMAN, QUEZON CITY, PHILIPPINES
- PASTOR, XAVIER. LAB OCEANOGRAFICODE BALEARES, MUELLE DE PELAIRES S/N, PALMA DE MALLORCA, BALEARIC ISLANDS, SPAIN
- PAUL, C.R.C. DEPARTMENT OF GEOLOGY, LIVERPOOL UNIVERSITY, LIVERPOOL, L69 3BX, ENGLAND, UK
- PAULS, SHEILA M. INSTITUTO DE ZOOLOGIA TROPICAL, UNIVERSIDAD CENTRAL VENEZUELA, APARTADO 47058, CARACAS, 1041-A, VENEZUELA
- PAWSON, DAVID L. ROOM W323, MAIL STOP 163, NAT. MUSEUM OF NATURAL HISTORY, SMITHSONIAN INSTITUTION, WASHINGTON, DC 20560
- PAYOT, FLORENCE. INSTITUTE OF BIOLOGY, ODENSE UNIVERSITY, CAMPUS VEJ 55, DK 5230 ODENSE, DENMARK
- PEARSE, JOHN S. INSTITUTE OF MARINE SCIENCES, UNIVERSITY OF CALIFORNIA, SANTA CRUZ, CA 95064, USA
- PEDROTTI, M.L. STATION ZOOLOGIQUE, QUAI DE LA DARSE, F-06230 VILLEFRANCE-SUR-MER, BP.28-06230 FRANCE
- PENCHASZADEH, PABLO. INTECMAR, UNIV. SIMON BOLIVAR, APARTADO 80659, CARACAS, VENEZUELA

PENNINGTON, J. TIMOTHY. KEWALO MARINE LAB, 41 AHUI STREET,  
HONOLULU, HI 96813, USA

PENTREATH, VICTOR W. BIOLOGICAL SCIENCES, UNIVERSITY OF  
SALFORD, SALFORD M5 4WT, ENGLAND, UK

PETR, VACLAV. GEOLOGICAL LIBRARY, CHARLES UNIVERSITY,  
ALBERTOV 6, 128 43 PRAHA 2, CZECH REPUBLIC

PHELAN, THOMAS. 122 WINNEBAGO STREET, WALLA WALLA, WA 99362,  
USA

PHILIPPE, MICHAEL. MUSEE GUIMET D'HIST NATURELLE, 28 BD DES  
BELGES, 69006 LYON, FRANCE

PINELA, JEAN E. INSTITUTO DE ZOOLOGIA, UNIVERSIDAD AUSTRAL DE  
CHILE, CASILLA 567, VALDIVIA, CHILE

POLSON, EMMA. DEPARTMENT OF BIOLOGY, UNIVERSITY OF SOUTH  
FLORIDA, TAMPA, FL 33620, USA

POMORY, CHRISTOPHER. DEPARTMENT OF BIOLOGY, UNIVERSITY OF  
SOUTH FLORIDA, TAMPA, FL 33620, USA

PORETZKAJA, E.S. V-34, 1 LINE, 10 APRTM. 21, LENINGRAD, RUSSIA 199034

PRESTEDGE, GEOFFREY. 16 GEEVES CRESCENT, MIDWAY POINT,  
TASMANIA 7171, AUSTRALIA

PRINCE, JANE. DEPARTMENT OF ZOOLOGY, UNIV OF WESTERN  
AUSTRALIA, NEDLANDS, WESTERN AUSTRALIA 6009, AUSTRALIA

PROFANT, ROBERT. DIV OF LIFE SCIENCE, SANTA BARBARA CITY  
COLLEGE, 721 CLIFF DR., SANTA BARBARA, CA 93109, USA

PROKOP, RUDOLF JAN. NATIONAL MUSEUM, PRAHA 1, VACLAVSKE  
NAMESTI, 115 79 PRAHA 1, CZECH REPUBLIC

PROPP, M. INSTITUTE OF MARINE BIOLOGY, VLADIVOSTOK 690041,  
RUSSIA

RAHAMAN, ABDUL ABDUL. DEPARTMENT OF ZOOLOGY, SRI VASAVI  
COLLOEGE, ERODE-G38 31B, INDIA

RAJA, P. VIVEK. DEPARTMENT OF BIOLOGY, MADRAS MEDICAL  
COLLEGE, MADRAS 600 003, INDIA

RAYMOND, ALAN M. GATTY MARINE LAB, UNIVERSITY OF ST.  
ANDREWS, ST. ANDREWS, FIFE KY168LB, SCOTLAND

REGIS, MARIE-BERTHE. CERAM-CASE 3UI, FACUL SCI & TECH DE ST.  
JEROME, 13397, MARSEILLE CEDEX 13, FRANCE

REGNELL, GERHARD. GEOLOGISKA INSTITUTIONEN, SOLVEGATAN 13, S-  
223 62 LUND, SWEDEN

RENAULT, SERGE. MUSEUM D'HISTOIRE NATURELLE, 12 RUE VOLTAIRE,  
44000 NANTES, FRANCE

RHO, BOON JO. NATURAL HISTORY MUSEUM, EWHA WOMEN'S  
UNIVERSITY, SEOUL, KOREA 120

RIDDLE, STEVEN. THE OHIO STATE UNIVERSITY, 125 SOUTH OVAL MALL,  
COLUMBUS, OH 43210, USA

ROBERTS, DAI. SCHOOL OF BIOLOGY AND BIOCHEM., QUEEN'S  
UNIVERSITY BELFAST, NORTHERN IRELAND BT9 7BL, UK

ROBISON, RICHARD. DEPARTMENT OF GEOLOGY, UNIVERSITY OF  
KANSAS, LAWRENCE, KS 66045, USA

RODRIGUEZ, SEBASTIAN. DEPT. BIOL. CELULAR Y MOLEC., FACULTAD  
DE CIENCIAS BIOLOGIA, UNIVERSIDAD CATOLICA DE CHILE, CASILLA  
114-D, SANTIAGO, CHILE

ROMINAS-CASTRO, LILY. NATIONAL INSTITUTE FISHERIES, PO BOX 1306,  
ENSENAD, BAJA CALIFORNIA, MEXICO

ROSE, EDWARD P.F. DEPARTMENT OF GEOLOGY, ROYAL HOLLOWAY &  
BEDFORD NEW COL, UNIVERSITY OF LONDON, EGHAM SURREY TW20  
OEX, ENGLAND, UK

ROTMAN, HELEN CLARK. NATURAL HISTORY UNIT, NATIONAL MUSEUM  
OF N.Z., PO BOX 467, WELLINGTON, NEW ZEALAND

ROUX, MICHEL. UNIVERSITE DE REIMS, LABORATOIRE DES SCIENCES  
DE LA TERRE, B.P. 347, 51062 REIMS CEDEX, FRANCE

ROWE, FRANK. GOLDBROOK BOARDING KENNELS, NUTTERY VALE,  
CROSS STREET, HOXNE, SUFFOLK IP21 5EB, ENGLAND, UK

ROZHNOV, S.V. PALEONTOLOGICAL INSTITUTE, RUSSIAN ACADEMY OF  
SCIENCES, PROFSOYUSNAYA STR. 123, 117647 MOSCOW, RUSSIA



RUMRILL, STEVEN S. SOUTH SLOUGH NATIONAL ESTUARINE RESERVE,  
PO BOX 5412, CHARLESTON, OR 97420, USA

RUZAFÁ PEREZ, ANGEL. DEPARTAMENTO AL ECOLOGIA, FACULTAD DE  
BIOLOGIA, UNIVERSIDAD DE MURCIA, APTDO 4.021, MURCIA, SPAIN

RYABUSHKO, VITALY. DEPARTMENT ANIMAL PHYSIOLOGY, INSTITUTE  
OF BIOLOGY OF THE SOUTHERN SEAS, 2 NAKHIMOV AVENUE,  
SEVASTOPOL 335011, UKRAINE

SALVAT, MARIANA BEATRIZ. CIENCIAS EXACTAS Y NATURALES,  
CIUDAD UNIVERSITARIA, PAB II 4 PISO, IY28 BUENOS AIRES, ARGENTINA

SANCHEZ, PATRICIO. FACULTAD DE CIENC. BIOLÓGICAS,  
UNIVERSIDADE CATOLICA, CASILLA 114-D, SANTIAGO, CHILE

SASTRY, DWDASI R.K. FPS BUILDING, ZOOLOGICAL SURVEY OF INDIA, 27  
JAWAHARLAL NEHRU ROAD, CALCUTTA 700016, INDIA

SATO, HIDEKI. FACULTY OF SOCIAL SCIENCE, NAGANO UNIVERSITY,  
SHIMONOGO, UEDA, NAGANO 386-12, JAPAN

SAUER DE AVILA PIRES, TERESA C. DEPTO DE ZOOLOGICA/UFRJ,  
CIUDADE UNIVERSITARIA, ILHA DO FUNDÃO, CCS-SALA A1-117, RIO DE  
JANEIRO, BRAZIL 21.941

SCALLY, KEVIN. HOSPITAL DENTAL SERVICES, CANTERBURY AREA  
HEALTH BOARD, PRIVATE BAG 4710 CHRISTCHURCH, NEW ZEALAND

SCHELTEMA, RUDOLF S. DEPARTMENT OF BIOLOGY, WOODS HOLE  
OCEANOGRAPHIC INST., WOODS HOLE, MA 02543, USA

SCHINNER, GOTTFRIED. INSTITUTE OF ZOOLOGY, UNIVERSITY OF  
VIENNA, A-1090 VIENNA, AUSTRIA

SCHMINCKE, SABINE. PALAEOONTOLOGISCHES INSTITUT, UNIV.  
WURZBURG, PLEICHERWALL 1, 8700 WURZBURG, GERMANY

SCHOPPE, SABINE. INST ALLGEMEINE SPEZ. ZOOLOGIE, JUSTUS-LIEBIG  
UNIVERSITÄT, D-6300 GIESSEN, GERMANY

SCHUETZ, ALLEN W. SCH HYGIENE & PUBLIC HEALTH, JOHNS HOPKINS  
UNIVERSITY, 615 N. WOLFE ST., BALTIMORE, MD 21205, USA

SCHUMACHER, HELMUT. UNIVERSITÄT ESSEN, INST. FÜR ÖKOLOGIE,  
POSTFACH 10 37 64, D4300 ESSEN 1, GERMANY

SCURRY, D.G.A. DEPT OF GEOLOGY & PHYSICAL SCI, OXFORD  
POLYTECHNIC, OXFORD OX3 OBP, ENGLAND

SERAFY, D. KEITH. DEPARTMENT OF BIOLOGY, SOUTHAMPTON  
COLLEGE, SOUTHAMPTON, NY 11968, USA

SEVASTOPULO, GEORGE. DEPT OF GEOLOGY, TRINITY COLLEGE,  
UNIVERSITY OF DUBLIN, DUBLIN 2, IRELAND

SHEPHERD, SCORESBY. S.A. RESEARCH AND DEV. INSTT., BOX 1625  
G.P.O., ADELAIDE, S.A. 5001, AUSTRALIA

SHICK, MALCOLM. DEPT. OF ZOOLOGY, UNIVERSITY OF MAINE, 5751  
MURRAY HALL, ORONO, ME 04469-5751, USA

SHIRAI, HIROKO. LAB OF REPRODUCTIVE BIOLOGY, NATIONAL INST  
FOR BASIC BIOLOG, OKAZAZI, AICHI-KEN 444, JAPAN

SHIRLEY, THOMAS C. SCHOOL OF FISHERIES, UNIVERSITY OF ALASKA,  
11120 GLACIER HWY., JUNEAU, AK 99801, USA

SIBUET, MYRIAM. IFREMER CENTRE DE BREST, BP 70, 29263 PLOUZANE,  
FRANCE

SIEGEL, I. MYRA. RFD 5 HANNAFORD COVE, CAPE ELIZABETH, ME 04107

SILVA DA HORA, MARIA MARGARETH. MESEU NACIONAL DA UNIV  
FEDERAL, QUSNTA BOA VISTA-SAO CRISTOVAE, RIO DE JANEIRO 20.940,  
BRAZIL

SIMMS, MICHAEL J. DEPARTMENT OF GEOLOGY, NATIONAL MUSEUM OF  
WALES, CATHAYS PARK, CARDIFF, WALES CF1 3NP, ENGLAND, UK

SIMPSON, RODNEY D. DEPARTMENT OF ZOOLOGY, UNIVERSITY OF NEW  
ENGLAND, ARMIDALE, NSW 2351, AUSTRALIA

SINGLETARY, ROBERT. L. BIOLOGY DEPARTMENT, UNIVERSITY OF  
BRIDGEPORT, BRIDGEPORT, CT 0661, USA

SLAMOVA, RADANA. STAVEBNI GEOLOGIE N.P., POD RAPIDEM 8,  
STRASNICE, PRAHA 10, CZECHOSLOVAKIA

SMILEY, SCOTT. INSTITUTE OF ARCTIC BIOLOGY, UNIVERSITY OF  
ALASKA, FAIRBANKS, AK 99775, USA

SMIRNOV, I.S. ZOOLOGICAL INSTITUTE, ACADEMY OF SCIENCES,  
UNIVERSITETSKAYA NAB., I, ST. PETERSBURG 199034, RUSSIA

SMIRNOV, ALEXEY V. ZOOLOGICAL INSTITUTE, ACADEMY OF SCIENCES,  
UNIVERSITETS NAB., I, ST. PETERSBURG 199034, RUSSIA

SMITH, ANDREW. DEPARTMENT OF PALAEOLOGY, NATURAL  
HISTORY MUSEUM, CROMWELL ROAD, LONDON SW7 5BD, ENGLAND, UK

SOLOVJEV, ANDREY N. PALEONTOLOGICAL INSTITUTE, RUSSIAN  
ACADEMY OF SCIENCES, PROFSOYUSNAYA STR., 123, 117647 MOSCOW,  
RUSSIA

SOUTH, G. ROBIN. INSTITUTE OF MARINE RESOURCES, UNIVERSITY OF  
THE SOUTH PACIFIC, PO BOX 1168, SUVA, FIJI

SPENCER, LARRY T. NATURAL SCIENCE DEPARTMENT, PLYMOUTH  
STATE COLLEGE, PLYMOUTH, NH 03264, USA

SPRINKLE, JAMES. DEPT OF GEOLOGICAL SCIENCES, UNIVERSITY OF  
TEXAS, AUSTIN, TX 78712, USA

STANCYK, STEPHEN E. BELLE W. BARUCH INSTITUTE, UNIVERSITY OF  
SOUTH CAROLINA, COLUMBIA, SC 29208, USA

STEELE, D.H. DEPARTMENT OF BIOLOGY, MEMORIAL UNIVERSITY OF  
NEWFOUNDLAND, ST. JOHN'S, NEWFOUNDLAND A1C 5S7, CANADA

STENKIL, M. NATURAL HISTORY DEPARTMENT, MALMO MUSEUM,  
MALMOHUSVAGEN, S21120, MALMO, SWEDEN

STEPHENSON, DAVID JEFFREY. DEPARTMENT OF GEOLOGY,  
UNIVERSITY OF KEELE, KEELE, STAFFORDSHIRE, ST5 5BG, UK

STICKLE, WILLIAM B. JR. ZOOLOGY & PHYSIOLOGY DEPT., LOUISIANA  
STATE UNIVERSITY, BATON ROUGE, LA 70803, USA

STOKES, ROBERT B. SCHOOL OF GEOLOGICAL SCIENCES, KINGSTON  
UNIVERSITY, M KINGSTON UPON THAMES, SURREY, KT1 2EE, ENGLAND,  
UK

STRATHMANN, RICHARD R. FRIDAY HARBOR LABORATORIES, 620  
UNIVERSITY ROAD, FRIDAY HARBOR, WA 98250, USA

STUBBS, T. GATTY MARINE LAB, UNIVERSITY OF ST. ANDREWS, FIFE,  
SCOTLAND KY16 8L3, UK

STUMP, R.J.W. ZOOLOGY DEPARTMENT, JAMES COOK UNIVERSITY, PO JCUNQ, TOWNSVILLE, QUEENSLAND 4811, AUSTRALIA

SUGIYAMA, MASAO. MARINE BIOLOGICAL STATION, NAGOYA UNIVERSITY, SUGASHIMA-CHO, TOBA-SHI, MIE-KEN 517, JAPAN

SUMRALL, COLIN D. DEPARTMENT OF GEOLOGICAL SCI., UNIVERSITY OF TEXAS, AUSTIN, TX 78712, USA

SUTER, SHERMAN J. COMMITTEE ON EVOLUTIONARY BIOL, C/O DEPT. GEOPHYSICAL SCIENCES, UNIVERSITY OF CHICAGO, CHICAGO, IL 60637, USA

SUZUKI, NORIO. NOTO MARINE LABORATORY, KANAZAWA UNIVERSITY, OGI, UCHIURA, ISHIKAWA 927-05, JAPAN

TABLADO, ALEJANDRO. MUS. ARGENTINA CS. NATURALES, AVDA. ANGEL GALLARDO 470, 1405 BUENOS AIRES, ARGENTINA

TAHARA, YUTAKA. DEPARTMENT OF BIOLOGY, OSAKA KYOIKU UNIVERSITY, TENNOJI, OSAKA 543K, JAPAN

TAHERA, QASEEM. MARINE REFERENCE COLLECT. CTR., UNIVERSITY OF KARACHI, CODE NO. 75270, KARACHI-32, PAKISTAN

TAJIKA, KEN-ICHI. DEPARTMENT OF BIOLOGY, NIHON UNIVERSITY, SCHOOL OF MEDICINE, OYAGUCHI, ITABASHI, TOKYO 173, JAPAN

TAKAHASHI, KEIICHI. DEPARTMENT OF BIOLOGY, INTERNATIONAL CHRISTIAN UNIVERSITY, 3-10-2 OSAWA, MITAKA, TOKYO 181, JAPAN

TAKI, JYO. HOKKAIDO ABASHIRI, FISHERIES EXPERIMENTAL STATION, ABASHIRI, HOKKAIDO, 099-31, JAPAN

TEGNER, MIA. DEPARTMENT OF ZOOLOGY, PKO A001, SCRIPPS INST OF OCEANOGRAPHY, LA JOLLA, CA 92093-0201, USA

TELFORD, MALCOLM. DEPARTMENT OF ZOOLOGY, UNIVERSITY OF TORONTO, ONTARIO M5S 1A1, CANADA

TERRY, RICHARD E. DEPARTMENT OF GEOLOGY, UNIVERSITY OF CINCINNATI, CINCINNATI, OH 45221, USA

THANDAR, A.S. DEPARTMENT OF ZOOLOGY, UNIVERSITY OF DURBAN-WESTVILLE, P/BAG X54001, DURBAN 4000, REPUBLIC OF SOUTH AFRICA

THIERRY, JACQUES. UNIVERSITE DE BOURGOGNE, CENTRE DES SCIENCES LA TERRE, 6 BD-GABRIEL, 21100 DIJON, FRANCE

THIES, JENNIFER. 7942 EMERALD BLUFF COURT, HOUSTON, TX 77095-4415, USA

THOMASSIN, BERNARD. CENT. D'OCEANOGRAPHIE MARSEILLE, STATION MARINE D'ENDOUME, H TRAVERSE LA BATTERIE LIONS, 13007 MARSEILLE, FRANCE

THOMPSON, GEOFFREY B. STATE POLLUTION CONTROL COMMIS, 157 LIVERPOOL ST., SYDNEY 2000, NSW, AUSTRALIA

TOMMASI, LUIZ ROBERTO. INSTITUTO OCEANOGRAFICO, UNIVERSIDAD DE SAO PAULO, CAIXA POSTAL NO 9075, SAO PAULO, S.P., BRAZIL

TRAERE, KURT. LEHRKANZEL FUR MEERESBIOLOGIE, UNIVERSITAT WIEN, WAHRINGERSTRASSE 17, 1090 AUSTRIA

TRONCOSO, J FRANKLIN F. INVESTIGADOR BIOLOGIA, MUSEO DE CONCEPCION, CASILLA 1054, EDMUNDO LARENAS 420, CONCEPCION, CHILE

TURNER, RICHARD L. DEPT. OF BIOLOGICAL SCIENCES, FLORIDA INST. OF TECHNOLOGY, 150 W. UNIVERSITY BLVD., MELBOURNE, FL 32901, USA

TYLER, PAUL. DEPARTMENT OF OCEANOGRAPHY, THE UNIVERSITY, SOUTHAMPTON, ENGLAND S09 5NH, UK

UBAGHS, GEORGES. AVENUE BOIS LE COMTE 28, 4140 SPRIMONT, BELGIUM

URBINA, MEDARO BURGOS. INSTITUTO CENTRAL DE BIOLOGIA, UNIVERSIDAD DE CONCEPCION, CONCEPCION, CHILE

VADAS, ROBERT L. DEPARTMENT OF PLANT BIOLOGY, UNIVERSITY OF MAINE, ORONO, ME 04469-5722, USA

VADET, ALAIN. 67 BLVD FURVINY, 62200 BOULOGNE SUR MER, FRANCE

VADON, CATHERINE. MUSEUM NATIONAL D'HISTOIRE NATURELLE, LAB. DE BIOLOGIE DES INVERTS., 55 RUE DE BUFFON, F-75005 PARIS, FRANCE

VAIL, LYLE. LIZARD ISLAND RESEARCH STATION, PMB 37, CAIRNS, QUEENSLAND 4871, AUSTRALIA

VALENTINE, JOHN. UNIVERSITY OF ALABAMA, DAUPHIN ISLAND SEA LAB, BOX 369-370, DAUPHIN ISLAND, AL 36528, USA

VANDENSPIEGEL, DIDIER. LAB DE BIOLOGIE MARINE, UNIVERSITE DE MONS, 19 AVE MAISTRIAU (BAT 4), 7000-MONS, BELGIUM

VARAKSINA, GALINA S. LABORATORY OF REGULATION AND REPRODUCTION, INSTITUTE OF MARINE BIOLOGY, VLADIVOSTOK 690032, RUSSIA

VASKOVSKY, V. INSTITUTE OF MARINE BIOLOGY, VLADIVOSTOK 690022, RUSSIA

VEGA, JUAN TORRES. LABORATORIO ECOL. EQUINODERMOS, INST. CIENC. DEL MAR LIMNOLOG., UNIVERSIDAD NACIONAL AUTONOMA, APARTADO POSTAL 70-305, CP 04510, MEXICO DF.

VEINUS, JULIA. DEPARTMENT OF GEOLOGY, SYRACUSE UNIVERSITY, SYRACUSE, NY 13210, USA

VELARDE, RONALD. CITY OF SAN DIEGO, MARINE BIOLOGY LAB. M.S. 45, 4077 NORTH HARBOR DR., SAN DIEGO, CA 92101, USA

VELAQUE, MARC. LABORATORIE D'E.B.V.M., UNIV. DES SCIENCES DE LUMINY, CASE 901-70 RT LEON LACHAMP, 13288 MARSEILLE CEDEX 9, FRANCE

VENTURA, C.R.R. DEPTO. DE BIOLOGIA MARINHA, UNIVERSIDAD FEDERAL DO R.J., BLOCO A, C.C.S. ILHA DO FUNDAO, CIDADE UNIVERSITARIA, RIO DE JANEIRO, RJ 21941, BRAZIL

VIADA, STEPHEN. 1410 CAVITT, LGL ECOLOGICAL RESEARCH, BRYAN, TX 77801, USA

VIKTOROVSKAYA, GALINA I. PACIFIC RESEARCH INSTITUTE OF FISHERIES AND OCEANOGRAPHY, LABORATORY OF PHYSIOLOGY, 4, SHEVCHENKO ALLEY, 690600 VLADIVOSTOK, RUSSIA

VIVIANI, CARLOS A. UNIVERSIDAD DE LA SERENA, CAMPUS ENRIQUE MOLINA, LA SERENA, CHILE

VOOGT, PETER A. CHEMICAL ANIMAL PHYSIOLOGY, UNIVERSITY OF UTRECHT, 5 PRINCETON PLEIN, 3584 CC UTRECHT, THE NETHERLANDS

VOSS, JOEACHIM. INSTITUT FUR POLAROKOLOGIE, CHRISTIAN ALBRECHTS UNIV KIEL, 2300 KIEL, GERMANY

WALENKAMP, J.H.C. RIJSMUSEUM VAN NAT. HISTORIE, RAAMSTEEG 3,  
LEIDEN 2311 PL, THE NETHERLANDS

WALKER, CHARLES W. DEPARTMENT OF ZOOLOGY, UNIVERSITY OF  
NEW HAMPSHIRE, DURHAM, NH 03824, USA

WAREN, ANDERS. BOX 50007, S-10405, STOCKHOLM, SWEDEN

WATERS, JOHNNY A. WEST GEORGIA COLLEGE, DEPARTMENT OF  
GEOLOGY, CARROLLTON, GA 30118, USA

WATTS, STEPHEN A. DEPARTMENT OF BIOLOGY, UNIV OF ALABAMA AT  
BIRMINGHAM, UAB STATION, BIRMINGHAM, AL 35294, USA

WEBER, WALTER. ZOOL. INSTITUTE, DEPT ANIMAL PHYSIOLOGY,  
UNIVERSITY OF COLOGNE, 5 COLOGNE 41, WEYERTAL 119, GERMANY

WEBSTER, GARY D. DEPARTMENT OF GEOLOGY, WASHINGTON STATE  
UNIVERSITY, PULLMAN, WA 99164-2812, USA

WEST, RON. DEPARTMENT OF GEOLOGY, THOMPSON HALL, KANSAS  
STATE UNIVERSITY, MANHATTAN, KS 66506, USA

WESTHEAD, STANLEY. "CROSSAL", 7 CLAREMONT DR., CLITHEROE,  
LANCASHIRE BB7 1JW, ENGLAND, UK

WIBLE, JOSEPH G. LIBRARY, BOX 398, CATALINA MARINE SCIENCE  
CENTER, AVALON, CA 90704, USA

WIEDMAN, LAWRENCE. DEPARTMENT OF GEOLOGY, MONMOUTH  
COLLEGE, MONMOUTH, IL 61462, USA

WILKIE, IAIN C. DEPT. OF BIOLOGICAL SCIENCES, GLASGOW  
POLYTECHNIC, 70 COWCADDENS ROAD, GLASGOW G4 OBA, SCOTLAND,  
UK

WILLCOX, MARK. DUNSTAFFNAGE MARINE LABORATORY, PO BOX 3,  
OBAN, ARGYLL PA34 4AD, SCOTLAND

WITZKE, BRIAN J. IOWA DEPT OF NATURAL RESOURCES, GEOLOGICAL  
SURVEY BUREAU, IOWA CITY, IA 52242, USA

WOODLEY, JEREMY D. DISCOVERY BAY MARINE LABORATORY, PO BOX  
35, DISCOVERY BAY, JAMAICA, WEST INDIES

WORHEIDE, GERT. FREIE UNIVERSITAT BERLIN, INSTITUT FUR  
PALAONTOLOGIE, MALTESER-STR 74-100, HAUS D, D 1000 BERLIN 46,  
GERMANY

WRIGHT, C.W. OLD RECTORY, SEABOROUGH, BEAMINSTER, DORSET DT8  
304, ENGLAND, UK

YAKOLEV, SERGHEY N. INSTITUTE OF MARINE BIOLOGY, 690032  
VLADIVOSTOK, RUSSIA

YAKOVLEV, YURI. INSTITUTE OF MARINE BIOLOGY, 690032  
VLADIVOSTOK, RUSSIA

YAMAGUCHI, MASASHI. DEPARTMENT OF MARINE SCIENCES,  
UNIVERSITY OF THE RYUKYUS, NISHIHARA, OKINAWA, 903-01, JAPAN

YANAGISAWA, TOMIO. SAITAMA MEDICAL SCHOOL JUNIOR COLLEGE,  
MOROYAMA, IRUMA-GUN, SAITAMA 350-04, JAPAN

YOSHIAKI, ISHIDA. 4-21-2 KAMIIGUSA, SUGINAMI-KU, TOKYO, JAPAN 167

YOSHIDA, MASAO. USHIMADO MARINE LABORATORY, 130-17 KASHINO,  
USHIMADO, OKAYAMA 7-1-43, JAPAN

YOSHIKATO, KATSUTOSHI. DEPARTMENT OF BIOLOGY, TOKYO  
METROPOLITAN UNIVERSITY, SETAGAYA-KU, TOKYO, JAPAN

YOUNG, CRAIG. HARBOR BRANCH OCEANOGRAPHIC INSTITUTION, INC.  
5600 OLD DIXIE HIGHWAY, FORT PIERCE, FL 34946, USA

ZAGHBIB-TURKI, D. DEPT. SCIENCES DE LA TERRE, FACULTE DES  
SCIENCES DE TUNIS, CAMPUS UNIVERSITAIRE, 1060 TUNIS, TUNISIA

ZAVODNIK, DUSAN. CENTER MARINE RESEARCH ROVINJ, "RUDJER  
BOSKOVIC" INSTITUTE, 52210 ROVINJ, CROATIA

ZEIDLER, WOLFGANG. SOUTH AUSTRALIAN MUSEUM, NORTH TERRACE,  
ADELAIDE, SOUTH AUSTRALIA 5000, AUSTRALIA

ZIESENHENNE, FRED C. RR#1 BOX 497-A, KULA, MAUI, HI 96790, USA

ZMARZLY, DEBORAH. CITY OF SAN DIEGO, OCEAN MONITORING  
PROGRAM MS-45, 4077 NORTH HARBOR DRIVE MS-45A, SAN DIEGO, CA  
92101, USA



**TELEPHONE - FAX - E-MAIL**

Individuals who desire to be listed in the next newsletter are requested to send their numbers to the editor.

Addresses of other echinoderm workers will be most welcome.

A network for individuals with e-mail addresses has been set up by

Win Hide  
Department of Biochemistry  
University of Houston  
Houston, TX

The Echinoderm Newsletter is distributed through the network.

Hide edits STARNET, an e-mail newsletter for echinoderm biologists.  
e-mail: whide@matrix.BCHS.UH.EDU

e-mail	Telephone	FAX
Arnaud, P.	91 52.12.94	
Ausich, W. ausich.1@osu.edu	614-422-2721	
Baker, A.	4 859-609	4 857-157
Banks, L. milnes_p@darwin.ntu.edu.au		
Barker, M. mfbarker@otago.ac.nz.	3-479-7584	3 34781825
Bartsch, I.	40 89693213	49 40 89693212
Basch, L. lbasch@cats.ucsc.edu	408 459-4026	408 459-4882
Beck, G. gbeck@path.som.sunysb.edu		
Berger, J.		416 978-3521
Birenheide, R. tmotokaw@cc.titech.ac.jp	3 3726-1111	3 3748-3017
Birkeland, C. chaz@uog.pacific.edu	671 734-2421	671 734-6767
Black, R.	9 380-2232	
Blake, D. blake@firefly.geology.uiuc.edu	217 333-3540	
Bosch, I.	407 465-2400	407 465-2446
Breton, G.	33 3541 3728	
Briscoe, S.	609 896-5000	
Burke, R.	604 721-710	604 721-8653
Bustos, E.		56 65-259 995
Byrne, M.	61-2-6922497	61-2-6922813
Cabot, E. ecec@miday.uchicago.edu		
Campbell, D.	609-895-5418	
Candia Carnevali, M.	2-236.63.30	

Carey, A. 503-754-3504  
 Castilla, J.C 56-2-222-4561 56-2-222-5515  
 Chen, C.-P. 886-785-8059  
 ZOCP@TWNAS886.bitnet  
 Chia, F.-S. 403 432-3499  
 Clark, H. 64 4 859 609 64 4 857 157  
 read-g@kosmos.wcc.govt.nz  
 Clarke, A. 223 61188  
 Clements, L. 904-744-3950  
 Cobb, J. 334 76161  
 Conand, C. 19 262 29 45 45 19 262 29 00 90  
 David, B. 80-39-63-71 80-39-50-66  
 bdavid@satie-u-bourgogne.fr  
 Davis, K. karendbb@mariposa.stanford.edu  
 Dearborn, J. 207 581-2552  
 Den Besten, P. 30-533084  
 De Ridder, C. 32-2-642 24 12  
 Dobson, W.E. 1 704 262-2668  
 dobsonwe@conrad.appstate.edu  
 Donovan, S. 809 927-2728 9 927-1640  
 Drozdov, A. kva@piboc.marine.su  
 Dubois, P. 2-642.25.17  
 phdubois@ulb.ac.be  
 Ebert, T. 419 594-6767  
 Eckelbarger, K. 207 563-3146 207 563-3119  
 Elsberry, W. elsberry@beta.tricity.wsu.edu  
 Ellers, O. 916 752-1449  
 Emlet, R. 503-888-2581 503-888-3250  
 remlet@oimb.uoregon.edu  
 Emson, R. 71 836 5454 71 937 5396  
 Epel, D. 408 373-0464  
 Ettensohn, F.R. 606 257-1401 606 258-1938  
 geofre@ukcc.uky.edu  
 Eylers, J. 919 220-5429 919 220-5575  
 Fankboner, P. 604 291-4475  
 Farmanfarmaian, A. 908 932-2829 908 932-5870  
 Fenaux, L. 93 76 66 13 93 76 38 34  
 Feral, J.-P. 33 6888 7318 33 6888 1699  
 Ferguson, J. 813 867-1166  
 Fischelson, L. 3 6409812 972 3 6409403  
 zool@taunos  
 Fontaine, A. 604 721-7131 604 721-7120  
 Gage, J. 631 62244 631 65518  
 Gagnon, J.-M. jm\_gagnon@iml1.iml.dfo.ca  
 Garret, R. rhg@uva.pcmail.virginia  
 Gentile, F. 98 69.72.30  
 Gilmour, T. 306 966-4404 306 966-4461  
 Glynn, P. 305 361-4151  
 Giudice, G. 616.26.322 91-616.56.65

Greenstein, B. 413 585-3804  
 BGREENSTEIN@SMITH  
 Guille, A. 68 88 00 40 68 88 16 99  
 Guillou, M. 98.31.62.73 98.31.63.11  
 Guisado A., C.B. 51 311287  
 Haedrich, R. 709 737-8833  
 Hammond, L. 651 1998  
 Harrold, C. 408 649-6466  
 Hart, M. 206 378-2165  
 Hayes, J.  
  
 josh@mowgli.cqs.washington.edu  
 Heck, K. 205 861-2141  
 Hendlar, G. 213 744-6391 213 746-2999.  
 Herrera, J.  
 herrera@zoo.ufl.edu  
 Hide, W.  
 whide@matrix.bchs.uh.edu  
 Hill, R. 401 792-2372  
 Himmelman, J. 418 656-5230 418-656-2339  
  
 Hodgson, A.N. 461 22023 461 24377  
 Holland, N. 619-259-0771  
 Horowitz, A.  
 LANE@UCS.INDIANA.EDU  
 Hopkins, T. 205-861-2141 205-861-4646.  
 Hoshi, M. 45-923-0368 45-923-0368  
 Haeckle, W. 407 465-6630 407 461-8154  
 Jacob, W.  
 rsiu42g@saupm00.bitnet  
 Janies, D.  
 daj@zoo.ufl.edu.  
 Jangoux, M. 2-642 24 12 2- 650 27 96  
 Jensen, M. 31 35 41 11 31 39 81 55  
 Johnsen, S. 919-962-5017 919-962-1625  
 sonkejo@uncmvs.oit.unc.edu  
 Johnson, C. 7-365-2482 7-365-1655  
 zljohanson@uqvax.cc.uq.edu.au  
 Johnson, D.  
 d\_johnson@aims.gov.au  
 Johnson, L.  
 johnson@inst.augie.edu  
 Kazyanov, V. 423-2 296-205 423-2 252-589  
 Keegan, B. 91 24411 91 25700  
 Kemp, P.  
 p.kemp@u.washington.edu  
 Klinger, T.S.  
 klin@bf486.bloomu.edu  
 Knott, K.E. 205 348-7709  
 Kocher, T.  
 t\_kocher@unhh.unh.edu  
 Korrube, J.  
 jkorrube@zoo.uct.ac.za  
 Lane, G.

lane@ucs.indiana.edu  
 Larrain, A. 56-234985 56-240280.  
 Lawrence, J. 813 974-3250 813 974-5273  
 lawr@chuma.cas.usf.edu  
 LeClair, E.  
 lecl@midway.uchicago.edu.  
 Lessios, H. 507 28-4304 407 28-0516  
 Lettiere, D.  
 lettieri\_daj@ccsua.ctstateu.edu  
 Levin, V.  
 service@post.kamchatka.su  
 Levitan, D. 916 752-1272 916 752-1449  
 Littlewood, T. 71-938-9228 71-938-9277  
 MBDTL@DLVH.DARESBURY.AC.UK  
 Lubchenco, J  
 lubchenj@bcc.orst.edu  
 Lucas, J. 77 814111 77 796371  
 Manahan, D. 213 743-7493 213 740-8123  
 Macurda, D. 713 781-6881 713 781-2998  
 Maples, C.  
 maples@ukanvax.bitnet  
 Martin, R. 61 2 325-222 61 2 325-000  
 martin@ml.csiro.au  
 Mariueux, J.  
 jean.mariaux@zool.unine.ch  
 Marlab, G.  
 s-jm@va.nsm.ac.uk  
 Marsh, L. 9 328 4411 9 328 8686  
 Martinez, de Saaverdra y Alvarez.  
 22 25 34 48  
 McClintock, J. 205 934-8308  
 BIOF004@UABDPO.DPO.UAB.EDU  
 McClarey, D.  
 ZOOLOGY@OTAGO.AC.NZ  
 McEdward, L. 904 9328738 904 392-3704  
 lmced@zoo.ufl.edu.  
 McKenzie, J. 631 62244 631 65518  
 S\_JM@UK.AC.NSM.VA  
 McKnight, D.G. 4 386-1189 4 386-2153  
 Meijer, L. 33 98 292339 33 98 292342  
 Menge, B. 503 737-0501  
 mengeb@bcc.orst.edu  
 Meyer, D. 512-556-6931  
 MEYER@UCBEH.SAN.UC.EDU  
 Meyers, G.  
 glm@pol.lanl.gov  
 Michel, E.  
 emichel@ccit.arizona.edu  
 Miller, R. 215 787-6646  
 V5222E@TEMPLEVM  
 Mladenov, P. 64 3 479 8304 64 3 479 8336  
 Mladenov@Otago.ac.nz.  
 Mooi, R. 415 750-7080 415 750-7090  
 Moran, P. 77 78 9211

P Moran@AIMS.GOV.AU  
 Nagle, D. maja@mendel.psycha.upenn.edu  
 Neely, M. neely\_mp@darwin.ntu.edu.au  
 Nebelsick, J. 7071 29 4039 7071 29 6990  
 Nichols, D. 392-263263 392-263700  
 biology@exeter.ac.uk  
 Nojima, S. 969-35-000 969-35-2413  
 Norris, D. 671 472-3002 671 472-3002  
 dnorris@uog.pacific.edu.  
 Ojeda, P. 56-2-222-4516 56-2-222-5515  
 O'Loughlin, P. 347 4211  
 Olsen, R. r-olsen@unhh.bitnet  
 Ozgunen, T. tuncay@trcuniv.bitnet  
 Parsley, R. 504 865-5918  
 Pawson, David. 202-786-2127 202-357-3043  
 Payne, R. rpayne@kean.ucs.mun.ca  
 Pearse, J. 408 459-245 408-459-2882  
 Pentcheff, D. dean2@tbone.biol.scarolina.edu  
 Pentreath, V. 61745 5000 61-745 5999  
 Perez-Ruzafa, A. 34-68-833000 34-68-835418  
 Pezzack, D. d\_pezzack@bionet.bio.df0.ca  
 Phillips, A. phillips@uconnvm.uconn.edu  
 Philippe, M. 7-893.22.44  
 Polson, E. 813-974-3250  
 emma@chuma.cas.usf.edu  
 Procop, R. 42 02 269451 42 02 2369489  
 Propp, M.V. 4232 310912 4232 310900  
 faribm@visenet.marine.su  
 Raff, R. Rraff@sunflower.bio.indiana.edu.  
 Read, G. Read\_g@kosmos.wcc.govt.nz  
 Regis, M. 91.28.84.41 91.28.80.30  
 Regnell, G. 46-121 475 46-121 477  
 Remsen, D. dremsen@hoh.mbl.edu  
 Roller, R. raroller@lub001.lamar.edu  
 Rose, E. 784 434455 784 71780  
 Roux, M. 26 5 33 96  
 Rowley, R. 643-479-8306 643-479-8306  
 RJROWLEY@OTAGO.AC.NZ.  
 Ryabushko, V. 59-19-60 690592813  
 Scheibling, R. 902-424-2211  
 Sewell, T. sewell@uctvax.uct.ac.za

Shick, M. 207-581 0146  
 Shick@Maine.bitnet  
 Shirley, T. 907 789-2101  
 JFTCS@ALASKA.  
 Sibuet, M. 9822 45 47 98 22 43 03  
 msibuet@ifremer.fr  
 Smiley, S. 907-474-7655 907-474-6967.  
 Smirnov, A. 812 218-00-11  
 zisp.pm@pcsti.spb.su  
 Smirnov, I. 812 218-13-11 812 218-29-41  
 zisp.pm@pcsti.spb.su  
 Smith, A. 71 938 8925 071 938 9277  
 MBDTL@DLVH.DARESBURY.AC.UK.  
 Smith, M.  
 msmith@darwin.sfu.ca  
 Spencer, L.  
 lts@oz.plymouth.edu  
 Spina, M.A.  
 mas8f@elvis.med.virginia.edu  
 Sprinkle, J. 512 471-4264 512 471-9425  
 Stancyk, S.  
 stancyk@univscvm.csd.scarlina.edu  
 Stickle, W. 504 388-1739 504 388-1763  
 zostic@lsuvm.bitnet  
 Strathmann, R.  
 fh1@u.washington.edu.  
 Suzuki, N. 768 74-1151 768 74-1644  
 Telford, M. 416 978-4843  
 gmt@zoo.toronto.edu  
 Thandar, A. 820-9111 31 820-2383  
 Thierry, J. 80 39 63 62 80 39 63 87  
 Tyler, P. 703 592557 703 593939  
 Vadas, R. 207-581-2970 207-581-2969  
 vadas@maine.edu  
 Vail, L. 70 60 3976 70 60 3977  
 Valentine, J. 205-861-2141 205-861-4646  
 Vasquez, J.A. 51-311 287  
 Velarde, R.G. 1 619 692-4903 1 619 692 4902  
 Ventura, R. 55021-2807993  
 ventura@ufrj.bitnet  
 Yakolev, Y. 423.2.252.589  
 Young, C. 407 465-6630 407 461-8154  
 Watts, S. 205-934-1045  
 Waters, J.  
 jwaters@uga.bitnet  
 Wilcox, M.  
 s\_mw\_t@vaxa.nerc-oban.ac.uk  
 Wilkie, I. 41-331-3000 41-331-3005  
 Williams, C. 2-20-2630 2-20-2745  
 christopher.williams@zoo.utas.edu.au  
 Zaghib-Turki, D. 1.512.600 216.1.500.666

### REQUESTS AND INFORMATION

Contents of GBRMPA. Crown-of-thorns starfish research update 1991/92. (U. Engelhardt, B. Lassig, eds.). Published by the Great Barrier Reef Marine Park Authority, PO Box 1379, Townsville, Q 4810, Australia.

- Lassig, B. The research continues.  
 Moran, P. Current status of outbreaks.  
 Babcock, R. Spawning.  
 Black, K. Computer models show us where starfish larvae disperse.  
 Keesing, J., Recruitment.  
 Johnson, C. Deep water recruitment hypothesis and the riddle of juveniles.  
 Stump, R., J. Lucas. Ageing of crown-of-thorns starfish.  
 Keesing, J. Feeding.  
 Done, T. How long will it take for the Great Barrier Reef to recover?  
 DeVantier, L. Impact of the crown-of-thorns starfish on massive coral reefs.  
 Lassig, B. So, what causes outbreaks?

### Port Erin Marine Laboratory - Centenary 1992

Symposium, 16-18 September 1992. The marine resources of the Irish Sea; experimental ecology; ecophysiology and behaviour; evolution and genetics.

Museum exhibit, 20 July - 6 January 1993. The surrounded Sea - 100 years of Manx Marine Biology. Liverpool City Museum.

Library Database. The Library, Marine Biological Association of the United Kingdom, Plymouth. The Library has catalogued papers of interest to the MBA and PML scientists as well as all papers on British waters, and all papers on marine pollution world-wide. References back to 1985 are being added to the Library Database, and this can be searched for all aspects of research on British waters and marine pollution.

Ophiuroid taxonomy. Seiichi Irimura is seeking students to assist in work on the taxonomy of Japanese ophiuroids.

Jack Cohen, B.D. Massey (Univ. Birmingham), A. Frettsome (Plymouth Marine Laboratory) are studying the effectiveness of sperm fertilization of Psammechinus miliaris. About 1/1000 sperm fertilize under experimental conditions regardless of the number of eggs.

V.G. Klikushin (St. Petersburg Paleontological Laboratory) is studying pentacrinids and has recently published a monograph on them.

Chantal Conand is now at: Laboratoire de Biologie Marine, Universite de La Reunion, 15 Avenue Rene Cassin, 97489 Saint-Denis cedex, France. La Reunion is a volcanic island in the Indian Ocean.

S. Johnsen (Univ. of North Carolina, Chapel Hill) is studying the responses of echinoderms to polarized light.

Botsford, L.W., J.F. Quinn (Bodega Marine Laboratory, University of California at Davis) at studying the spatial management of the northern California red sea urchin fishery. The population of *S. franciscanus* in northern California has declined nearly one-third. This study is designed to describe the variability in population dynamics over space and time to assist agencies regulating the fishery.

D.J. McKenzie and M.S. Kelly (Scottish Marine Biological Association, Dunstaffnage Marine Laboratory) have been studying the sub-cuticular bacteria of echinoderms from the British Isles. Twenty-five of fifty-one species examined have SCB. The distribution of SCB among species and taxa is very curious.

Maeve Kelly was awarded the Bank of Scotland Waitangi Fellowship to do research at the Portobello Marine Laboratory, University of Otago, Dunedin, New Zealand in the fall of 1992. She collaborated with Michael Barker in studies on the potential vertical transmission of sub-cuticular bacteria in ophiuroids.

John Himmelman (Universite Laval) is spending a sabbatical year (1992-1993) at the Instituto Oceanografico de Venezuela, Universidad del Oriente and the Gatty Marine Laboratory, University of St. Andrews.

Disease in *Strongylocentrotus purpuratus* and *Strongylocentrotus franciscanus*: Mortality from disease for these species was reported by Dayton et al in 1992 (P.K. Dayton, M.J. Tegner, P.B. Edwards. 1992. Temporal and spatial patterns of disturbance and recovery in a kelp forest community. *Ecological Monographs*. 62, 421-445). Dayton reports he has noted the disease since he started studying in the southern California area in 1970, and that Wheeler North and John Pearse saw it before that. He has observed it to cover areas ranging from hectares to ca. 1 square kilometer. He is willing to collect specimens for anyone interested in identifying the pathogen.

J.D. Gage (Scottish Marine Biological Association, Dunstaffnage Marine Laboratory) is comparing growth rates of shallow-water and deep-water echinoids. He and Paul Tyler (Oceanography, Southampton) are collaborating in a dietary study of starfish species from the Rockall Trough. Both of them and Craig Young



(Harbor Branch Institution) have been studying the reproductive biology of deep-sea echinoids.

Mark Wilcox (Dunstaffnage Marine Laboratory) is studying the genetic basis of salinity tolerance in *Asterias rubens*.

Aykie Agatsuma (Hokkaido Central Fisheries) is interested in the population dynamics and reproductive biology of sea urchins. He is studying *Strongylocentrotus nudus*, *S. intermedius*, and *Hemicentrotus pulcherrimus*.

K. Emily Knott (University of Alabama) is interested in the use of molecular techniques in studying the phylogenetic systematics of echinoderms.

Nina L. Leibson (Institute of Marine Biology, Vladivostok) is studying the regeneration of holothuroid tissue, emphasizing the digestive tract.

#### Echinoderms at the Australian Museum (Sydney)

(communicated by Dr. Penny Barents, Collection Manager)  
 Many of you are no doubt aware that Dr. Frank Rowe has left research and the Australian Museum, Sydney, to return to less sunny climes. Our large, well maintained collection bears testament to Frank's diligent labours. Comprehensive in the eastern Australian fauna, the collection also represents shallow and deep-water species of the wider Indo-Pacific, Antarctic regions and beyond. And of course, the Australian Museum is host *Xyloplax* spp. (Concentricycloidea) - fondly known as "Daisy". The collection continues to grow with the collection manager and four technical officers. The dry collections are in controlled environment storage and the records are being put in computers. The Lizard Island Research Station, far North Queensland, is a facility of the Australian Museum. The directors there, Dr. Anne Hoggett and Dr. Lyle Vail, are both echinoderm researchers. So although Frank is no longer with us, our collections and interest in echinoderms continues. We invite any workers to make use of our collections and facilities. Enquiries may be directed to the Collection Manager, Marine Invertebrates.

#### **SCHEDULED MEETINGS**

##### LARVAL ECOLOGY MEETINGS

Marine Sciences Research Center  
 State University of New York  
 Stony Brook, New York, 11764-5000  
 20-22 August 1993 (tentative)  
 S.G. Morgan (516-632-8668), R.K. Cowen (516-632-8711), W. Wise (516-632-8656)  
 FAX (516-632-8820)

#### **PUBLICATIONS OF THE PALEONTOLOGICAL RESEARCH INSTITUTE**

1259 Trumansburg Road

Ithaca, NY 14850-1398 USA

Weisbord, N.E. 1969. Some Late Cenozoic Echinoidea from Cabo Blanco, Venezuela.

Pabian, R.K., H.L. Strimple. 1974. Crinoid studies. part I. Some Pennsylvanian crinoids from Nebraska. part II. Some Permian crinoids from Nebraska, Kansas, and Oklahoma.

Brower, J.C., J. Veinus. Middle Ordovician crinoids from southwestern Virginia and eastern Tennessee.

Parsley, R.L., L.W. Mintz. 1975. North American Paracrinoidea (Ordovician: Paracrinozoa, New, Echinodermata).

Cutress, B.M. 1980. Cretaceous and Tertiary Cidaroida (Echinodermata: Echinoidea) of the Caribbean area.

Guensburg, T.E. 1984. Echinodermata of the Middle Ordovician Lebanon Limestone, central Tennessee.

Chesnut, D.R., F.R. Etnensohn. 1988. Hombergian (Chesterian) echinoderm paleontology and paleoecology, south central Kentucky.

Parsley, R.L. 1991. Review of selected North American mitrate stylophorans (Homalozoa; Echinodermata).

Strimple, H.L. 1949. Studies of Carboniferous crinoids.

Brower, J.C. Crinoids from the Girardeau limestone.

Tuoney, M., F.S. Holmes. Pleiocene fossils of south Carolina, containing descriptions and figures of the Polyparia, Echinodermata and Mollusca.

#### BOOKS

Arakawa, K.Y. 1990. A handbook on the Japanese sea cucumber - its biology, propagation and utilization. (translated by M. Izumi). Midori-Shobo Publishers, 2-14-4 Ikebukuro, Toshima-ku, Tokyo 171).

Dettlaff, T.A., S.G. Vassetzky. (eds.) 1990. Animal species for developmental studies. I. Invertebrates. Plenum. Describes methods of laboratory maintenance, obtaining gametes, artificial fertilization, rearing embryos and larvae. Includes:

Buznikov, G.A., V.I. Podmarev. The sea urchins Strongylocentrotus drobachiensis, S. nudus, and S. intermedius.

Podmarev, V.I. The starfish Asterina pectinifera (Muller et Trochel, 1842).

Johnson, C. (ed.). 1992. Crown-of-thorns starfish on the Great Barrier Reef: reproduction, recruitment and hydrodynamics. CSIRO. Australia. see also: Australian Journal of Marine and Freshwater Research. 43 (1992).

Klikushin, V.G. 1992. Fossil pentacrinid crinoids and their occurrence in the USSR. St. Petersburg. This monograph summarizes the history of the study of pentacrinids, their skeletal morphology, and nomenclatural problems. The taxa are diagnosed, and their geographical and stratigraphical limits defined. The author's concepts of taxonomy and phylogeny are given. The occurrence of pentacrinids in the USSR is discussed in detail. Most known species are illustrated. The life-style of fossil and recent forms is described. Williams, D.I. 1992. Larvae and evolution. Toward a new zoology. Chapman and Hall. Williams suggests the larvae of marine invertebrates may result from captured genes of

non-related organisms. Much of his interpretation is based on the larvae of echinoderms, particularly ophiuroids and echinoids.

Williamson, D.I. 1992. Larvae and evolution. Chapman & Hall. Williamson suggests the anomalies between adults and larvae of many groups, including echinoderms, resulted from the capture by organisms of genes from other organisms. These genes were incorporated into the genome of the host organisms and expressed in larval or juvenile stages. The book considers echinoderms and their larvae, the affinities of echinoderms, the metamorphosis of echinoderms, the relationship between sea-urchins and brittle-stars.

#### **NEWSLETTERS**

Beche-de-mer Information Bulletin. No. 4. July 1992.  
Fisheries Information Project  
PO Box D5, New Caledonia

Group coordinator: Chantal Conand, Laboratoire Oceanographie Biologique, Universite de Bretagne Occidentale, 29287 Brest, France.

#### **Contents:**

Tuwo, A., C. Conand. Developments in beche-de-mer production in Indonesia during the last decade.

Byrne, M., C. Conand. Request for information on spawning behaviour of tropical holothurians.

Arakawa, K.Y.A handbook on the Japanese sea cucumber--its biology, propagation, and utilization.

Lokani, P. First results of an internal tag retention experiment on sea cucumber.

Beumer, J. Queensland's beche-de-mer fishery.

Adames, T. Resource aspects of the Fiji beche-de-mer industry.  
Beche-de-mer correspondence.

Sea Urchin, Kelp, Abalone News. No. 1. March 1993.

Available from the editor:

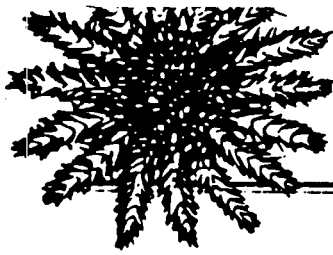
Dr. Leon T. Davies

Sea Grant Extension Program

University of California

Department of Wildlife and Fisheries Biology

Davis, CA 95616-8751



# COTS COMMS

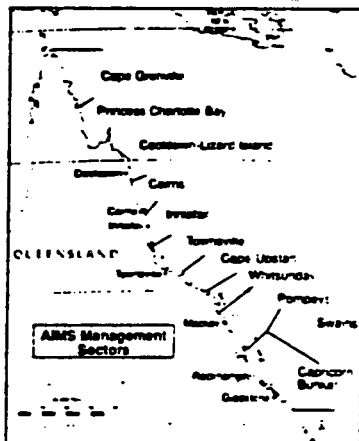
*Dr Brian Lassig and Udo Engelhardt*

## Current COTS

Since the last issue of COTS COMMS the AIMS survey team has been to the two extremes of the Marine Park - the Swain and Capricorn/Bunker Sectors in the south and the Far Northern Section (I'll resist the tautology!). Active crown-of-thorns starfish (COTS) outbreaks were observed on two reefs in the Swains (Gannet Cay and Snake Reef) and, with coral cover on these reefs exceeding 50% away from the localised aggregations, the outbreaks seem likely to be around for some time to come.

The Capricorn/Bunker reefs maintained their consistency of having no COTS visible to manta towers, but they too have problems. The cover of hard corals was less than 30% on all of the four reefs in this sector surveyed. The coral cover at One Tree Island and Lady Musgrave Island had decreased significantly since last year's survey, probably as a result of cyclone Fran which visited the area in March last year. Lady Musgrave in particular seems to be suffering, going from a hard coral cover of >75% in 1986/87 to a level of <10% this year. A bid by the Authority for additional funds from the Federal Government to more closely investigate this dramatic decline was unsuccessful.

Sixteen reefs were surveyed in the Far Northern Section which includes three of AIMS' Sectors - Cape Grenville, Princess Charlotte Bay and Cooktown/Lizard Island. No outbreaks were observed although several of the surveyed reefs are recovering from previous outbreaks.



## Swan Song Poem

As the last issue of COTS COMMS announced, Professor John Swan has retired from the chair of the Crown-of-thorns Starfish Research Committee (COTSREC) and been succeeded by Professor Graham Mitchell. John commemorated the occasion of his retirement by bequeathing the following literary legacy:

### *The Age of Reason and Acanthaster planci*

Under the December moon, more often under sun,  
Chance encounters during spawning, there are  
almost none.

The female egg, as ever was the case,  
Attracts the male sperm to her embrace.  
And chemotaxis, as the name implies,  
Makes no distinction 'twixt the foolish and the wise.  
And what is more, the oft-observed communal  
aggregation,  
Ensures for all a most successful starfish  
propagation.

Gastrula! Bipinnaria! Brachiolaria!  
Fourteen days of buoyant freedom,  
sometimes twenty-eight,  
A pea-soup diet, *Dunaliella*, presently their  
fate.

Then down they go! To bed with crustose algae and  
*Lithothamnium pseudosorum*.

Now watch each starfish grow from its  
remarkable primordium.

Then come the fish, the crabs, the worms  
Their brothers, sisters, cousins,  
That finally reduce our *Acanthaster* population  
From the millions to the dozens.

A few more months and then, the promised land!  
Where milk and honey flow from coral strand.  
White scars appear on branching staghorn corals,  
And far away - the muted roar of academic  
quarrels.

"It's predator removal"; "Heavy rainfall  
after drought";

"It's the farmers"; "It's El Nino"; "Larval  
transport without doubt".

It seems the reef is "under threat",

And who to blame? - the Managers, I'll bet.

But Johnson points to H. McCallum's optimistic simulations:

"The patterns of the starfish plagues in reef sub-populations,  
Are cyclical, or quite chaotic, numbers high or low,  
Depending on the larval pulses, water movement,  
ebb and flow."

The oceans are resilient, their systems are robust,  
Life forms adapt, evolve and flourish; corals will adjust.

So shoot the foxes, feral cats, the pigs, the goats and rabbits,

But learn to love the crown-of-thorns;  
accept its natural habits.

### Causes and Consequences

The proceedings of a COTS workshop ('The Possible Causes and Consequences of Outbreaks of the Crown-of-Thorns Starfish') held in Townsville in June last year has finally been printed. The publication contains 17 contributions covering a variety of research and management aspects of the Authority's COTS program. Abstracts of the papers were included in Issue #10 of COTS COMMS. The real thing (168 pages) can be obtained by contacting either Brian Lassig or Udo Engelhardt of the Authority.

The Possible Causes and  
Consequences of Outbreaks  
of the  
Crown-of-Thorns Starfish



### COTSREC Confabulates

COTSREC met in Townsville on 19 January 1993 for its first meeting with Professor Graham Mitchell as Chair. The meeting was primarily concerned with the research program's progress with some discussion of directions in the 1993/94 financial year.

The Committee made a number of recommendations that will be put to the Marine Park Authority for approval. These included support for a project by Dr Kerry Black of the Victorian Institute of Marine Sciences to investigate field testing of hydrodynamic

models (contingent upon substantial Australian Research Council funding); maintenance of the Australian Institute of Marine Science crown-of-thorns starfish rearing program for a further 3 years (subject to the availability of funds and specimens), and collaboration with the Cooperative Research Centre (a consortium of agencies comprised of AIMS, GBRMPA, James Cook University, Queensland Department of Primary Industries and the Association of Marine Park Tourist Operators) in hydrodynamic and water quality programs to achieve mutual goals.

### Local Controls

Can and should outbreaking populations of COTS be controlled? This rather emotive question has split public and scientific opinion for many years. The long standing policy of the Great Barrier Reef Marine Park Authority (GBRMPA) on this issue is not to interfere on a large scale unless it can be shown that outbreaks are initially caused or exacerbated by human activity (Kelleher 1993). However, local control efforts may be initiated in an attempt to protect sites of particular value to tourism or science. Scientific reviews of this matter have supported the Authority's policy.

Historically (ie. in the past 10-15 years), local-scale attempts to control starfish populations have relied largely on the use of a toxic agent such as copper sulphate (CuSO<sub>4</sub>). Preliminary field experiments into methods suitable for controlling the starfish were conducted at Green Island in the early 1980s. The researchers concluded that of the variety of chemical agents tested, CuSO<sub>4</sub> had some outstanding properties, making it the most effective and efficient agent available (Hicks and Blackford 1981). Their work suggested that approximately 7 - 9.5 ml of CuSO<sub>4</sub> may be sufficient to kill individual starfish. However, their report did not provide any information on possible seasonal or size dependent variability in the actual amount of toxic compound required to kill starfish. Copper sulphate injections have since been the preferred means for controlling local scale populations of starfish.

Although the latest wave of COTS outbreaks appears to be coming to an end, some relatively large populations remain on reefs in the Whitsunday Region and further south in the Swains complex. A relatively large, outbreaking (?) population of COTS can still be found at Bait Reef off the Whitsunday Islands. This typical mid-shelf reef is located some 35 nautical miles off the coast in the Central Section of the Great Barrier Reef Marine Park.

Bait Reef is the main focus for the local diving

FOSSIL ECHINODERM COLLECTIONS -- MUSEUM FÜR NATURKINDE ZU BERLIN

During April 1992, I had the opportunity to examine the fossil echinoderm collections at the Museum für Naturkunde, der Humboldt-Universität zu Berlin (the Natural History Museum in the former East Berlin) during the Berlin Week, sponsored by the German Fulbright Commission. With this brief report, I wish to inform echinoderm paleontologists of the availability, magnitude, general content, and crinoid holdings of this important collection.

The Museum für Naturkunde is best known for its specimens of Archaeopteryx (the oldest bird; holdings include the best single individual, arguably the most significant fossil specimen, and an isolated feather) and for the largest mounted dinosaur, a splendid specimen of Brachiosaurus. In addition the Museum für Naturkunde houses a quite significant fossil echinoderm collection, including many type specimens and other referred collections. The Echinoderm collection fills approximately 35 ranges (13 drawers per range). Material is arranged systematically and is divided among Paleozoic crinoids (seven ranges), Mesozoic and Cenozoic crinoids (five ranges), blastozoans and homalozoans (two and one-half ranges together), Paleozoic echinoids (less than one range), Mesozoic and Cenozoic echinoids (17 ranges), asteroids (two ranges), and ophiuroids (one and one-half ranges).

Time permitted only a careful examination of the crinoids. This material is curated very well and includes more than 70 primary types, principally from localities in Germany. Primary types are from the work of H.E. von Beyrich, O. Jaekel, C.F. Roemer, and W.E. Schmidt. The collection was apparently amassed largely during the beginning of this century under the curatorships of Beyrich and Jaekel. Strengths of the crinoid collection are materials from Germany. Excellent collections include the pyritized crinoids from the Lower Devonian Hunsrückschiefer, Budenbach and vicinity, Germany; the Lower Devonian Coblenzian, Germany; the Middle Devonian Eifelian fauna; the fauna from the Lower Triassic Muschelkalk; and crinoids from the Jurassic Solenhofen Limestone.

Collections from outside Germany were presumably part of trades or purchases, perhaps from Frank Springer, U.S. National Museum of Natural History, and F.A. Bather, British Museum (Natural History). These presumed acquisitions include representative collections from the Lower Mississippian of Crawfordsville, Indiana, USA; the Silurian of Gotland, Sweden; and the Lower Carboniferous (Tournaisian) of Yorkshire, England. Smaller collections from outside Germany include Late Ordovician of Cincinnati, Ohio, USA; Silurian of western Tennessee, USA; southern Indiana, USA; northeastern Illinois, USA; and Dudley, England; Lower Mississippian of central Tennessee, USA and eastern Iowa, USA; Jurassic of England; and the Cretaceous of England. The only type and figured specimens from the United States are specimens from Roemer (1960), from the Late Silurian of Decatur County, Tennessee. A few other non-German type specimens include material from Bohemia, Columbia, and Russia.

During this visit I also learned of the existence of the

Roemer-Pelizaeus-Museum in Hildesheim, Germany<sup>1</sup>. Work at this museum includes a determination of the deposition of fossil collections which formed the basis of C.F. Roemer's scientific papers. Roemer was one of the earlier European scientists to make fossil collections in the New World and to name new species. Fossil echinoderms were named in at least (9) publications from 1844 to 1881, and many type and figured specimens were deposited in European museums.

Dr. Hermann Jaeger, Director of the Paleontological Institute, and Dr. Erika Pietrzeniuk were gracious hosts during my visit to the Museum für Naturkinde. Please feel free to contact me if you would like any additional information, including further data on Paleozoic crinoid primary types.

Roemer, C.F. 1860. Die silurische Fauna des westlichen Tennessee. E. Trewendt; Breslau, Poland; 100 p.

<sup>1</sup>Dr. Helga Stein  
Roemer-Pelizaeus-Museum  
Am Steine 1-2  
W-3200 Hildesheim  
GERMANY

William I. Ausich  
Department of Geological Sciences  
155 South Oval Mall  
The Ohio State University  
Columbus, OH 43210 USA

# Crown-of-thorns Starfish on the Great Barrier Reef: Reproduction, Recruitment and Hydrodynamics



Editor: Craig Johnson



# Crown-of-thorns Starfish on the Great Barrier Reef: Reproduction, Recruitment and Hydrodynamics

Edited by Craig Johnson  
*Department of Zoology, University of Queensland*

(Reprinted from *Australian Journal of Marine and Freshwater Research*, Volume 43, Number 3, 1992)

## Contents

- 
- Reproduction, recruitment and hydrodynamics in the crown-of-thorns phenomenon on the Great Barrier Reef: introduction and synthesis. *Craig Johnson.*
- Reproductive biology, spawning and field fertilization rates of *Acanthaster planci*. *R C Babcock and C N Mundy.*
- Observations of crown-of-thorns starfish spawning. *William Gladstone.*
- Enhancement of larval and juvenile survival and recruitment in *Acanthaster planci* from the effects of terrestrial runoff: a review. *Jon E Brodie.*
- Pattern of outbreaks of crown-of-thorns starfish (*Acanthaster planci* L.) along the Great Barrier Reef since 1966. *P J Moran, G De'ath, V J Baker, D K Bass, C A Christie, I R Miller, B A Miller-Smith and A A Thompson.*
- Larval dispersal simulations: correlation with the crown-of-thorns starfish outbreaks database. *M K James and J P Scandol.*
- Hydrodynamics and larval dispersal: a population model of *Acanthaster planci* on the Great Barrier Reef. *J P Scandol and M K James.*
- Review of the genetics, dispersal and recruitment of crown-of-thorns starfish (*Acanthaster planci*). *J A H Benzie.*
- Settlement and recruitment of *Acanthaster planci* on the Great Barrier Reef: questions of process and scale. *Craig Johnson.*
- Recruitment of *Acanthaster planci* over a five-year period at Green Island Reef. *David A Fisk.*
- Importance of postsettlement processes for the population dynamics of *Acanthaster planci* (L.). *Jolin K Keesing and Andrew R Halford.*
- Completing the circle: stock-recruitment relationships and *Acanthaster*. *Hamish McCallum.*

THE UNIVERSITY OF CHICAGO

PHYSICS DEPARTMENT  
530 CHICAGO  
ILLINOIS 60637  
TEL. 733-4331

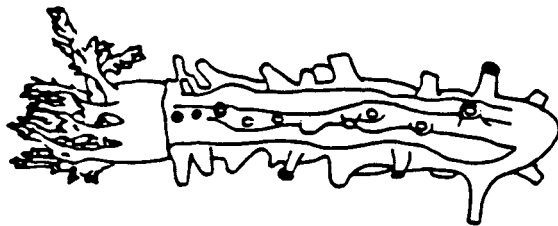


THE FAUNA OF  
THE CLYDE SEA AREA  
ECHINODERMATA

57

by

I.C. WILKIE, B.Sc., Ph.D.



OCCASIONAL PUBLICATION  
No. 6

UNIVERSITY MARINE BIOLOGICAL STATION  
MILLPORT  
ISLE OF CUMBRAE

1989



The Kyles of Butc. from Forbes (1841).

# ECHINODERM STUDIES

*Edited by*  
**MICHEL JANGOUX**  
*Université Libre de Bruxelles, Belgium*

**JOHN M. LAWRENCE**  
*University of South Florida, Tampa, USA*

## VOLUME 4

How to study evolution in echinoderms? <i>Bruno David</i>	1
Comparative physiology of echinoderm muscle <i>Robert B. Hill</i>	81
Pharmacological effects of compounds from echinoderms <i>J.F. Verbist</i>	111
An index of names of recent Asteroidea – Part 2: Valvatida <i>Ailsa M. Clark</i>	187
Note from the Editors	367



A.A. BALKEMA / ROTTERDAM / BROOKFIELD / 1993

## RECENT PUBLICATIONS ON ECHINODERMS

- Agatsuma, Y. 1992. Annual reproductive cycle of the sea urchin, Hemicentrotus pulcherrimus in southern Hokkaido. *Suisanzoshoku*. 40, 475-478.
- Agatsuma, Y., T. Nishikiori. 1991. Gonad development of the northern sea urchin, Strongylocentrotus nudus, that experimentally fed fishes. I. Gonad development. *Sci. Repts. Hokkaido Fish. Exp. Stn.* (37), 59-66.
- Ahearn, C. 1992. Phillipine brittlestars (Echinodermata: Ophiuroidea) described by R. Koehler (1922): a corrected and annotated list of type specimens. *Smithsonian Contrib. Zool.* No. 536.
- Ali, M.S.M. 1992. A new genus and species of regular echinoid from the Late Cretaceous of Gebel El Rowdah, Hatta area, Oman-U.A.E. *J. Paleont.* 66, 693-696.
- Ali, M.S.M. 1992. Additional echinoids from the Late Maestrichtian (Cretaceous) of Gebel El Rowdah, Oman - United Arab Emirates. *N. Jb. Geol. Paläont. Mh.*, H(2), 65-74.
- Ameziane-Cominardi, N., J.P. Bourseau, M. Roux. 1991. Les crinoïdes pedoncules de l'W Pacifique: un modèle zoobathmétrique pour l'analyse des calcaires à Entroques et du tectono-eustatisme au Jurassique. *Doc. et Trav. IGAL*, Paris. no. 15, 182-207.
- Amano, T., Y. Okita, T. Matsui, M. Hoshi. 1992. Pretreatment effects of jelly components on the sperm acrosome reaction and histone degradation in the starfish, Asterina pectinifera. *Biochem. Biophys. Res. Comm.* 187, 268-273.
- Amano, T., Y. Okita, T. Okinaga, T. Matsui, I. Nishiyama, M. Hoshi. 1992. Egg jelly components responsible for histone degradation and acrosome reaction in the starfish, Asterina pectinifera. *Biochem. Biophys. Res. Comm.* 187, 274-278.
- Amano, T., Y. Okita, M. Hoshi. 1992. Treatment of starfish sperm with egg jelly induces the degradation of histones. *Dev. Growth Differ.* 34, 99-106.
- Anstrom, J.A. 1992. Organization of the ciliary basal apparatus in embryonic cells of the sea urchin, Lytechinus pictus. *Cell Tissue Res.* 269, 305-313.
- Anstrom, J.A.. 1992. Microfilaments, cell shape changes, and the formation of primary mesenchyme in sea urchin embryos. *J. Exp. Zool.* 264, 312-322.
- Amemiya, S., R.B. Emlet. 1992. The development and larval form of an echinothuriid echinoid, Asthenosoma ijimai, revisited. *Biol. Bull.* 182, 15-30.
- Amemiya, S., Y. Nakajima. 1992. First electron microscopical study on the sperm morphology of the sea lily (Crinoidea, Echinodermata). *Zool. Sci.* 9, 897-904.
- Antonelli, P.L., R.H. Bradbury, N.D. Kazarinoff, X. Lin, R.E. Reichelt. 1992. Large-scale starfish waves and reefal connectance. *Ecol. Modelling.* 61, 187-194.
- Aronson, R.B. 1992. The effects of geography and hurricane disturbance on a tropical predator-prey interaction. *J. Exp. Mar. Biol. Ecol.* 162, 15-33. (opiuroids)
- Aronson, R.B. 1992. Biology of a scale-independent predator-

prey interaction. *Mar. Ecol. Prog. Ser.* 89, 1-13.

Ausich, W.I., T.W. Kammer. 1992. Dizygocrinus: Mississippian camerate crinoid (Echinodermata) from the midcontinental United States. *Paleontol.* 66, 637-658.

Ayling, A.M., A.L. Ayling. 1991. Crown-of-thorns and coral trout density on three central section reefs - 1983-1989. Great Barrier Reef Authority, Res. Pub. No. 15.

Babcock, R.C., C.N. Munday. 1992. Reproductive biology, spawning and field fertilization rates of Acanthaster planci. *Austr. J. Mar. Freshwater Res.* 43, 525-534.

Balser, E.J., E.D. Ruppert. 1993. Ultrastructure of axial vascular and coelomic organs in comasterid featherstars (Echinodermata: Crinoidea). *Acta Zool.* 74, 87-101.

Baker, V.J., D.K. Bass, C.A. Christie, I.R. Miller, A.A. Thompson. 1992. Broadscale surveys of crown-of-thorns starfish on the Great Barrier Reef 1991 to 1992. Australian Institute of Marine Science, Townsville.

Baldwin, J., A. Patak, K. Mortimer. 1992. Echinoderm (Holothuria atra) lactate dehydrogenase: affinities with the putative ancestral chordate enzyme. *Biochem. Syst. Ecol.* 20, 535-539.

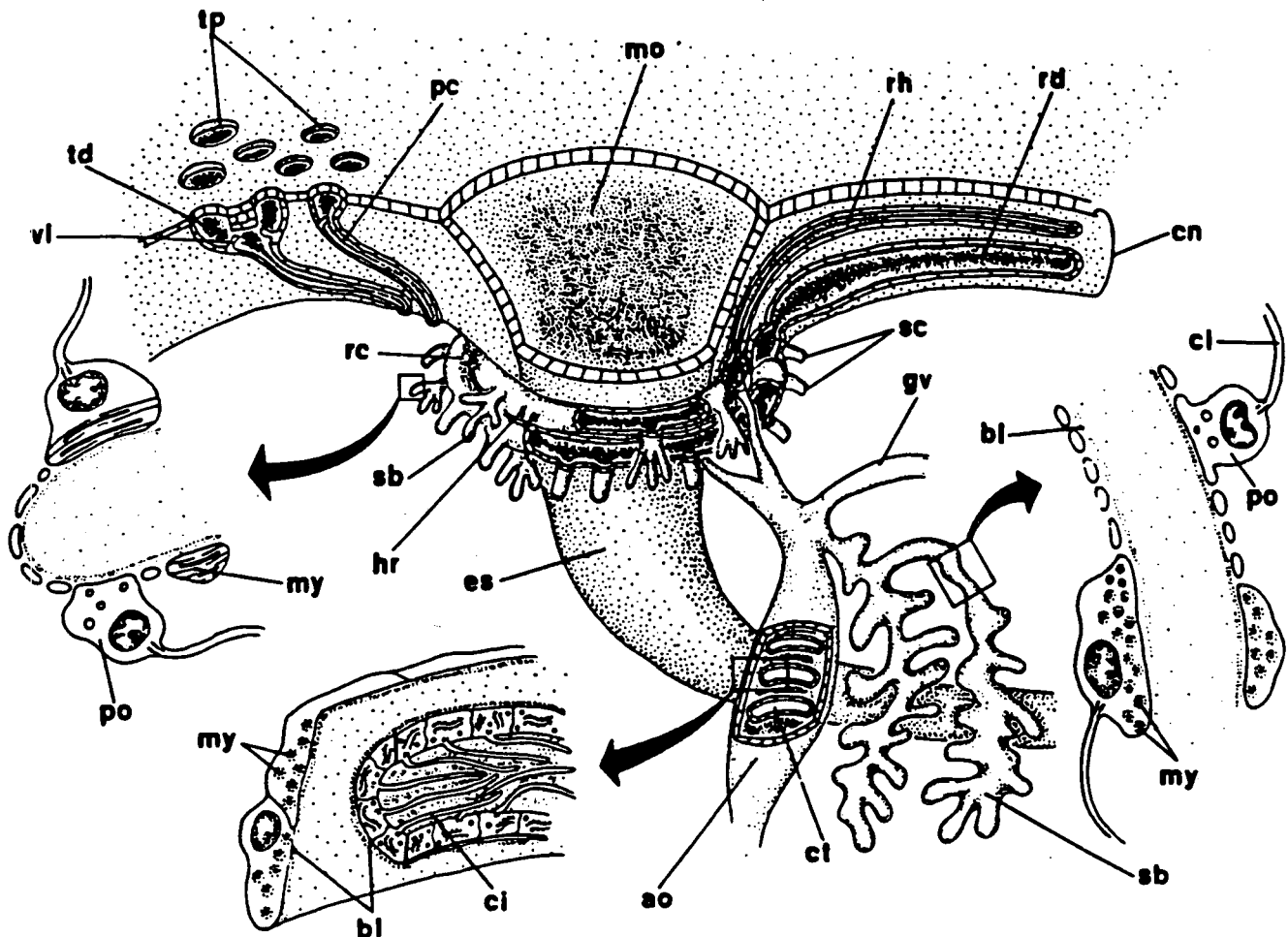


Fig. 1. Reconstruction of the oral axial organs of a comasterid crinoid based on serial thick sections for light microscopy and representative thin sections for electron microscopy. *ao* Axial organ; *bl* basal lamina; *ci* cilium; *cn* connective tissue; *ct* cellular tube; *es* esophagus; *gv* genital blood vessel; *hr* hemal ring; *mo* mouth; *my* myocyte, myofilaments; *pc* parietal canal; *po* podocyte; *rc* ring canal; *rd* radial canal; *rh* radial haemal canal; *sb* oral and axial parts of spongy body; *sc* stone canal; *td* tegmental duct; *tp* tegmental pore; *vl* valve.

BALSER & RUPPERT 1993

Baumiller, T.K. 1992. Importance of hydrodynamic lift to crinoid autecology, or, could crinoids function as kites? *J. Paleontol.* 676, 658-665.

Baumiller, T.K., W.I. Ausich. 1992. The Broken-Stick model as a null hypothesis for crinoid stalk taphonomy and as a guide for the distribution of connective tissue in fossils. *Paleobiology.*

Barthel, D., J. Gutt. 1992. Sponge associations in the eastern Weddell Sea. *Antarctic Sci.* 4, 137-150. (ophiuroids and asteroids)

Barton, N.R., E.M. Bonder, J.D. Fishkind, R.H. Warren, M.M. Pratt. A novel vesicle-associated protein (VAP-1) in sea urchin eggs containing multiple RNA-binding consensus sequences. *J. Cell Sci.* 103, 797-809.

Beckley, L.E., G.M. Branch. 1992. A quantitative scuba-diving survey of the sublittoral macrobenthos at subantarctic Marion Island. *Polar Biol.* 11, 553-563.

Bell, J., B.R. Char, R. Maxson. 1992. An octamer element is required for the expression of the alpha H2B histone gene during the early development of the sea urchin. *Dev. Biol.* 150, 363-371.

Benedetti-Cecchi, L., F. Cinelli. 1992. Effects of canopy cover, herbivores and substratum type on patterns of *Cystoseira* spp. settlement and recruitment in littoral rockpools. *Mar. Ecol. Prog. Ser.* 90, 193-200. (*Paracentrotus*)

Benzie, J.A.H. 1992. Review of the genetics, dispersal and recruitment of crown-of-thorns starfish (*Acanthaster planci*). *Aust. J. Mar. Freshwater Res.* 43, 597-610.

Benzie, J.A.H. 1992. Genetic structure of outbreaking and non-outbreaking crown-of-thorns starfish (*Acanthaster planci*) populations on the Great Barrier Reef. *Mar. Biol.* 112, 119-130.

Benzie, J.A.H. 1992. Genetic structure of crown-of-thorns starfish *Acanthaster planci* in Australia. *Mar. Biol.* 112, 631-

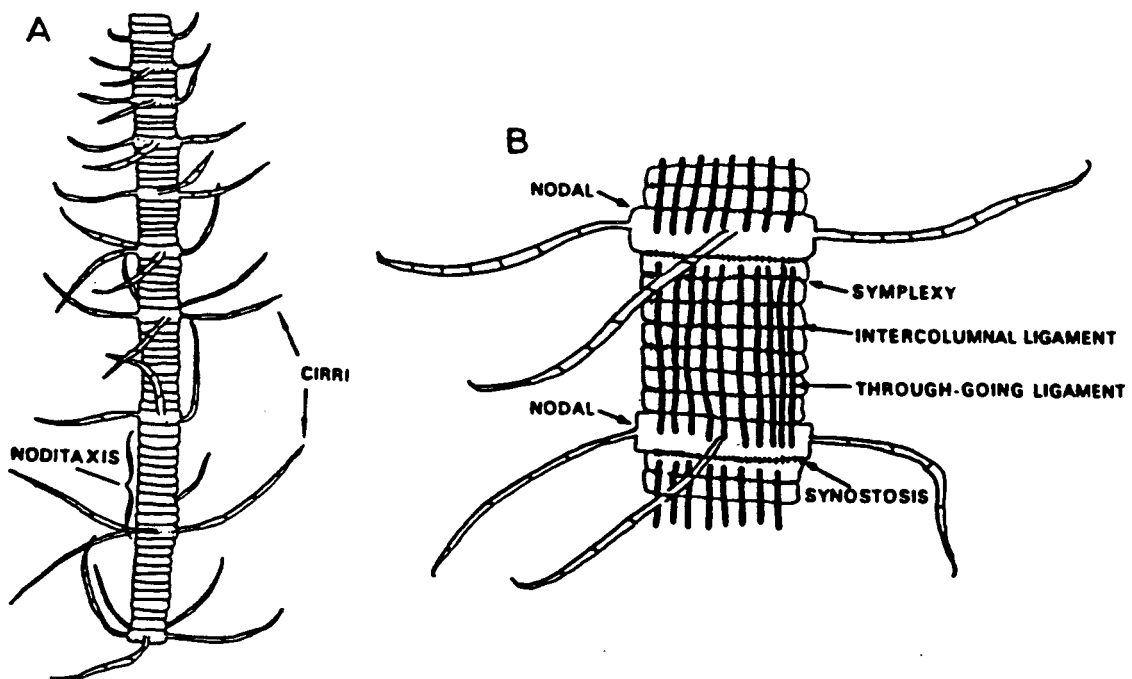


FIGURE 3. A schematic diagram of an isocrinid stalk. A. The proximal (top) and medial parts of the stalk are represented. Nodals are stippled. Note that the number of columnals/noditaxis increases distally in the proximal portion; it reaches a more or less constant value in the medial and distal portions. B. Partial segment of the medial or distal portion of the stalk. The through-going ligament connects a single noditaxis. Note that only intercolumnal ligament is present at synostoses whereas both types of ligament are present at symplectial articulations.

Baumiller + Ausich 1992

639.

- Berger, J., D.H. Lynn. 1992. Hydrogenosome-methanogen assemblages in the echinoid endocommensal plagiopylid ciliate, Lechriopyla mystax Lynch, 1930 and Plagiopyla minuta Powers, 1933. *J. Protozool.* 39, 4-8.
- Bergman, M.J.N., M. Hup. 1992. Direct effects of beam trawling on macrofauna in a sandy sediment in the southern North Sea. *J. Mar. Sci.* 49, 5- 11. (Echinocardium, ophiuroids)
- Bergwerff, A.A., S.H.D. Hulleman, J.P. Kamerling, J.F.G. Vliegthart, L. Shaw, G. Reuter, R.Schauer. 1992. Nature and biosynthesis of sialic acids in the starfish Asterias rubens: Identification of sialo-oligomers and detection of A-adenosyl-L-methionine: N-acylneuraminate 8-O- methyltransferase and CMP-N-acetylneuraminate moxygenase activities. *Biochimi.* 74, 25-37..c
- Bett. B.J., A.L. Rice. 1992. The influence of hexactinellid sponge (Pheronema carpenteri) spicules on the patchy distribution of macrobenthos in the Porcupne Seabight (bathyal NE Atlantic). *Ophelia.* 36, 217-226.
- Bilinkis, A.A., G.G. Sekirina, M.N. Skoblina, S.G. Vasetskii. 1992. Maturation in vitro of oocytes and cell hybrids of the starfish Aphelasterias japonica. *Ontogenez.* 23, 511-517.
- Bing, H.X., R.R. Schmidt. 1992. Glycosyl imidates: 55. Synthesis of the pentassacharide moiety of an asterosaponin. *Liebigs Ann. Chem.* (8), 817-823.
- Bishop, C.D., S.A. Watts. 1992. Biochemical and morphometric study of growth in the stomach and intestine of the echinoid Lytechinus variegatus (Echinodermata). *Mar. Biol.* 114, 459-467.
- Black, K.P., P.J. Moran, L.S. Hammond. 1991. Numerical models show coral reefs can be self-seeding. *Mar. Ecol. Prog. Ser.* 74, 1-11. (Acanthaster)
- Bode, L., I.J. Dight, M.K. James, L.B. Mason, J.P. Scandol. 1992. Modelling approach to hydrodynamics and the large-scale larval dispersal of Acanthaster planci. Report to COTSREC, Great Barrier Reef Marine Park Authority.
- Borri, M., P. Righini, A. Piras. 1991. The echinological fauna of the Upper Tyrrhenian Sea soft bottoms and notes on the relative biocenosis. *Atti Soc. Ital. Sci. Nat. Mus. Civ. Stor. Nat. Milano.* 131, 377-410.
- Bosch, I. 1992. Symbiosis between bacteria and oceanic sea star larvae in the western north Atlantic Ocedan. *Mar. Biol.* 114, 495-402.
- Bourseau, J.P., R. Avocat, N. Ameziane-Cominardi, M. Roux. 1991. Crinoides pedoncules de Nouvelle-Caledonie. *Mem. Mus. natn. Hist. nat. (A), Paris.* 8, 229-333.
- Brey, T., S. Hain. 1992. Growth, reproduction and production of Lissarca notocadensis (Bivalvia: Philobryidae) in the Weddell Sea, Antarctica. *Mar. Ecol. Prog. Ser.* 82, 219-226. (commensal on spatangoids)
- Bressan, M., R. Brunetti, S. Casellato, G.C. Fava, P. Giro, M. Marin, P. Negrisol, L. Tallandini, S. Thomann, L. Tosoni, M. Turchetto. 1989. Effects of linear alkylbenzene (LAS) on benthic organisms. *Tenside Surfactants Detergents.* 26, 148-158. (Paracentrotus larvae)
- Bressan, M., M.G. Mrin, R. Brunetti. 1991. Effects of linear



- alkylbenzene sulfphonate (LAS) on skeletal development of sea urchin embryos (Paracentrotus lividus LMK). *Wat. Res.* 25, 613-616.
- Brodie, J.E. 1992. Enhancement of larval and juvenile survival and recruitment in Acanthaster planci from the effects of terrestrial runoff: a review. *Aust. J. Mar. Freshwater Res.* 43, 539-554.
- Brower, J.C. 1992. Cupulocrinid crinoids from the Middle Ordovician (Galena Group, Dunleith Formation) of northern Iowa and southern Minnesota. *J. Paleontol.* 66, 99-128.
- Brunetti, R., M. Bressan, M.G. Marin, M. Fuolega. 1989. Sodium nitrilotriacetate (NTA) influences the larval development and metamorphosis of marine invertebrates. *Bull. Environ. Contam. Toxicol.* 42, 234-240.
- Brunetti, R., M. Marin, M. Bressan, M. Zordan, A. Soggia. 1991. Effects of the chelating agent nitrolotriacetic acid (NTA) on the toxicity of metals (Cd, Cu, Zn and Pb) in the sea urchin Paracentrotus lividus LMK. *Vie Milieu.* 41, 39-43.
- Bruno, I., M.V. D'Auria, M. Iorizzi, L. Minale, R. Riccio. 1992. Marine eicosanoids: occurrence of 8,11,12-trihydroxylated eicosanoic acids in starfishes. *Experientia.* 48, 114-116.
- Bulteel, P., M. Jangoux, P. Coulon. 1992. Biometry, bathymetric distribution, and reproductive cycle of the holothuroid Holothuria tubulosa (Echinodermata) from Mediterranean seagrass beds. *Mar. Ecol.* 13, 53-62.
- Bustos R., E., C. Godoy A., S. Olave M., Ricardo Troncoso T. 1991. Desarrollo de técnicas de producción de semillas y repoblación de recursos bentónicos. I. Investigaciones en el erizo chileno Loxechinus albus (Molina, 1782). Instituto de Fomento Pesquero, Santiago.
- Bulteel, P., M. Jangoux, P. Coulon. 1992. Biometry, bathymetric distribution, and reproductive cycle of the holothuroid Holothuria tubulosa (Echinodermata) from Mediterranean seagrass beds. *Mar. Ecol. Pbb. Staz. Zool. Nap.* 13, 63-68.
- Byrne, M. 1992. Reproduction of sympatric populations of Patiriella gunnii, P. calcar, and P. exigua in New South Wales, asterinid seastars with direct development. *Mar. Biol.* 114, 297-316.
- Canicatti, C. 1992. The echinoderm lytic system. *Boll. Zool.* 59, 159-166.
- Canicatti, C., P. Pagliara, L. Stabili. 1992. Sea urchin coelomic fluid agglutinin mediates coelomocyte adhesion. *Eur. J. Cell Biol.* 58, 291-295.
- Canicatti, C., F. Rossigno. 1992. Cellular reactions mimicking encapsulation in Holothuria polii (Echinodermata). *Eur. Arch. Biol.* 103, 51-55.
- Cantera K., J.R., R. Neira O. 1987. Primer registro del genero Echineulima Lutzen y Nielsen (Gastropoda: Eulimidae), moluscos parasitos de erizos de mar en la isla de Gorgona (Pacífico Colombiano). *An. Inst. Inv. Mar. Punta de Betin.* 17, 87-93.
- Candia Carnevali, M.D., I.C. Wilkie. 1992. Gli straordinari tessuti connettivi degli echinodermi. *Le Scienze.* (286), 58-70.
- Cantera, J.R., H. von Prahl, R. Neira O. 1987. Moluscos, crustaceos y equinodermos asociados a la gorgonia Lophogorgia

- alba Duchassaing y Michelotti, 1864 en la isla de Gorgona, Colombia. Boletín Ecotropical. (17), 3-23.
- Carginale, V., A. Capasso, L. Madonna, L. Borrelli, E. Parisi. 1992. Adenylate cyclase from sea urchin eggs is positively and negatively regulated by D-1 and D-2 dopamine receptors. Exp. Cell Res. 203, 491-494.
- Carter, B.D., M.L. McKinney. 1992. Eocene echinoids, the Suwannee Strait, and biogeographic taxonomy. Paleobiology. 18, 299-325.
- Cartes, J.E., P. Abello. 1992. Comparative feeding habits of polychelid lobsters in the Western Mediterranean deep-sea communities. Mar. Ecol. Prog. Ser. 84, 139-150. (asteroids, echinoids, ophiuroids as prey)
- Caso, M.E. 1990. Une nuevo asteroideo del Caribe Mexicano Astropecten caribemexicanensis sp. nov. y comparacion con la especie afin Astropecten nitidus Verrill. An. Inst. Cienc. Mar. Limnol. Univ. Nac. Auton. Mex. 17, 107-130.
- Cervello, M., D. Di Ferro, L. D'Amelio, F. Zito, V. Matranga. 1992. Calcium-dependent self-aggregation of toposome, a sea urchin embryo cell adhesion molecule. Biol. Cell. 74, 231-234.
- Chang, S., F.W. Steimle, R.N. Reid, S.A. Fromm, V.S. Zdanowicz, R.A. Pikanowski. 1992. Association of benthic macrofauna with habitat types and quality in the New York Bight. Mar. Ecol. Prog. Ser. 89, 237-251. (Echinarachnius)
- Chao, S.-M., C.-P. Chen, P.S. Alexander. 1993. Reproductive periodicity of a tropical dendrochirote holothurian, Phyrella fragilis (Echinodermata: Holothuroidea), in Taiwan. Bull. Inst. Zool., Academia Sinica. 32, 111-119.
- Chen, B.-Y., C.-P. Chen. 1992. Reproductive cycle, larval development, juvenile growth and population dynamics of Patiriella pseudoexigua (Echinodermata: Asteroidea) in Taiwan. Mar. Biol. 113, 271-280.
- Chen, B.-Y., C.-P. Chen. 1993. Reproduction and development of a miniature sand dollar, Sinaechinocyamus mai (Echinodermata: Echinoidea) in Taiwan. Bull. Inst. Zool., Academia Sinica. 32, 100-110.
- Chen, C.-P., B.-Y. Chen. 1992. Effects of high temperature on larval development and metamorphosis of Arachnoides placenta (Echinodermata: Echinoidea). Mar. Biol. 112, 445-449.
- Chen, J., R. Maxson, P.A. Jones. 1993. Direct induction of DNA hypermethylation in sea urchin embryos by microinjection of 5-methyl-dCTP stimulates early histone gene expression and leads to developmental arrest. Dev. Biol. 155, 75-86.
- Chernyshev, A.V. 1991. Tetrastemma commensalis and Asteronemertes gibsoni gen et sp new (Hoploneimertini, Tetrastemmidae): commensals of starfish. Zooll. Zhur. 70, 34-39.
- Chiba, K., H. Tadenuma, M. Matusumoto, K. Takahashi, T. Katada, M. Hoshi. 1992. The primary structure of the alpha subunit of a starfish guanosine-nucleotide-binding regulatory protein involved in 1-methyladenine-induced oocyte maturation. Eur. J. Biochem. 207, 833-838.
- Cho, Y., R. Higuchi, N. Marubayashi, I. Ueda, T. Komori. 1992. Biologically active glycosides from Asteroidea. 29. X-ray crystallographic analysis of desulfated asterosaponin-P1 isolated

- from the starfish Asterina pectinifera. Liebig's Ann. Chem. 79-81.
- Cole, R.G., Creese, R.G. Grace, P. Irving, B.R. Jackson. 1992. Abundance patterns of subtidal benthic invertebrates and fishes at the Kermadec Islands. Mar. Ecol. Prog. Ser. 82, 207-218. (asteroids, echinoids)
- Coulon, P., M. Jangoux, P. Bulteel. 1992. Respiratory rate and assessment of secondary production in the holothuroid Holothuria tubulosa (Echinodermata) from Mediterranean seagrass beds. P.S.Z.N.I: Mar. Ecol. 13, 63-68.
- Coyer, J.A., R.F. Ambrose, J.M. Engle, J.C. Carroll. 1993. Interactions between corals and algae on a temperate zone rocky reef: mediation by sea urchins. J. Exp. Mar. Biol. Ecol. 167, 21-37.
- Czihak, G., M.K. Kojima. 1992. Nuclear migration and spindle formation in the fourth cleavage of sea urchin eggs under the influence of inhibitors. Cell Struct. Funct. 17, 145-150.
- Dafni, J. 1992. Growth rate of the sea urchin Tripneustes gratilla elatensis. Israel J. Zool. 38, 25-33.
- Dales, R.P. 1992. Phagocyte interactions in echinoid and asteroid echinoderms. J. Mar. Biol. Assoc. U.K. 72, 473-482.
- D'Auria, M.V., L. G. Paloma, L. Minale, R. Riccio, A. Zampella, J.-I. Tanaka, T. Higa. 1992. Isolation, structure characterization and conformational analysis of a unique 4 $\alpha$ -9 $\alpha$ -epoxysteroid sulphate from the Okinawan ophiuroid Ophiomastix annulosa. Tetrahedron Lett. 33, 4641-464.
- Dautov, S.S., S.D. Kashenko, E.S. Kornienko. 1991. Study of the topography of catecholamine-containing neurons in the sea slug Stichopus japonicus larvae during their development. Dokl. Akad. Nauk. SSSR. 320, 723-725.
- David, B., B. Laurin. 1991. Modeles morphometriques en paleontologie evolutive. Geobios. M.S. no. 13. 91-69.
- David, B., B. Laurin. 1991. L'ontogenese complexe du spatangue Echinocardium cordatum: un test des standards des trajectoires heterochroniques. Geobios. no. 24, 569-583.
- Davault, D., N. Degros, M.A. Janmquin, B. Sozey. 1992. Biometrics, carbon and nitrogen content in the ophiuroid Ophiothrix fragilis. J. Mar. Biol. Ass. U.K. 72, 915-918.
- Dayton, P.K., M.J. Tegner, P.E. Parnell, P.B. Edwards. 1992. Temporal and spatial patterns of disturbance and recovery in a kelp forest community. Ecol. Monogr. 62, 421-445. (echinoids)
- Dericcardis, F., M. Iorizzi, L. Minale, R. Riccio, C. Debitus. 1992. The first occurrence of polyhydroxylated steroids with phosphate conjugation from the starfish Tremaster novaecaledoniae. Tetrahedron Lett. 33, 1097-1100.
- De Riccardis, F., M. Iorizzi, L. Minale, R. Riccio, C. Debitus. 1992. The first occurrence of polyhydroxylated steroids with phosphate conjugation from the starfish Tremaster novaecaledoniae. Tetrahedron Lett. 33, 1097-1100.
- Diaz-Miranda, L., D.A. Price, M.J. Greenberg, T.D. Lee, K.E. Doble, J.E. Garcia-Arras. 1992. Characterization of two novel neuropeptides from the sea cucumber Holothuria glaberrima. Biol. Bull. 182, 241-247.
- Diaz-Miranda, L., D.A. Price, M.J. Greenberg, T.D. Lee, K.E. Doble, J.E. Donovan, S.K., E.N. Doyle, D.A.T. Harper. 1992. A

- flexible crinoid from the Llandovery (Silurian) of western Ireland. *J. Paleontol.* 66, 262-266.
- Dolo, V., C. Forti, S. Dell'Utri, G. Gherzi, M.J.L. Vittorelli. 1992. An acid extract from dissociation medium of sea urchin embryos, induces mesenchyme differentiation. *Cell Biol. Int. Rep.* 16, 517-532
- Done, T. 1992. Constancy and change in some Great Barrier Reef coral communities: 1980-1990. *Amer. Zool.* 32, 655-662.  
(Acanthaster)
- Donovan, S.K. 1992. Jamaican Cretaceous Echinoidea (Echinodermata): 3. Scoliechinus axiologus Arnold and Clark, 1927, and an indeterminate cassiduloid. *Proc. Biol. Soc. Wash.* 105, 23-31.
- Donovan, S.K. 1992. New cladid crinoids from the Late Ordovician of Girvan, Scotland. *Palaeontology.* 35, 149-158.
- Donovan, S.K. 1992. Scanning EM study of the living cyrtocrinid Holopus rangii (Echinodermata, Crinoidea) and implications for its functional morphology. *J. Paleontol.* 66, 665-675.
- Donovan, S.K., N.D.L. Clark. 1992. An unusual crinoid columnal morphospecies from the Llandovery of Scotland and Wales. *Palaeontology.* 35, 27-35.
- Drummond, A.E. 1992. Reproduction of the sea urchin Stomopneustes variolaris (Lam.) on the east coast of South Africa. *Invertebr. Reprod. Dev.* 20, 259-265.
- Dunton, K. 1992. Arctic biogeography: the paradox of the marine benthic fauna and flora. *Trends Ecol. Evol.* 7, 183-189.  
(echinoderms)
- Ebert, T.A., M.P. Russell. 1992. Growth and mortality estimates for red sea urchin, Strongylocentrotus franciscanus, from San Nicolas Island, California. *Mar. Ecol. Prog. Ser.* 81, 31-41.
- Eckelbarger, K.J., C.M. Young. 1992. Ovarian ultrastructure and vitellogenesis in ten species of shallow-water and bathyal sea cucumbers (Echinodermata: Holothuroidea). *J. Mar. Biol. Ass. U.K.* 72, 759-781.
- Emura, T., S. Fujimoto, K.p. Takahashi, S. Morisawa, A. Inoue. 1992. Nuclear S1 proteins from the starfish Asterina pectinifera. *Biochem. Int.* 26, 531-535.
- Ellers, O., M. Telford. 1992. Causes and consequences of fluctuating coelomic pressure in sea urchins. *Biol. Bull.* 182, 424-434.
- Eshel, D., C. Shingyoji, K. Yoshimura, I.R. Gibbons, K. Takahashi. 1992. The phase of sperm flagellar beating is not conserved over a brief imposed interruption. *Exp. Cell Res.* 202, 552-555.
- Exposito, J.-Y., M. D'Alessio, F. Ramirez. 1992. Novel amino-terminal propeptide configuration in a fibrilla procollagen undergoing alternative splicing. *J. Biol. Chem.* 267, 17404-17408.
- Exposito, J.-Y., M. D'Alessio, M. Solurshi, F. Ramirez. 1992. Sea urchin collagen evolutionarily homologous to vertebrate pro-alpha2(I) collagen. *J. Biol. Chem.* 267, 15559-15562.
- Fabricius, K.E., F.H. Fabricius. 1992. Re-assessment of ossicle frequency patterns in sediment cores: rate of sedimentation related to Acanthaster planci. *Coral Reefs.* 11, 109-114.

Fenaux, L., Y. De Greef, C. Cellario. 1992. Effets du retard dans la premiere alimentation sur le developpement de la larve planctotrophe de l'oursin Paracentrotus lividus. Int. Revue ges. Hydrobiol. 77, 651-663.

Ferguson, J.C. 1992. The function of the madreporite in body fluid volume maintenance by an intertidal starfish, Pisaster ochraceus. Biol. Bull. 482-489.

Finamore, E., F. Zollo, L. Minala, T. Yasumoto. 1992. Starfish saponins. 47. Steroidal glycoside sulfates and polyhydroxyasteroids from Alphelasteris japonica. J. Nat. Prods. Lloydia. 55, 767-772.

Fisk, D.A. 1992. Recruitment of Acanthaster planci over a five-year period at Green Island Reef. Aust. J. Mar. Freshwater Res. 43, 929-633.

Flammang, P., M. Jangoux. 1992. Functional morphology of the locomotory podia of Holothuria forskali (Echinodermata: Holothuroidea). Zoomorphology. 111, 167-178.

Foote, M. 1992. Paleozoic record of morphological diversity in blastozoan echinoderms. Proc. Natl. Acad. Sci. U.S.A. 89, 7325-7329.

Foster, M.S., A.P. De Vogelaere, J.S. Oliver, J.S. Pearse, C. Harrold. 1991. Open coast intertidal and shallow subtidal ecosystems of the northeast Pacific. In: A.C. Mathieson, P.H. Nienhuis (eds.). Ecosystems of the World., Elsevier, Amsterdam. 24, 235-272.

Foster, M.S. 1992. How important is grazing to seaweed evolution and assemblage structure in the north-east Pacific. In: D.M. John, S.J. Hawkins, J.H. Price (Eds.). Plant-animal interactions in the marine benthos. Clarendon Press, Oxford. pp. 61-85.

Frantzis, A., A. Gremare, G. Vétion. 1992. Growth rates and RNA:DNA ratios in Paracentrotus lividus (Echinodermata: Echinoidea) fed on benthic macrophytes. J. Exp. Mar. Biol. Ecol. 156, 125-138.

Francour, P. 1991. Statut de Centrostephanus longispinus en Mediterranee. In: C.F. Boudouresque et al. (eds.). Les Espèces Marines à Protéger en Méditerranée. 187-202.

Flammang  
+ Jangoux 1992

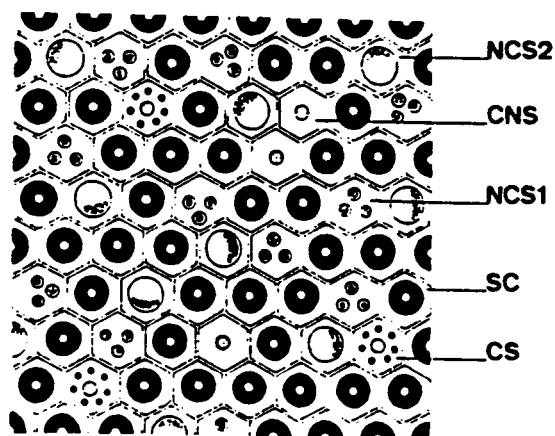


Fig. 14. Diagrammatic representation of a transverse section through the apex of the disc epidermis (not to scale). CNS, ciliated non-secretory cell; CS, ciliated secretory cell; NCS1, type 1 non-ciliated secretory cell; NCS2, type 2 non-ciliated secretory cell; SC, support cell

C.A. Freire et al.: Growth and spatial distribution of *Cassidulus mitis*

Table 1. *Cassidulus mitis*. Total numbers of females, males and individuals of indeterminate sex collected from February 1986 to January 1987, and sex ratio (female:male) as a function of arbitrary size class (gonopores were not detectable at < 10 mm length)

Sex	Test length (mm)					Total
	≤ 10.00	10.05–20.00	20.05–30.00	30.05–40.00	≥ 40.05	
Female	0	58	91			
Male	0	16	83	95	36	280
Indeterminate	88	76	8	67	27	193
Total	88	150	182	162	0	172
Sex ratio	-	3.6	1.1	1.4	1.3	1.5

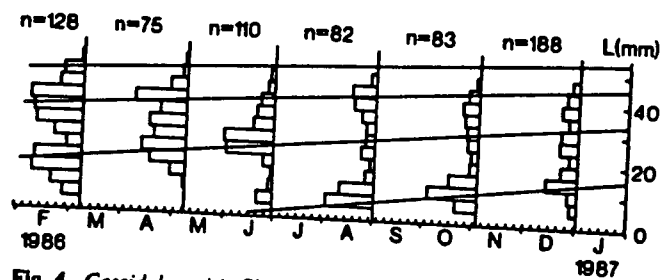


Fig. 4. *Cassidulus mitis*. Size-frequency (length,  $L$ ) distribution over investigation period. Data are grouped bi-monthly, and growth curves of four size-cohorts are superimposed on histograms

Freire, C.A., P.J.P. Santlos, N.F. Fontoura, R.A.O. Magalhaes, P.A. Grohmann. 1992. Growth and spatial distribution of *Cassidulus mitis* (Echinodermata: Echinoidea) on a sandy beach in southeastern Brazil. Mar. Biol. 112, 625-630.

Frid, C.L.J. 1992. Foraging behavior of the spiny starfish *Marthasterias glacialis* ion Lough Ine, Co. Cork. Mar. Behav. Physiol. 19, 227-239.

Fronk, J., G.A. Tank, J.P. Langmore. 1992. DNA methylation pattern changes during development of a sea urchin. Biochem. J. 283, 751-753.

Fujino, Y., A. Fujiwara, I. Yasumasu, T. Fujii. 1992. Chromogranin A-like proteins in the heat-stable fraction of sea urchin eggs, embryos, and the substances secreted with sperm. Zool. Sci. 9, 329-444.

Fujiwara, A., I. Yasumasu. 1992. SCN ion blocks hardening of the fertilization envelope of sea urchin eggs and adhesion of blastomeres. Growth Differ. 34, 309-318.

- Fuhrman, M.H., J.P. Suhan, C.A. Etnensohn. 1992. Developmental expression of echinoectin and endogenous lectin of the sea urchin embryo. *Dev. Growth Differ.* 34, 137-150. Gage, J.D. 1992. Growth bands in the sea urchin Echinus esculentus. Results from tetracycline-mark/recapture. *J. Mar. Biol. Assoc. U.K.* 72, 257-260.
- Gage, J.D. 1992. Natural growth bands and growth variability in the sea urchin Echinus esculentus: results from tetracycline tagging. *Mar. Biol.* 114, 607-616.
- Gale, A.S. S.K. Donovan. 1992. Predatory asteroids and articulate brachiopods: a reply. *Lethaia.* 25, 346-348.
- Ganster, R., Y. Paul, K.S. Katula. 1992. Analysis of the DNA binding proteins interacting with specific upstream sequences of the Strongylocentrotus purpuratus CyI actin gene. *Mol. Reprod. Dev.* 33, 392-406.
- Garcia-Arrijas. 1992. Characterization of two novel neuropeptides from the sea cucumber Holothuria glaberrima *Biol. Bull.* 182, 241-247.
- Garneau, F.-X., C. Harvey, J.-L. Simard, J.W. Apsimon, D.J. Burnell, J.H. Himmelman. 1989. The distribution of asterosaponins in various body components of the starfish Leptasterias polaris. *Comp. Biochem. Physiol.* 92B, 411-416.
- George, S.B., J.M. Lawrence, L. Fenaux. 1991. The effect of food ration on the quality of eggs of Luidia clathrata (Say) (Echinodermata: Asteroidea). *Invertebr. Reprod. Dev.* 237-242.
- Gerdes, D., M. Klages, W.E. Arntz, R.L. Herman, J. Galeron, S. Hain. 1992. Quantitative investigations on macrobenthos communities of the southeastern Weddell Sea shelf based on multibox corer samples. *Polar Biol.* 12, 291-301.
- Geys, J.F. 1992. Regular echinoids, other than Hemicidaroida, from Upper Cretaceous deposits in the Wadi Qena-ara (eastern Desert, Egypt). *Bull. Inst. R. Sci. Nat. Belg. Sci. Terre.* 62, 139-154.
- Gherardi, F. 1991. Eco-ethological aspects of the symbiosis between the shrimp Athanas indicus (Coutiere 1903) and the sea urchin Echinometra mathaei (de Blainville 1825). *Trop. Zool.* 4, 107-128.
- Gilliland, P.M. 1992. Holothurian faunal changes at the Triassic-Jurassic boundary. *Lethaia.* 25, 69-84.
- Gilliland, P.M. 1992. Holothurians in the Blue Lias of southern Britain. *Paleontology.* 35, 159-210.
- Gittings, S.R., T.J. Bright, W.W. Schroeder, W.W. Sager, J.S. Laswell, R. Rezak. 1992. Invertebrate assemblages and ecological features in the Northeast Gulf of Mexico. *Bull. Mar. Sci.* 50, 435-455. (crinoids, ophiuroids, echinoids, holothuroids)
- Gladstone, W. 1992. Observations of crown-of-thorns starfish spawning. *Aust. J. Mar. Freshwater Res.* 43, 535-537.
- Gliksman, N.R., S.F. Parsons, E.D. Salmon. 1992. Okadaic acid induces interphase to mitotic-like microtubule dynamic instability by inactivating rescue. *J. Cell Biol.* 119, 1271-1276. (echinoid)
- Glynn, P.W., M.W. Colgan. 1992. Sporadic disturbances in fluctuating coral reef environments: El Nino and coral reef development in the eastern Pacific. *Amer. Zool.* 32, 707-718.

**(Acanthaster)**

- Gonzalez-Martinez, M.T., A. Guerrero, E. Morales, L. de la Torre, A. Darszon. 1992. A depolarization can trigger calcium uptake and the acrosome reaction when preceded by a hyperpolarization in Lytechinus pictus sea urchin sperm. *Dev. Biol.* 150, 193-202.
- Gordon, D.M., S.K. Donovan. 1992. Disarticulated echinoid ossicles in paleoecology and taphonomy: the last interglacial Falmouth Formation of Jamaica. *Palaios.* 7, 157-166.
- Greenstein, B.J. 1992. Taphonomic bias and the evolutionary history of family Cidaridae (Echinodermata: Echinoidea). *Paleobiology.* 18, 50-79.
- Grissom, P.M., M.E. Porter, J.R. McIntosh. 1992. Two distinct isoforms of sea urchin egg dynein. *Cell Motil. Cytoskeleton.* 21, 281-292.
- Guensburg, T.E. 1992. Paleoecology of hardground encrusting and commensal crinoids, Middle Ordovician, Tennessee. *J. Paleontol.* 66, 129-147.
- Guensburg, T.E. 1992. Glaucocrinus falconeri Parks and Alcock, 1912 (Crinoidea) and its systematic status. *R. Ont. Mus. Life Sci. Occas. Pap.* (39), 1-7.
- Guensburg, T.E., J. Sprinkle. 1992. Rise of echinoderms in the Paleozoic evolutionary fauna: significance of paleoenvironmental controls. *Geology.* 20, 407-410.
- Guillou, M., C. Michel. 1993. Reproduction and growth of Sphaerechinus granularis (Echinodermata: Echinoidea) in southern Brittany. *J. Mar. Biol. Ass. U.K.* 73, 179-192.
- Gustafson, T. 1991. Pharmacological control of muscular activity in the sea urchin larva -- IV. Effects of monoamines and adenosine. *Comp. Biochem. Physiol.* 98C, 307-315.
- Gutt, J. 1991. Are Weddell Sea holothurians typical representatives of the Antarctic benthos. *Meeresforschung.* 33, 312-329.
- Gutt, J., M. Klages. 1991. In situ observations on the genus Bathyploetes (Holothuroidea: Echinodermata) in Antarctica and its relevance to taxonomy. *Zool. Scripta.* 20, 301-306.
- Gutt, J., D. Gerdes, M. Klages. 1992. Seasonality and spatial variability in the reproduction of two Antarctic holothurians (Echinodermata). *Polar Biol.* 11, 533-544.
- Hagen, N.T. 1992. Macroparasitic epizootic disease: a potential mechanism for the termination of sea urchin outbreaks in northern Norway. *Mar. Biol.* 114, 469-478.
- Hagen, N.T. 1992. Macroparasitic epizootic disease: a potential mechanism for the termination of sea urchin outbreaks in northern Norway. *Mar. Biol.* 114, 469-478.
- Hagen, N.T., K.H. Mann. 1992. Functional response of the predators American lobster Homarus americanus (Milne-Edwards) and Atlantic wolfish Anarhichas lupus (L.) to increasing numbers of the green sea urchin Strongylocentrotus droebachiensis (Muller). *J. Exp. Mar. Biol. Ecol.* 159, 89-112.
- Hajduk, S.L. 1992. Ultrastructure of the tube-foot of an ophiuroid echinoderm, Hemipholis elongata. *Tissue Cell.* 24, 111-119.
- Hamel, J.-F., J.H. Himmelman. 1992. Sexual dimorphism in the



- sand dollar Echinarachnius parma. Mar. Biol. 113, 379-383.
- Hardin, J., J.A. Coffman, S.D. Black, D.R. McClay. 1992. Commitment along the dorsalventral axis of the sea urchin embryo is altered in response to nickel chloride. Development. 116, 671-685.
- Harmelin-Vivien, M.L., M. Peyrot-Clausade, J.-C. Romano. 1992. Transformation of algal turf by echinoids and scarid fishes on French Polynesia coral reefs. Coral Reefs. 11, 45-50.
- Harrington, M.J.G., J.A. Coffman, F.J. Calzone, L.E. Hood, R.J. Britten, E.H. Davidson. 1992. Complex of sea urchin embryo nuclear proteins that contain basic domains. Proc. Natl. Acad. Sci. U.S.A. 89, 6252-6256.
- Harris, P.J., E.L. Clason. 1992. Conditions for assembly of tubulin-based structures in unfertilized sea urchin eggs: spirals, monasters and cytasters. J. Cell Sci. 102, 557-567.
- Harumi, T., K. Hoshino, N. Suzuki. 1992. Effects of sperm-activating peptide I on Hemicentrotus pulcherrimus spermatozoa in high potassium sea water. Dev. Growth Differ. 34, 163-172.
- Harumi, T., M. Kurita, N. Suzuki. 1992. Purification and characterization of sperm creatine kinase and guanylate cyclase of the sea urchin Hemicentrotus pulcherrimus. Dev. Growth Differ. 34, 151-162.
- Haude, R. 1992. Scyphocrinoids, the Buoy crinoids in the uppermost Silurian-lowermost Devonian. Paleontogr. Abt. A. Palaeozool. Stratigr. 222, 141-187.
- Hawkins, S.J., R.G. Hartnoll, J.M. Kain (Jones), T.A. Norton. 1992. Plant-animal interactions on hard substrata in the north-east Atlantic. In: D.M. John, S.J. Hawkins, J.H. Price (eds.). Plant-animal interactions in the marine benthos. Clarendon Press, Oxford. pp. 1-32.
- Heip, C., D. Basford, J.A. Craeymeersch, J.-M. Dewarvmez, J. Dorjes, P. De Wildel, G. Duneveld, A. Eleftherious, P.M.J. Herman et al. 1992. Trends in biomass density, and diversity of North Sea macrofauna. J. Mar. Sci. 49, 13-22. (echinoderms)
- Henderson, R.A. 1992. Assessment of crown-of-thorns skeletal elements in surface sediment of the Great Barrier Reef. Coral Reefs. 11, 103-108.
- Henderson, R.A., P.D. Walbran. 1992. Interpretation of the fossil record of Acanthaster planci from the Great Barrier Reef: a reply to criticism. Coral Reefs. 11, 95-101.
- Henry, J.J., K.M. Klueg, R.A. Raff. 1992. Evolutionary dissociation between cleavage, cell lineage and embryonic axes in sea urchin embryos. Development. 114, 931-938.
- Hess, H. 1991. New ophiuroids from the Toarcian of the Swabian Jurassic (Baden-Wuerttemberg). Stuttg. Beitr. Naturkd. Ser. B. (Geol. Palaeontol). (80), 1-11.
- Himmelman, J.H., C. Dutil. 1991. Distribution, population structure and feeding of subtidal seastars in the northern Gulf of St. Lawrence. Mar. Ecol. Prog. Ser. 76, 61-72.
- Himmelman, J.H. 1991. Diving observations of subtidal communities in the northern Gulf of St. Lawrence. Can. Spec. Publ. Fish. Aquat. Sci. 113. pp 319-332. (asteroids, echinoids, holothuroids, ophiuroids)
- Hines, G.A., S.A. Watts, S.A. Sower, C.W. Walker. 1992. Sex

- steroid levels in the testes, ovaries, and pyloric caeca: gametogenesis in the sea star Asterias vulgaris. Gen. Comp. Endocrinol. 87, 451-460.
- Hines, G.A., S.A. Watts, C.W. Walker, P.A. Voogt. 1992. Androgen metabolism in somatic and germinal tissues of the sea star Asterias vulgaris. Comp. Biochem. Physiol. 102B, 521-526.
- Hisanga, S.-I., S. Endo, N. Hirokawa, H. Sakai, J. Pudles. 1992. Ultrastructure of detergent-resistant cytoskeletons in the noncortical domain of sea urchin eggs as revealed by the quick-freeze deep-etch technique. Cell Struct. Funct. 17, 277-285.
- Hobson K.A., H.E. Welch. 1992. Determination of trophic relationships within a Arctic marine food web using  $^{13}C$  and  $^{15}N$  analysis. Mar. Ecol. Prog. Ser. 84, 9-18. (asteroids, echinoid)
- Hodgson, A.N., R.T.F. Bernard. 1992. Spermatozoan structure of eight species of South African holothurians (Echinodermata). J. Morph. 211, 179-186.
- Hoshino, K., T. Shimizu, T. Sendai, T. Harumi, N. Suzuki. 1992. Differential effects of the egg jelly molecules FSG and SAP-I on elevation of intracellular calcium and pH in sea urchin spermatozoa. Dev. Growth Differ. 34, 403-411.
- Ikegami, S., N. Kajiyama, Y. Ozaki, Y. Myotoishi, S. Miyashiro. 1992. Selective inhibition of membrane fusion events in echinoderm gametes and embryos by halenaquinol sulfate. FEBS Letts. 301, 284-286.
- Ikegami, S., N. Kajiyama, Y. Ozaki, Y. Ooe, T. Shimizu, T. Kasahara, D. Uemura, M. Shioda. 1992. Effects of okadaic acid on embryonic development of the starfish Asterina pectinifera. Biosci. Biotechnol. Biochem. 56, 1007-1011.
- Inagaki, F., S.-I. Tate, H. Kubo, M. Hoshi. 1992. A novel difucosylated neutral glycosphingolipid from the eggs of the sea urchin, Hemicentrotus pulcherrimus: II. Structural determination by two-dimensional NMR. J. Biochem. 112, 286-289.
- Inoue, C., M. Kiyomoto, H. Shirai. 1992. Germ cell differentiation in starfish: the posterior enterocoel as the origin of germ cells in Asterina pectinifera. Dev. Growth Differ. 34, 413-418.
- Iorizzi, M., L. Minale, R. Riccio, T. Yasumoto. 1992. Starfish saponins: 48. Isolation of fitenn sterol constituents (six glycosides and nine polyhydroxyxsteroids) from the starfish Solaster borealis. J. Nat. Prod. (Lloydia). 55, 866-867.
- Ishida, Y. 1992. Fossil ophiuroids from the Oligocene Asagai Formation of Iwaki, Fukushima, Japan. Bull. Natl. Sci. Mus. Ser. C. 18, 65-78.
- Ito, T., T. Matsutani, K. Mori, T. Nomura. 1992. Phagocytosis and hydrogen peroxide production by phagocytes of the sea urchin Strongylocentrotus nudus. Dev. Comp. Immunol. 16y, 187-194.
- Jagt, J.W.M. 1992. Campanian-Maastrichtian pelagic crinoids from NE Belgium and SE Netherlands: preliminary observations. Bull. Inst. R. Sci Nat. Belg. Sci. Terre. 62, 155-161.
- Jagus, R., W.-I. Huan, L.J. Hansen, M.A. Wilson. 1992. Changes in rates of protein synthesis and eukaryotic initiation factor-4 inhibitory activity in cell-free translation systems of sea urchin eggs and early cleavage stage embryos. J. Biol. Chem. 267, 15530-15536.

- James, M.K., J.P. Scandol. 1992. Larval dispersal simulations: correlation with the crown-of-thorns starfish outbreaks database. *Aust. J. Mar. Freshwater Res.* 43, 569-582.
- Jans, D., M. Jangoux. 1992. Rejection of intracoelomic invading material by Leptosynapta inhaerens (Echinodermata: Holothuroidea): a process of ecological significance? *Mar. Ecol.* 13, 255-231.
- Jesus Vassquez, M., E. Quinoa, R. Riguera, A. San Martin, J. Darias. 1992. Santiagoside, the first asterosaponin from an Antarctic starfish (Neosmilaster georgianus). *Tetrahedron.* 48, 3739-3746.
- Jensen, P., I. Aagaard, R.A. Burke Jr., P.R. Dando, N.O. Jogensen, A. Kuijpers, T. Laier, S.C.M. O'Hara, R. Schmaljohann. 1992. 'Bubbling reefs' in the Kattegat: submarine landscapes of carbonate-cemented rocks support a diverse ecosystem at methane seeps. *Mar. Ecol. Prog. Ser.* 83, 103-112. (asteroids)
- Jiang, Z.-H., R.R. Schmidt. 1992. Glycosyl imidates: 56. Synthesis of the hexassacharide moiety of pectinioside E. *Liebigs Ann. Chem.* (9), 975-982.
- Johnson, C. 1992. Reproduction, recruitment and hydrodynamics in the crown-of-thorns phenomeon on the Great Barrier Reef: introduction and synthesis. *Aust. J. Mar. Freshwater Res.* 43, 517-523.
- Johnson, C. 1992. Settlement and recruitment of Acanthaster planci on the Great Barrier Reef: questions of process and scale. *Aust. J. Mar. Freshwater Res.* 43, 611-627.
- Johnson, D.B., P.J. Moran, V.J. Baker, C.A. Christie, I.R. Miller, B.A. Millersmith, A.A. Thompson. 1992. An attempt to locate high-density populations of juvenile crown-of-thorns starfish (Acanthaster planci)
- Justice, R.W., G.N. Nagel, C.F. Gottschling, M.F. Damis, E.J. Carroll, Jr. 1992. A 9.6S protein is the third calcium-insoluble component of the sea urchin hyaline layer. *Arch. Biochem. Biophys.* 294, 297-305.
- Kamata, Y., S. Furuya, K. Takei-Mikami, A. Fujiwara, I. Yasumasuj. 1992. Identification of GTP-binding proteins by ADP-ribosylation in the presence of cholera toxin, pertussis toxin and botulinum toxin D in plasma membrane isolated from eggs and embryos of sea urchin. *Dev. Growth Differ.* 34, 311-222.
- Kamimura, S., R. Kamiya. 1992. High-frequency vibration in flagellar axonemes with amplitudes reflecting the size of tubulin. *J. Cell Biol.* 1161, 1443-1454. (echinoid)
- Kammer, T.W., W.I. Ausich. 1992. Advanced cladid crinoids from the Middle Mississippian of the east-central United States: primitive-grade calyces. *J. Paleontol.* 66, 461-480.
- Kamiya, H., K. Muramoto, R. Goto, M. Sakai. 1992. Lectins in the hemolymph of a starfish, Asterina pectinifera: purification and characteristics. *Dev. Comp. Immunol.* 16, 243-250.
- Katow, H., Y. Nakajima. 1992. Behavior and ultrastructure of primary mesenchyme cells at sessile site during termination of cell migration in early gastrulae. *Dev. Growth Differ.* 34, 107-114.
- Katow-Fukul, Y., T. Noce, T. Ueda, Y. Fujiwara, N. Hashimoto, S. Tanaka, T. Higashinakagawa. 1992. Isolation and

- characterization of cDNA coding a spicule matrix protein in Hemicentrotus pulcherrimus micromeres. *Int. J. Dev. Biol.* 36, 353-361.
- Kasyanov, V.L. 1991. Correlation of the reproduction time in bivalves and echinoderms to the dynamics of abiotic environmental factors in Vostok Bay, the Sea of Japan. *Mar. Biol.* (3), 102-105.
- Katow, H., K. Ishida. 1992. Elevation of cyclic AMP-dependent protein kinase activity during migration of primary mesenchyme cell in sand dollar blastulae. *Dev. Growth. Differ.* 34, 529-533.
- Kenchington, R., G. Kelleher. 1992. Crown-of-thorns starfish Management conundrums. *Coral Reefs.* 11, 53-56.
- Keesing, J.K., R.H. Bradbury, L.M. Devantier, M.J. Riddle, G. De'ath. 1992. Geological evidence for recurring outbreaks of the crown-of-thorns starfish: a reassessment from an ecological perspective. *Coral Reefs.* 11, 79-85.
- Keesing, J.K., A.R. Halford. 1992. Field measurement of survival rates of juvenile Acantaster planci: techniques and preliminary results. *Mar. Ecol. Prog. Ser.* 4, 85, 107-114.
- Keesing, J.K., A.R. Halford. 1992. Importance of postsettlement processes for the population dynamics of Acanthaster planci (L.). *Aust. J. Mar. Freshwater Res.* 43, 635-651.
- Keesing, J.K., J.S. Lucas. 1992. Field measurement of feeding and movement rates of the crown-of-thorn starfish Acanthaster planci (L.). *J. Exp. Mar. Biol. Ecol.* 156, 89-104.
- Kerr, A.M., D.R. Norris, P.J. Schupp, K.D. Meyer, T.J. Pitlik. 1992. Range extensions of echinoderms (Asterozoa, Echinozoa and Holothurozoa) to Guam, Mariana Islands. *Micronesica.* 25, 201-216.
- Kerr, A.M., E.M. Stoffel, R.L. Yoon. 1993. Abundance, distribution of holothurozoans (Echinodermata: Holothurozoa) on a windward and leeward fringing coral reef, Guam, Mariana Islands. *Bull. Mar. Sci.*
- Kicha, A.A., A.I. Kalinovskii, V.A. Stonir. 1991. Steroid sulfates from the starfish Lethasterias nanimensis chelifera. *Khim. Prir. Soedin.* (4), 520-523.
- Kikuyama, M., Y. Hiramoto. 1991. Change in intracellular calcium ions upon maturation in starfish. *Dev. Growth Differ.* 33, 633-638.
- Kingsley, P.D., L.M. Angerer, R.C. Angerer. 1993. Major temporal and spatial patterns of gene expression during differentiation of the sea urchin embryo. *Dev. Biol.* 155, 216-234.
- Kishimura, H., K. Hayashi. 1991. Purification and properties of carboxypeptidase A-like enzyme from the starfish Asterias amurensis. *Nippon Suisan Gakkaishi.* 57, 1939-1944.
- Klikushin, V.G. 1991. The importance of crinoids for the Jurassic stratigraphy of the northeastern USSR. *Paleontol. Zh.* (3), 77-84.
- Klikushin, V.G. 1992. Systematics of Pentacrinida (Crinozoa). in: *Questions in paleontology.* St. Petersburg. 122-132.
- Knowlton, N. 1992. Thresholds and multiple stable states in coral reef community dynamics. *Amer. Zool.* 32, 674-682. (Diadema)
- Kong, F., M.-K. Harper, D.J. Faulkner. 1992. Fuscusine, a tetrahydroisoquinoline alkaloid from the seastar Perknaster

- fuscus antarcticus. Nat. Prod. Lett. 1, 71-74.
- Kreimer, D.I., V.Y. Fedoseev, V.M. Grishchenko, T.G. Orlova, A.A. Freidin, N.Y. Orlov. 1992. Water-soluble calcium-calmodulin-binding proteins in embryo of the sea urchin Strongylocentrotus intermedius. Comp. Biochem. Physiol. B. 103, 951-954.
- Kubo, H., G.J. Jiang, A. Irie, M. Morita, T. Matsubara, M. Hoshi. 1992. A novel ceramine trihexoside from the eggs of the sea urchin, Hemicentrotus pulcherrimus. J. Biochem. 111, 726-731.
- Kubiak, J.Z. 1991. Cleavage divisions of bisected sea urchin eggs and zygotes: implications for centrosome role and inheritance. Eur. Arch. Biol. 102, 103-109.
- Kubo, H., G.J. Jiang, A. Irie, M. Suzuki, F. Inagaki, M. Hoshi. 1992. A novel difucosylated neutral glycosphingolipid from the eggs of the sea urchin, Hemicentrotus pulcherrimus: I. Purification and structural determination of the glycolipid. J. Biochem. 112, 281-285.
- Kuraishi, R., K. Osanai. 1992. Cell movements during gastrulation of starfish larvae. Biol. Bull. 183, 258-268.
- Kuznetsov, L.A.P., L.L. Demina, I.P. Shmelev. 1992. Ecological differentiated approach to studying the bioaccumulation of chemical elements and compounds in marine organisms. Doklady Akad. Nauk. 324, 1336-1338. (asteroids, echinoids, holothuroids)
- Kvitek, R.G., J.S. Oliver, A.R. DeGange, B.S. Anderson. 1992. Changes in Alaskan soft-bottom prey communities along a gradient in sea otter predation. Ecology. 73, 413-423.
- Kvitek, R.G., J.S. Oliver. 1992. Influence of sea otters on soft-bottom prey communities in southeast Alaska. Mar. Ecol. Prog. Ser. 82, 103-113.
- Lacalli, T.C. 1993. Ciliary bands in echinoderm larvae: evidence for structural homologies and a common plan. Acta Zoologica. 74, 127-133.
- Lacalli, T.C., J.E. West. 1993. A distinctive nerve cell type common to diverse deuterostome larvae: comparative data from echinoderms, hemichordates and amphioxus. Acta Zool. 74, 1-8.
- Lamash, N.E., Y.S. Khotimkichenko. 1992. Properties of the adenylate cyclase in the oocytes of starfish Asterias amurensis. Comp. Biochem Physiol. C. 103, 379-382.
- Lambert, C.C., D.E. Bataglia. 1993. Loss of the paternal mitochondrion during fertilization. Zool. Sci. 10, 31-38. (includes echinoderms)
- Lawrence, J.M., P. Moran. 1992. Proximate composition and allocation of energy to body components in Acanthaster planci (Linnaeus) (Echinodermata: Asteroidea). Zool. Sci. 9, 321-438.
- Lazzari, C. 1991. Signalling fossil tracts of Asterozoans near Bragarezza in Val Zoldana. Soc. Veneziana Sci. Nat. Lav. 16, 213-261.
- Lebeche, D., B. Kaminer. 1992. Characterization of a calsequestrin-like protein from sea urchin eggs. Biochem. J. 287, 741-747.
- Leclerc, M., M. Bajelan. 1992. Homologous antigen for T cell receptor in axial organ cells from the asterid Acanthaster rubens. Cell Biol. Int. Rep. 16, 487-490.
- Leibson, N.L. 1992. Regeneration of digestive tube in holothurians Stichopus japonicus and Eupentacta fraudatrix. In:

Keys for regeneration. Monogr. Develop. Biol. 23, 51-61.

Lepage, T., C. Sardet, C. Gache. 1992. Spatial expression of the hatching enzyme gene in the sea urchin embryo. Dev. Biol. 150, 23-32.

Leslie, R.J. 1992. Chromosomes attain a metaphase position on half-spindles in the absence of an opposing spindle pole. J. Cell Sci. 103, 125-130.

Levitan, D.R. 1992. Community structure in times past: influence of human fishing pressure on algal-urchin interactions. Ecology. 73, 1597-1605.

Levitan, D.R. 1993. The importance of sperm limitation to the evolution of egg size in marine invertebrates. Amer. Nat. 141, 517-536. (echinoids)

Litvinova, N.M. 1992. Revision of *Ophiotholia* (Echinodermata, Ophiuroidea). Zool. Zh. 71, 47-57.

McGehee, M.A. 1992. Distribution and abundance of two species of *Echinometra* (Echinoidea) on coral reefs near Puerto Rico. Carib. J. Sci. 28, 173-183.

McNamara, K.J. 1992. Geographical and stratigraphical distribution of the echinoid *Echinometra mathaei* (Blainville) in Western Australia. Rec. West Aust. Mus. 16, 79-86.

Maekawa, S., M. Toriyama, H. Sakai. 1992. A novel 24-kDa microtubule-associated protein purified from sea urchin eggs. Eur. J. Biochem. 204, 1195-1200.

Malatesta, R.J., P.J. Auster, B.P. Carlin. 1992. Analysis of transect data for microhabitat correlations and faunal patchiness. Mar. Ecol. Prog. Ser. 87, 189-195. (asteroids)

Mallyla, S.K., J.S. Partin, M.C. Valdizan, W.J. Lennarz. 1992. Proteolysis of the major yolk glycoproteins is regulated by acidification of the yolk platelets in sea urchin embryos. J. Cell Biol. 117, 1211-1222.

Manni, R., U. Nicosia, M. Pilgliucci. 1991. Computer simulation of the behaviour of Jurassic crinoid larvae. Geol. Rom. 27, 437-447.

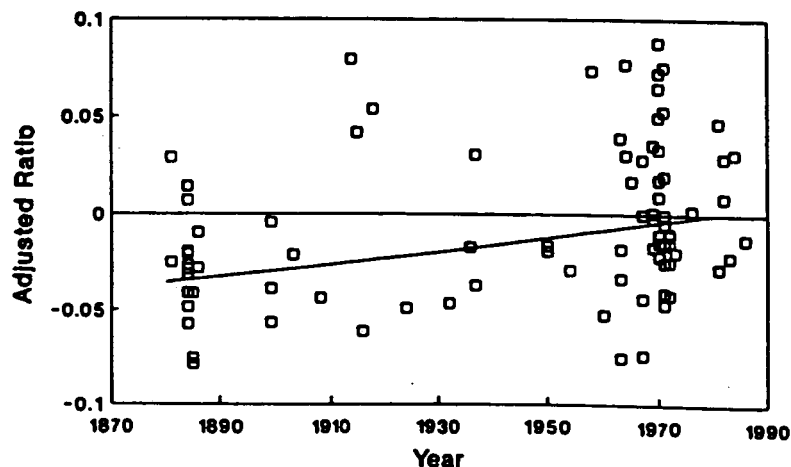


FIG. 2. *Diadema antillarum* demipyramid/test length ratio as a function of time. Ratios are allometrically adjusted (see Table 1). Regression equation:  $\ln \text{ adjusted ratio} = 0.00435\text{time} - 10.923$ ,  $R^2 = 0.114$ ,  $P < .002$ . (Time is in years and 0.1 is added to the ratio to remove negative numbers and allow for a natural logarithmic transformation.)

Levitan 1992

Markel, K., U. Roser. 1992. Functional anatomy of the valves in the ambulacral system of sea urchins (Echinodermata: Echinoidea). *Zoomorphology*. 111, 179-192.

Markel, K., U. Mackenstedt, U. Roser. 1992. The sphaeridia of sea urchins: ultrastructure and supposed function (Echinodermata: Echinoidea). *Zoomorphology*. 112, 1-10.

Markov, A.V. 1992. The most deep-sea representative of the family Schizasteridae (Echinoidea). *Zool. Zh.* 70, 153-155.

Marquet, P.A., S.A. Navarrete, J.C. Castilla. 1990. Scaling population density to body size in rocky intertidal communities. *Science*. 250, 1125-1127. (asteroids, echinoids)

Marshall, D.J., A.N. Hodgson, R.A. Pretorius. 1991. New southern geographical records of intertidal sea urchins (Echinodermata: Echinoidea), with notes on abundance. *S. Afr. J. Zool.* 26, 204-205.

Marshall, C.R. 1992. Character analysis and the integration of molecular and morphological data in an understanding of sand dollar phylogeny. *Mol. Biol. Evol.* 9, 309-322.

Marshall, C.R., H. Swift. 1992. DNA-DNA hybridization phylogeny of sand dollars and highly reproducible extent of hybridization

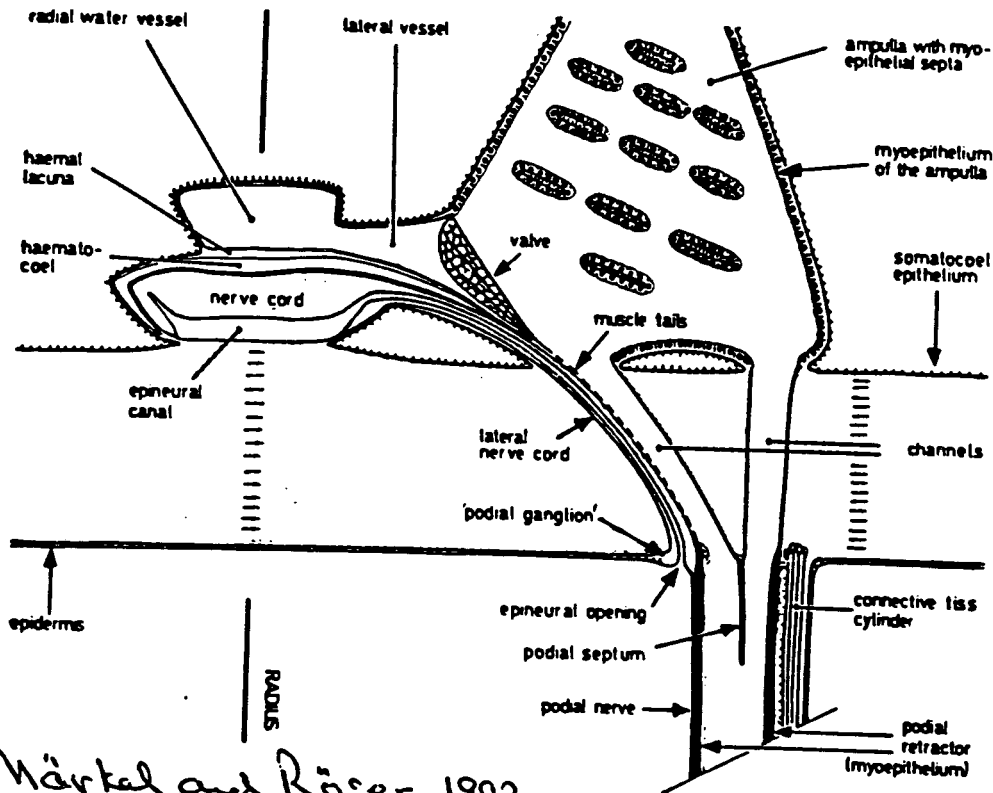


Fig. 11. Schematic representation of the radial organ complex with lateral branches to the right. The pore slit lies in the drawing plane and is not shown. The fact that the lateral nerve cord turns for about 90° around the perradial channel is ignored. Take note of the connection between the podial retractor muscle and the muscle tails (broken line) opposite to the 'podial ganglion'. Hatched the test plates. Not to scale

Markel and Roser 1992

values. *J. Mol. Evol.* 34, 31-44.

Marshall, D.J., A.N. Hodgson, R.A. Pretorius. 1991. New southern geographical records of intertidal sea urchins (Echinodermata: Echinoidea), with notes on abundance. *S.-Afr. Tydskr. Dierk.* 26, 204-206.

Martinez V., J. C. 1989. Atribucion inequivoca de las senales de RMN de un neolignano arilnaftalenico utilizando espectros bidimensionales. *REV. Col. Quim.* 18, 27-36. ((Echinaster)

Marty, P., Y. Martin. 1992. Bacteries heterotrophes aerobies isolees d'invertebres benthiques des eaux cotieres mediterraneennes: caracteristiques des souches, production d'exoenzymes et d'agents antibacteriens. *Mar. Life.* 1, 1-8. (Paracentrotus lividus, Holothuria tubulosa)

Massey, C.B., Jr., S.A. Watts. 1992. Patterns of sperm-specific histone variation in sea stars and sea urchins: primary structural homologies in the N-terminal region of spermatogenic H1. *J. Exp. Zool.* 262, 9-15.

Massin, C. 1992. Holothurians (Echinodermata) from Marion and Prince Edward Islands: new and little-known species. *Zool. Scripta.* 21, 311-324.

Massin, C. 1992. Three new species of Dendrochirotida (Holothuroidea, Echinodermata) from the Weddell Sea (Antarctica). *Bull. Inst. Roy. Sci. Nat. Belg. Biol.* 62, 179-191.

Matsuno, A., T. Motokawa. 1992. Evidence of calcium translocation in catch connective tissue of the sea cucumber Stichopus chloronotus. *Cell Tissue Res.* 267, 307-312.

Matsuoka, N., M. Inamori, M. Sugawara. 1992. High genetic variability in the starfish Distolasterias nippon revealed by enzyme electrophoresis. *Comp. Biochem. Physiol.* 104B, 75-79.

Matsuoka, N. 1992. Phylogenetic relationships of echinoderms deduced from kinetic similarity of glucose-6-phosphate dehydrogenase. *Comp. Biochem. Physiol.* 103B, 133-137.

Mayhook, A.G., A.-M. Rinaldi, H.T. Jacobs. 1992. Replication origins and pause sites in sea urchin mitochondrial DNA. *Proc. R. Soc. Lond. Ser. B. Biol. Sci.* 248, 85-94.

McCallum, H. 1992. Completing the circle: stock-recruitment relationships and Acanthaster. *Aust. J. Mar. Freshwater Res.*, 43, 653-662.

McCulloh, D.M., E.L. Chambers. 1992. Fusion of membranes during fertilization: increases of the sea urchin egg's membrane capacitance and membrane conductance at the site of contact with the sperm. *J. Gen. Physiol.* 99, 137-175.

McDonald, G.D., L. Davidson, G.B. Kitto. 1992. Amino acid sequence of the coelomic C globin from the sea cucumber Caudina arenicola. *J. Protein Chem.* 11, 1 29-37.

McEdward, L.R. 1992. Morphology and development of pelagic larva in the starfish Pteraster tesselatus (Echinodermata: Asteroidea). *Biol. Bull.* 182, 177-187.

McGehee, M.A. 1992. Distribution and abundance of two species of Echinometra (Echinoidea) on coral reefs near Puerto Rico. *Carib. J. Sci.* 28, 173-183.

McKinney, M.L., K.J. McNamara, B.D. Carter, S.K. Donovan. 1992. Evolution of Paleogene echinoids: A global and regional view. In: D. Prothero, Berggren (eds.). *Eocene-Oligocene Climatic and*



- Biotic Evolution. Princeton Univ. Press. pp. 349-367.
- McNamara, K.J. 1990. Echinoids. In: K.J. McNamara (ed.). Evolutionary trends. pp. 205-231. Belhaven, London. McPherson, S.M., P.S. McPherson, L. Mathews, K.P. Cambell, F.J. Longo. 1992. Cortical localization of a calcium release channel in sea urchin eggs. *J. Cell Biol.* 116, 1111-1121.
- McNamara, K.J. 1991. Murder and mayhem in the Miocene. *Natural History*. 8/91. 41-46. (echinoid slaughter)
- McNamara, K.J. 1992. Geographical and stratigraphical distribution of the echinoid Echinometra mathaei (Blainville) in Western Australia. *Rec. West. Aust. Mus.* 16, 79-86.
- McMillan, W.O., R.A. Raff, S.R. Palumbi. 1992. Population genetic consequences of developmental evolution in sea urchins (genus Heliocidaris). *Evolution*. 46, 1299-1312.
- Menge, B.A. 1992. Community regulation: under what conditions are bottom-up factors important on rocky shores. *Ecology*. 73, 755-765. (asteroids, echinoids)
- Mespoulhe, P., B. David. 1992. Strategie de croissance d'un oursin subantarctique: Abatus cordatus des iles Kerguelen. *C.R. Acad. Sci. Paris*. 314. Ser. III, 205-211.
- Mita, M. 1992. Involvement of cyclic adenosine 3'-5'-monophosphate in methylation during 1-methyladenine production by starfish ovarian follicle cells. *Gen. Comp. Endocrinol.* 87, 54-62.
- Mita, M. 1992. Diacyl choline phosphoglyceride: the endogenous substrate for energy metabolism in sea urchin spermatozoa. *Zool. Sci.* 9, 563-568.
- Mita, M., M. Nakamura. 1993. Phosphatidylcholine is an endogenous substrate for energy metabolism in spermatozoa of sea urchins of the Order Echinoidea. *Zool. Sci.* 10, 73-84.
- Mita, M., M. Nakamura. 1992. Ultrastructural study of an endogenous energy substrate in spermatozoa of the sea urchin Hemicentrotus pulcherimus. *Biol. Bull.* 182, 298-304.
- Mitsunaga-Nakatsubo, K., A. Fujiwara, I. Yasumasu. 1992. Change in the activity of sodium, potassium-ATPase in embryos of the sea urchin, Hemicentrotus pulcherrimus, during early development. *Dev. Growth Differ.* 34, 379-385.
- Mogami, Y., K. Watanabe, C. Ooshima, A. Kawano, S.A. Baba. 1992. Regulation of ciliary movement in sea urchin embryos: dopamine and 5-HT change the swimming behaviour. *Comp. Biochem. Physiol.* 101C, 251-254.
- Momma, H., Y. Agatsuma, M. Sawazaki. Migration with the passage of time and dispersion in the cultured seeds of the sea urchin. *Nippon Suisan Gakkaishi*. 58, 1437-1442.
- Moore, S.J., M.C. Thorndyke. 1992. The immunocytochemical mapping of the salmfamide neuropeptides in the echinoderm Asterias rubens. *Reg. Peptides*. 39, 277.
- Moran, P.J. 1992. Preliminary observations of the decomposition of crown-of-thorns starfish, Acanthaster planci (L.). *Coral Reefs*. 11, 115-118. Moran, P.J., G. De'ath. 1992. Suitability of manta tow techniques for estimating relative and absolute abundances of crown-of-thorn starfish (Acanthaster planci L.) and corals. *Aust. J. Mar. Freshwater Res.* 43, 357-378.
- Moran, P.J., G. De'ath, V.J. Baker, D.K. Bass, C.A. Christie,

- I.R. Miller, B.A. Miller-Smith, and A.A. Thompson. 1992. Pattern of outbreaks of crown-of-thorns starfish (Acanthaster planci L.) along the Great Barrier Reef since 1966. *Aust. J. Mar. Freshwater Res.* 43, 555-568.
- Morris, S.C. 1993. The fossil record and the early evolution of the Metazoa. *Nature.* 361, 219-. (echinoderms)
- Multigner, L., J. Gagnon, A. Van Dorsselaer, D. Job. 1992. Stabilization of sea urchin flagellar microtubules by histone H1. *Nature.* 360, 33-39.
- Munk, J.E. 1992. Reproduction and growth of green urchins Strongylocentrotus droebachiensis (Mueller) near Kodiak, Alaska. *J. Shellfish Res.* 11, 245-254.
- Nabata, S.-I., E. Abe, M. Kakiuchi. 1992. On the 'Isoyake' condition in Taisei-cho, southwestern Hokkaido. *Sci. Repts. Hokkaido Fish. Exp. Sta.* (38), 1-14.
- Nakamura, S., R. Kagotani, H. Fujisaki, M.K. Kojima. 1992. The acid-insoluble organic matrix of spicules in the sea urchin, Hemicentrotus pulcherrimus. *Zool. Sci.* 9, 741-747.
- Nasir, A., S.D. Reynolds, P.C. Keng, L. M. Angerer, R.C. Angerer. 1992. Centrifugal elutriation of large fragile cells: isolation of RNA from fixed embryonic blastomeres. *Anal. Biochem.* 203, 22-26.
- Nebelsick, J.H. 1992. Components analysis of sediment composition in Early Miocene temperate carbonates from the Austrian Paratethys. *Palaeogeog., Palaeoclim., Palaeoecol.* 91, 59-69.
- Nebelsick, J.H. 1992. Echinoid distribution by fragment identification in the Northern Bay of Safaga, Red Sea. *Palaios.* 7, 316-328.
- Neira, C., M. Silva, M. Iorizi, L. Minale. 1992. Marine organic chemistry. 5. Asterosaponins from the starfish Heliaster helianthus. *Bol. Soc. Chilena Quim.* 37, 139-142.
- Neira O., R., R. Pardo A., R. Cantera. 1991. Equinodermos de la costa Pacifica del Valle del Cauca. *Mem. I Simp. Nac. de la Fauna del Valle del Cauca. Inst. Vallecaucano de Investigaciones Cientificas. INCIVA. Cali.* 41-63.
- Nemoto, S., K. Yamamoto, N. Hashimoto. 1992. A nuclear extract, prepared from mass-isolated germinal vesicles, retains a factor able to sustain a cytoplasmic cycle of starfish oocytes. *Dev. Biol.* 151, 485-490.
- Neraudeau, D. 1992. Transgressions-regressions and echinoid morphoclines. *Lethaia.* 25, 219-220.
- Nickell, T.D., P.G. Moore. 1992. The behavioural ecology of epibenthic scavenging invertebrates in the Clyde Sea Area: laboratory experiments on attractions to bait in moving water, underwater TV observations in situ and general conclusions. *J. Exp. Mar. Biol. Ecol.* 159, 15-35. (ophiuroid)
- Nicosia, U. 1991. Mesozoic crinoids from the northwestern Turkey. *Geol. Rom.* 27, 389-436.
- Nisson, P.E., M.F. Gaudette, B.P. Brandhorst, W.R. Crain. 1992. Mutually exclusive expression of the Strongylocentrotus purpuratus *Spe1* gene and its Lytechinus pictus homologue in cells of hybrid embryos. *Development.* 114, 193-201.
- Norris, D. 1992. The distribution of two irregular echinoids

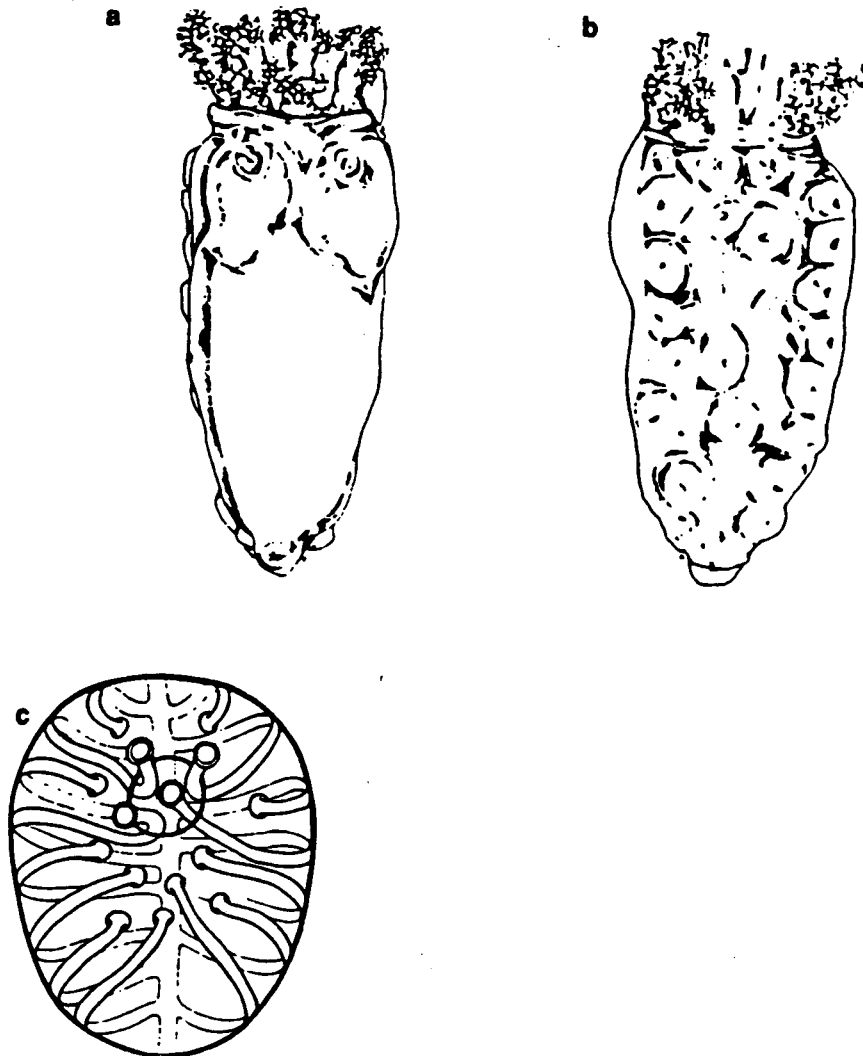
(*Maretia planulata* Lamarck and *Metalia dicrana* Clark) in Agra Harbor, Guam. *Galaxea*. 10, 89- 95.

Ohba, M., T. Mukaihira, T. Jujii. 1992. Synthesis of 5-arylthio-3-methyl-L- histidine, a model for the starfish alkaloid imbricatine. *Heterocycles*. 33, 21-26.

Okinaga, T., Y. Ohashi, M. Hoshi. 1992. A novel saccharide structure, Xyl-1- 3-Gal-1-(sulfate)<sub>3,4</sub>-Fuc, is present in acrosome reaction-inducing substance of the starfish, *Asterias amurensis*. *Biochem. Biophys. Acta*. 186, 405-410 .

O'Loughlin, P.M., T.D. O'Hara. 1992. New cucumariid holothurians (Echinodermata) from southern Australia, including two brooding and one fissiparous species. *Mem. Mus. Victoria*. 53, 227-266.

NEW HOLOTHURIANS FROM SOUTHERN AUSTRALIA



O'LOUGHLIN & O'HARA 1992

Omoto, C.K. 1992. Sea urchin axonemal motion supported by fluorescent, ribose-modified analogues of ATP. *J. Muscle Res. Cell Motil.* 13, 635-639.

Ookata, K., S.-I. Hisanaga, T. Okano, K. Tachibana, T. Kishimoto. 1992. Relocation and distinct subcellular localization of p34-cyclin B complex at meiosis reinitiation in starfish oocytes. *Eur. Mol. Biol. Organ.* 11, 1763-1772.

Page, L., S. Benson. 1992. Analysis of competence in cultured sea urchin micromeres. *Exp. Cell Res.* 203, 305-311.

Palumbi, S.R., B.D. Kessing. 1991. Population biology of the trans-Arctic exchange: MtDNA sequence similarity between Pacific and Atlantic sea urchins. *Evolution.* 45, 1790-1805.

Pandolfi, J.M. 1992. A palaeobiological examination of the geological evidence for recurring outbreaks of the crown-of-thorns starfish, *Acanthaster planci* (L.). *Coral Reefs.* 11, 87-93.

Parsley, R.L. 1991. Review of selected North American mitrate stylophorans (Homalozoa: Echinodermata). *Bull. Am. Paleontol.* 100, 5-54.

Pawlik, J.R. Chemical ecology of the settlement of benthic marine invertebrates. *Oceanogr. Mar. Biol. Annu. Rev.* 30, 273-335. (includes echinoderms)

Pawson, D.L., Y. Liao. 1992. Molpadiid sea cucumbers of China, with descriptions of five new species (Echinodermata: Holothuroidea). *Proc. Biol. Soc. Wash.* 105, 373-388.

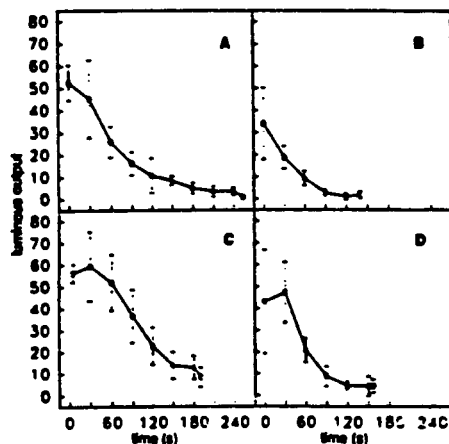
Pawson, D.L., J.E. Miller. 1992. *Phyllophorus* (*Urodemella*) *arenicola*, a new sublittoral sea cucumber from the southeast United States (Echinodermata: Holothuroidea). *Proc. Biol. Soc. Wash.* 105, 483-489.

Pedrotti, M.L., L. Fenaux. 1992. Dispersal of echinoderm larvae in a geographical area marked by upwelling (Ligurian Sea, NW Mediterranean). *Mar. Ecol. Prog. Ser.* 86, 217-227.

Perason, T.H., R. Rosenberg. 1992. Energy flow through the SE Kattegat: a comparative examination of the eutrophication of a coastal marine ecosystem. *Neth. J. Sea Res.*, 28, 317-334.

Pergent-Martini, C., P. Bulteel, P. Francour, M.C. Gambi, M. Harmelin-Vivien, M. Lorenti, L. Mazzella, G. Pergent, J. Romero, G. Russo, J.L. Sanchez-Lizaso. Signalisations de *Centrostephanus longispinus* autour d'Ile d'Ischia (Italie). In: C.F. Boudouresque et al. (eds.). *Les Expeces Marines a Proteger en Mediterranee.* 203-207.

Pickart, C.M., R.G. Summers, H. Shim, E.M. Kasperek. 1991.



Robison 1992

Figure 3. Video image-analysis plots of the decay rate of luminescent output over time. Luminescent output is measured as the mean number of individual light sources within four 100 cm<sup>2</sup> areas on the face of the experimental vessel. The error bars are standard deviations for the four areas. (A) is the result of the initial stimulation of specimen #1; (B) is the re-stimulation of specimen #1, after all the light particles from the initial stimulation had gone dark. (C) & (D) are the initial and re-stimulation decay plots of a second specimen.

- Dynamics of ubiquitin pools in developing sea urchin embryos. *Dev. Growth Differ.* 33, 587-598.
- Porter, J.W., O.W. Meier. 1992. Quantification of loss and change in Floridian reef coral populations. *Amer. Zool.* 32, 625-640. (*Diadema*)
- Pollyakov, V.Y., D. Fais, I.I. Kireyev, O.A. Oliva, A.M. Rinaldi, I. Salcher. 1992. The dynamics of structural modification of mitochondria at the early stages of sea urchin embryonic development. *Cell Biol. Int. Rep.* 16, 155-163.
- Prince, J.S., W.G. Leblanc. 1992. Comparative feeding preference of *Strongylocentrotus droebachiensis* (Echinoidea) for the invasive seaweed *Codium fragile* ssp. *tomentosoides* (Chlorophyceae) and four other seaweeds. *Mar. Biol.* 113, 159-163.
- Prokop, R.J., P. Vaclav. 1991. *Torrocrinus grandis*, new genus new species (Crinoidea) from the lower Devonian Koneprusy limestone of the Barrandian area (Czechoslovakia). *Vestn. Ustred Ustavu. Geol.* 66, 365-368.
- Punnett, T., R.L. Miller, B.-H. Yoo. 1992. Partial purification and some chemical properties of the sperm chemoattractant from the forcipulate starfish *Pycnopodia helianthoides* (Brandt, 1835). *J. Exp. Zool.* 262, 87-96.
- Quest, A.F.G., J.K. Chadwick, D.D. Wothe, R.A. J. McIlhinney, B.M. Shapiro. 1992. Myristoylation of flagellar creatine kinase in the sperm phosphocreatine shuttle is linked to its membrane association properties. *J. Biol. Chem.* 267, 14080-15085.
- Rappaport, R. 1992. Cleavage inhibition by cell shape change. *J. Exp. Zool.* 264, 26-31. (echinoid)
- Raff, R. 1992. Direct-developing sea urchins and the evolutionary reorganization of early development. *BioEssays.* 14, 211-218.
- Reimer, C.L., B.J. Crawford, T.J. Crawford. 1992. Basement membrane lectin binding sites are decreased in the esophageal endoderm during the arrival of presumptive muscle mesenchyme in the developing asteroid *Pisaster ochraceus*. *J. Morphol.* 21, 291-303.
- Reynolds, S.D., L.M. Angerer, J. Palis, A. Nasir, R.C. Angerer. 1992. Early mRNAs, spatially restricted along the animal-vegetal axis of sea urchin embryos, include one encoding a protein related to tolloid and BMP-1. *Development.* 114, 769-786.
- Reunov, A.A., A.L. Drozdov. 1991. The ultrastructure of spermatogonia, spermatocytes, and spermatids of the Far East sea cucumber. *Tsitologiya.* 33, 27-32.
- Rickards, R.B. 1993. Echinodermata. In: M.J. Benton, M.A. Whyte (eds.). *The fossil record.* Chapman and Hall.
- Roa, R. 1992. Design and analysis of multiple-choice feeding-preference experiments. *Oecologia.* 89, 509-515. (echinoids)
- Robinson, B..H. 1992. Bioluminescence in the benthopelagic holothurian *Enypniastes eximia*. *J. Mar. Biol. Ass. U.K.* 72, 463-472.
- Robinson, J.J., 1992. Purification and characterization of a 32-kDa protein that localizes to the sea urchin extraembryonic matrix, the hyaline layer. *Biochem. Cell Biol.* 70, 623-628.
- Robinson, J.J., D. Hall, C. Brenna, P. Kean. 1992. Hyalin, a sea urchin extraembryonic matrix protein: relationship between

- calcium binding and hyalin gelation. Arch. Biochem. Biophys. 298, 129-134.
- Roch, P., C. Canicatti, S. Sammarco. 1992. Tetrameric structure of the active phenoloxidase evidenced in the coelomocytes of the echinoderm Holothuria tubulosa. Comp. Biochem. Physiol. 102B, 349-355.
- Roman, J., A. Vadet, A. Boullier. 1991. Echinoides et brachiopodes de la limite Jurassique-Cretace a Canjuers (Var, France). Rev. Paleobiol. 10, 21-27.
- Roman, J., J Le Renard. 1992. Diversite et variabilite de la forme dans deux populations de Cassiduloides (Echinoidea) du Lutetien de Grignon (Yvelines, France). Geobios. Mem. Spec. 13, 105-112.
- Roman, J., B. Cahuzac. 1992. Une riche faune d'echinoides de l'Eocene d'Angoume (Sud-Aquitaine, France). Geobios Mem. Spec. 14, 113-121.
- Roman, J., J.-P. Platel, J. Roger. 1992. Echinoides cretaces et paleogenes du Sud-Est de l'Arabie (Dhofar, Sultanat d'Oman). Actes 115e Congr. natn. Soc. sav., Avignon, Sect. Sci. (3e Coll. Geol. afric.). 71-87.
- Rosenberg, R., B. Hellman, B. Johansson. 1991. Hypoxic tolerance of marine benthic fauna. Mar. Ecol. Prog. Ser. 79, 127-131. (Ophiuroids)
- Rozhnov, S.V., A.B. Fedorov, T.A. Sayutina. 1992. Lower Cambrian Echinodermata on USSR territory. Paleontol. Zh. (1), 53-66.
- Rumohr, H., P. Krost. 1991. Experimental evidence of damage to benthos by bottom trawling with special reference to Artica islandica. Meeresforschung. 33, 340-345. (Asterias).
- Sakashita, H. 1992. Sexual dimorphism and food habits of the clingfish, Diademichthys lineatus and its dependence on the host sea urchin. Env. Biol. Fish. 34, 95-101.
- Salata, L. L. I Wase, K. Kato, K. Ina, Y. Machiguchi. 1991. A simple feeding inhibitor assay for marine herbivorous gastropods and the sea urchin Strongylocentrotus intermedius and its application to unpalatable algal extracts. Nippon Suisan Gakkaishi. 57, 261-265.
- Sakata, K., Y. Iwase, K. Ina, D. Fujita. 1991. Halogenated terpenes isolated from the red alga Plocamium leptophyllum as feeding inhibitors for marine herbivores. Nippon Suisan Gakkaishi. 57, 743-746. (Strongylocentrotus)
- Sanghera, J. S., L.A. Charlton, H.B. Paddon, S.L. Pelech. 1992. Purification and characterization of echinoderm casein kinase II: Regulation by protein kinase C. Biochem. J. 283, 829-837.
- Sardet, C., I. Gillot, A. Ruscher, P. Payan, J.-P. Girard, G. De Renzis. 1992. Ryanodine activates sea urchin eggs. Dev. Growth Differ. 34, 37-42.
- Sawada, M.T., T. Someno, M. Hoshi, H. Sawada. 1992. Participation of 650-kDa protease (20s-proteasome) in starfish oocyte maturation. Dev. Biol. 150, 414-418.
- Scandol, J.P., M.K. James. 1992. Hydrodynamics and larval dispersal: a population model of Acanthaster planci on the Great Barrier Reef. 43, 583-596.
- Schatten, H., M. Walter, H. Biessmann, G. Schatten. 1992.

- Activation of maternal centrosomes in underfertilized sea urchin eggs. *Cell Motil. Cytoskeleton.* 23, 61-70.
- Schinner, G.O., J.B. McClintock. 1993. Structural characteristics of marsupial brood pouches of the Antarctic sea urchins Abatus nimrodi and Abatus shackletoni (Echinoidea: Spatangoidea). *J. Morphol.* 216-79-93.
- Schoppe, S. 1991. Echinometra lucunter (Linnaeus) (Echinoidea, Echinometridae) as host of a complex association in the Caribbean SEa. *Helgoland. Meeresunters.* 45, 373-379.
- Schubert, J.K., D.J. Bottjer, M.J. Simms. 1992. Paleobiology of the oldest known articulate crinoid. *Lethaia.* 25, 97-110.
- Shulman, M.J. 1990. Aggression among sea urchins on Caribbean coral reefs. *J. Exp. Mar. Biol. Ecol.* 140, 197-207.
- Scott, L.B., W.J. Lennarz, R.A. Raff, G.A. Wray. 1990. The "lecithotrophic" sea urchin Heliocidaris erythrogramma lacks typical yolk platelets and yolk glycoproteins. *Dev. Biol.* 138, 188-193.
- Seike, Y., H. Shibata, T. Suiyemitsu. 1992. Purification of a sperm lectin extracted from spermatozoa of the sea urchin Hemicentrotus pulcherrimus. *Dev. Growth Differ.* 34, 285-291.
- Sewell, M.A. 1992. Reproduction of the temperate aspidochirote Stichopus mollis (Echinodermata: Holothuroidea) in New Zealand. *Ophelia.* 35, 103-122.
- Sewell, M.A., D.R. Levitan. 1992. Fertilization success during a natural spawning of the dendrochirote sea cucumber Cucumaria miniata. *Bull. Mar. Sci.* 51, 161-162.
- Shick, J.M., W.C. Dunlap, B.E. Chalker, A.T. Banaszak, T.K. Rosenzweig. 1992. Survey of ultraviolet radiation-absorbing mycosporine-like amino acids in organs of coral reef holothuroids. *Mar. Ecol. Prog. Ser.* 90, 139-148.
- Shin, S. 1992. A systematic study on the Ophiuroidea in Korea. I. Species from the sea of Japan and the Korea Strait. *Korean J. Syst. Zool.* 8, 107-131.
- Shin, S. 1992. A systematic study on the Ophiuroidea in Korea. II. Cheju Island. *Korean J. Zool.* 35, 350-361.
- Sluder, G., F.J. Miller, K. Lewis. 1992. Centrosome inheritance in starfish zygotes II: selective suppression of the maternal centrosome during meiosis. *Dev. Biol.* 155, 58-67.
- Smith, A.B. 1992. Echinoderm phylogeny: morphology and molecules approach accord. *Trends Ecol. Evol.* 7, 224-229.
- Smith, A.B., B. Lafay, R. Christen. 1992. Comparative variation of morphological and molecular evolution through geologic time - 28S and RNA morphology in echinoids. *Phil. Trans. Roy. Soc. London. B.* 338, 265-282.
- Soliman, F. El-S. 1991. Studies on Egyptian Echinodermata: Ophiocoma aegyptica new species (Ophiuroidea: Ophiocomidae), from the Red Sea. *Galaxea.* 10, 79-88.
- Sousa, M., P. Moradas-Ferreira, C.A. Zevedo. 1992. Presence of a trypsin-like protease in starfish sperm acrosome. *J. Exp. Zool.* 261, 349-354.
- Sprinkle, J. 1992. Radiation of echinoderms. In: J.H. Lipps, P.W. Signor (eds.). *Origin and early evolution of the Metazoa.* Plenum Press. 375-398.
- Starr, M., J.H. Himmelman, J.-C. Therriault. 1992. Isolation

- and properties of a substance from the diatom Phaeodactylum tricornutum which induces spawning in the sea urchin Strongylocentrotus droebachiensis. Mar. Ecol. Prog. Ser. 79, 275-287.
- Stabili, L., P. Pagliara, M. Metrangolo, C. Canicatti. 1992. Comparative aspects of Echinoidea cytolytins: the cytolytic activity of Spherechinus granularis (Echinoidea) coelomic fluid. Comp. Biochem. Physiol. 101A, 553-556.
- Stephens, R.E. 1992. Tubulin in sea urchin embryonic cilia: post-translational modifications during regeneration. J. Cell Sci. 101, 837-845.
- Stevenson, J.P. 1992. A possible modification of the distribution of the intertidal seastar Patiriella exigua (Lamarck) (Echinodermata: Asteroidea) by Patirella calcar (Lamarck). J. Exp. Mar. Biol. Ecol. 155, 41-54.
- Steinberg, P.P., I. van Altema. 1992. Tolerance of marine invertebrate herbivores to brown algal phlorotannins in temperate Australasia. Ecol. Monogr. 62, 189-222.
- Stickle, W.B., D.W. Foltz, M. Katoh, H.L. Nguyen. 1992. Genetic structure and mode of reproduction in five species of sea stars (Echinodermata: Asteroidea) from the Alaskan coast. Can. J. Zool. 70, 1723-1728.
- Strathmann, R.R., L. Fenaux, M.F. Strathmann. 1992. Heterochronic developmental plasticity in larval sea urchins and its implications for evolution of nonfeeding larvae. Evolution. 46, 972-986.
- Sumrall, C.D. 1992. Spiraclavus nacoensis, new genus new species of clavate agelacrinitid edrioasteroid from central Arizona. J. Paleontol. 66, 90-98.
- Svetashev, V.I., V.S. Levin, C. Ngok Lam, D. Tuet Nga. 1991. Lipid and fatty acid composition of holothurians from tropical and temperate waters. Comp. Biochem. Physiol. 98B, 489-494.
- Swezey, R.R., D. Epel. 1992. The use of caged substrates to assess the activity of 6-phosphogluconate dehydrogenase in living sea urchin eggs. Exp. Cell Res. 201, 366-372.
- Swift, D.J. 1993. The macrobenthic infauna off Sellafield (north-eastern Irish Sea) with special reference to bioturbation. J. Mar. Biol. Ass. U.K. 73, 143-162. (echinoderms, particularly Echinocardium, Amphiura, Leptosynapta)
- Syasina, I.G., M.A. Vashchenko, V.B. Durkina. 1992. Histopathological changes in the gonads of sea urchins exposed to heavy metals. Mar. Biol. (Vlad.). (4), 244-251.
- Tadenuma, H., K. Takahashi, K. Chiba, M. Hoshi, T. Katada. 1992. Properties of 1-methyladenine receptors in starfish oocyte membranes: Involvement of pertussis toxin-sensitive GTP-binding protein in the receptor-mediated transduction. Biochem. Biophys. Res. Comm. 186, 114-121.
- Takagi, S., M. T. Someno, M. Hoshi, H. Sawada. 1992. Participation of 650-kDa protease (20S proteasome) in starfish oocyte maturation. Dev. Biol. 150, 414-418.
- Taniguchi, Y. 1992. Chymotrypsin-like and trypsin-like protease activities in the sea urchin (Hemicentrotus pulcherrimus) egg. Experientia. 48, 287-290.
- Tatarenko, D.E., A.B. Poltarau. 1991. Affiliation of the sea

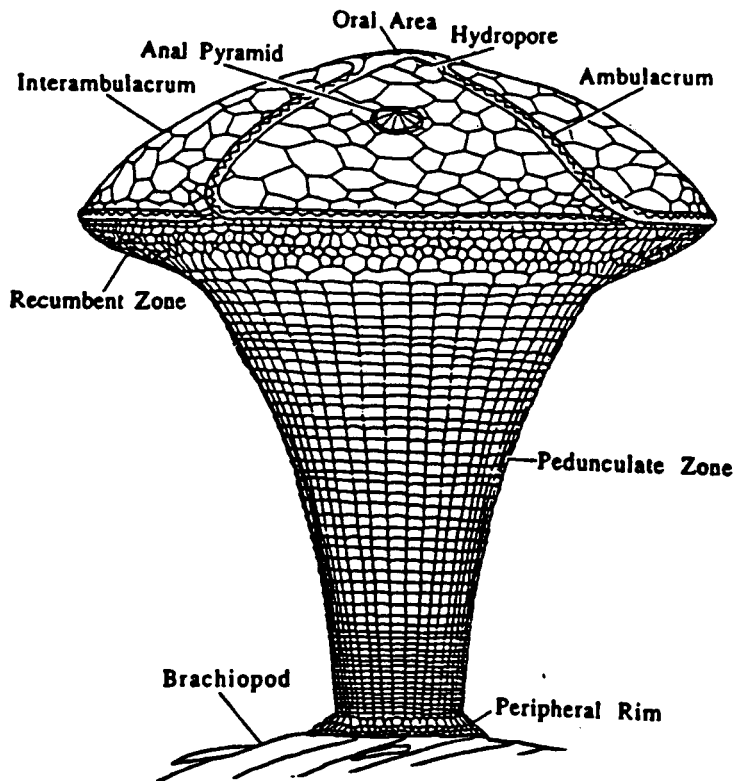


- urchins Strongylocentrotus echinoides and S. sachalinicus to the species S. pallidus based on a comparison of their genomes. *Biol. Morya.* (3), 69-75.
- Temara, A., C. De Ridder, M. Kaisin. 1991. Presence of an essential polyunsaturated fatty-acid in intradigestive bacterial symbionts of a deposit-feeder echinoid (Echinodermata). *Comp. Biochem. Physiol.* 100B, 503-505.
- Temara, A., C. De Ridder, J.G. Kuenen, L.A. Robertson. 1993. Sulfide-oxidizing bacteria in the burrowing echinoid, Echinocardium cordatum (Echinodermata). *Mar. Biol.* 115, 179-185.
- Thandar, A.S. 1991. The cucumariid holothurians of southern Africa with the erection of a new genus. *S. Afr. J. Zool.* 26, 115-139.
- Thorndyke, M.C., B.D. Crawford, R.D. Burke. 1992. Localization of a SALMFamide neuropeptide in the larval nervous system of the sand dollar Dendraster excentricus. *Acta Zool.* 73, 207-212.
- Timiyamoto, K. Togawa, R. Higuchi, T. Komori, T. Sasaki. 1992. Structures of four new triterpenoid oligoglycosides: DS-penaustrosides A, B, C, and D from the sea cucumber Pentacta australis. *J. Nat. Prod. (Lloydia).* 55, 940-946.
- Tosuji, H., I. Mabuchi, N. Fusetani, T. Nakazawa. 1992. Calcyculin A induces contractile ring-like apparatus formation and condensation of chromosomes in unfertilized sea urchin eggs. *Proc. Natl. Acad. Sci. U.S.A.* 89, 10613-10617.

STICKLE ET AL. 1992

TABLE 6. Summary statistics on genetic structure in five species of sea stars in Alaska

	Species				
	Le	Lh	Lp	Et	Po
No. of locations collected	4	1	1	2	3
Total no. of animals collected	344	69	60	193	322
No. of loci examined*	16 (7)	16 (9)	25 (7)	25 (11)	24 (8)
Observed heterozygosity ( $H_o$ )	0.132	0.145	0.084	0.038	0.084
Expected heterozygosity ( $H_e$ )	0.135	0.151	0.083	0.050	0.092
Mean fixation index ( $F_{IS}$ )	0.008	0.045	-0.030	0.247	0.088
Population heterogeneity ( $F_{ST}$ ) <sup>†</sup>	0.156	—	—	0.023	0.006



SUMRALL 1992

Tsuchiya, T., T. Obinata, M. Sato, T. Mori, E. Suzuki, S. Amemiya. 1992. Non-catch contraction in paramyosin-containing muscle in an echinothuriid sea urchin Asthenosoma ijimai. J. Exp. Biol. 162, 361-365.

Tuwo, A., C. Conand. 1992. Reproductive biology of the holothurian Holothuria forskali (Echinodermata). J. Mar. Biol. Ass. U.K. 72, 745-758.

Tyler, P.A., J.D. Gage, G.J.L. Paterson, A.L. Rice. 1993. Dietary constraints on reproductive periodicity in two sympatric deep-sea asteropectinid seastars. Mar. Biol. 115, 267-277.

Tyler, P.A., C.M. Young, D.S.M. Billett, L.A. Giles. 1992. Pairing behaviour, reproduction and diet in the deep-sea holothurian genus Paroriza (Holothurioidea: Synallactidae). J. Mar. Biol. Ass. U.K. 72, 447-462.

Tyler, P.A., H. Zibrowius. 1992. Submersible observations of the invertebrate fauna on the continental slope southwest of Ireland (northeast Atlantic Ocean). Oceanol. Acta. 15, 211-216.

Vadas, R.L., Sr., R.W. Elner. 1992. Plant-animal interactions in the north-west Atlantic. In: D.M. John, S.J. Hawkins, J.H. Price (ed.). Plant-animal interactions in the marine benthos. Clarendon Press, Oxford. pp. 33-60.

VandenSpiegel, D., M. Jangoux. 1993. Fine structure and behaviour of the so-called Cuvierian organs in the holothuroid genus Actinopyga (Echinodermata). Acta Zool. 74, 43-50.

VandenSpiegel, D., A. Ovaere, C. Massin. 1992. On the association between the crab Hapalonotus reticulatus (Crustacea, Brachyura, Eumedonidae) and the sea cucumber Holothuria (Metriatyla) scabra (Echinodermata, Holothuridae). Bull. Inst. Roy. Sci. Nat. Belg. Biol. 62, 167-177.

Van Voorhis, B.J., G.D. Snyder, F.J. Longo. 1992. Effects of

quinacrine on egg activation: a possible role for phospholipase A2 in sea urchin fertilization. *Invertebr. Reprod. Dev.* 21, 33-41.

Varaksina, G.S., A.A. Varaksin. 1991. Localization of steroid dehydrogenase in testes and ovaries of sea urchins. *Biol. Morya* (2), 77-82.

Vasquez, J.A. 1993. Effects on the animal community of dislodgment of holdfasts of *Macrocystis pyrifera*. *Pac. Sci.* 47, 180-184.

Vasquez, M.J., E. Quinoa, R. Riguera, A. Sanmartin, J. Darias. 1992. Santiagoside, the first asterosaponin from an Antarctic starfish (*Neosmilaster georgianus*). *Tetrahedron.* 48, 6739-6746.

Voogt, P.A., J.G.D. Lambert, J.C.M. Granneman, M. Jansen. 1992. Confirmation of the presence of oestradiol-17beta in sea star *Asterias rubens* by GC-MS. *Comp. Biochem. Physiol.* 101B, 13-16.

Walker, C.W., J.D.G. Boom, A.G. Marsh. 1992. First non-vertebrate member of the myc gene family is seasonally expressed in an invertebrates testis. *Oncogene.* 7, 2007-2012. (asteroid)

Wang, X.-D., M.-J. Chen. 1992. Carboniferous-Permian holothurian sclerites from Kalpin area, Xinjiang. *Acta. Micropalaeontol.* 9, 71-79.

Webster. G.D., P.A. Jell. 1992. Permian echinoderms from Western Australia. *Mem. Queensl. Mus.* 32, 311-373.

Webster, G.D. 1991. An evaluation of the Gupta V.J. echinoderm papers, 1971- 1989. *J. Paleontol.* 65, 1006-1008.

Wessel, G.M., S.W. Chen. 1993. Transient, localized accumulation of alpha-spectrin during sea urchin morphogenesis. *Dev. Biol.* 155, 161-171.

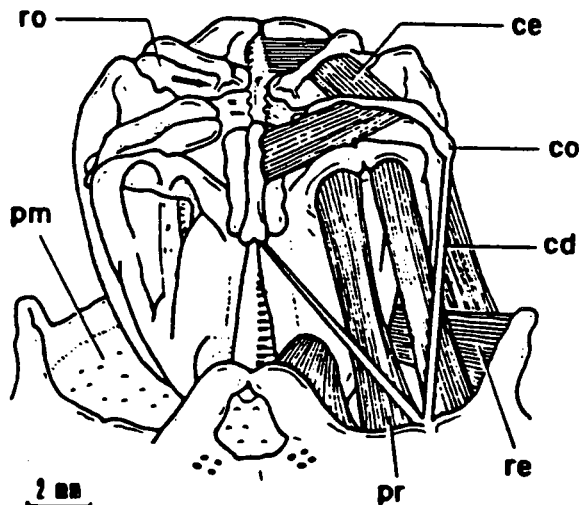


Fig. 1. Diagrammatic representation of the lantern of *Paracentrotus lividus*. The peripharyngeal coelomic membranes are omitted. Only the soft tissue components of the right side have been included: on the left side the insertion facets of the muscles are indicated by stippling. *cd* compass depressor; *ce* compass elevator; *co* compass; *pm* peristomial membrane; *pr* protractor muscle; *re* retractor muscle; *ro* rotula. Arrow indicates one of the pair of ligaments that link the compass to the rotula

Wilkie, Candia Carnevali  
and Bonasoro. 1992

Westervelt, C.A., Jr., E.N. Kozloff. 1992. Two new species of *Syndesmis* (Turbellaria: Neorhabdocoela: Umagillidae) from the sea urchins *Strongylocentrotus droebachiensis* and *Allocentrotus fragilis*. *Cah. Biol. Mar.* 33, 115-124.

Whaley, T., A. McDougall, I. Crossley, K. Swann, M. Whitaker. 1992. Internal calcium release and activation of sea urchin eggs by cGMP are independent of the phosphoinositide signaling pathway. *Mol. Biol. Cell.* 3, 373-383.

Wilkie, I.E. 1989. The fauna of the Clyde Sea Area. Echinodermata. *Univ. Mar. Biol. Sta., Millport. Occ. Pub. No. 6.*

Wilkie, I.E., C. Carnevali, F. Bonasoro. 1992. The compass depressors of *Paracentrotus lividus* (Echinodermata: Echnoida): ultrastructural and mechanical aspects of their variable tensility and contractility. *Zoomorphology.* 112, 143-153.

Wilkie, I.E. 1992. Variable tensility of the oral arm plate ligaments of the brittlestar *Ophiura ophiura* (Echinodermata: Ophiuroidea). *J. Zool.* 228, 5-26.

Williams, E.H., Jr. 1991. Threat to black sea urchins. *Nature.* 352, 385.

Witman, J., K.P. Sebens. 1992. Regional variation in fish predation intensity: a historical perspective in the Gulf of Maine. *Oecologia.* 90, 305-315.

Wray, G.A. 1992. The evolution of larval morphology during the post-Paleozoic radiation of echinoids. *Paleobiology.* 18, 258-287.

Wray, G.A. 1992. Rates of evolution in developmental

GREGORY A. WRAY 1992

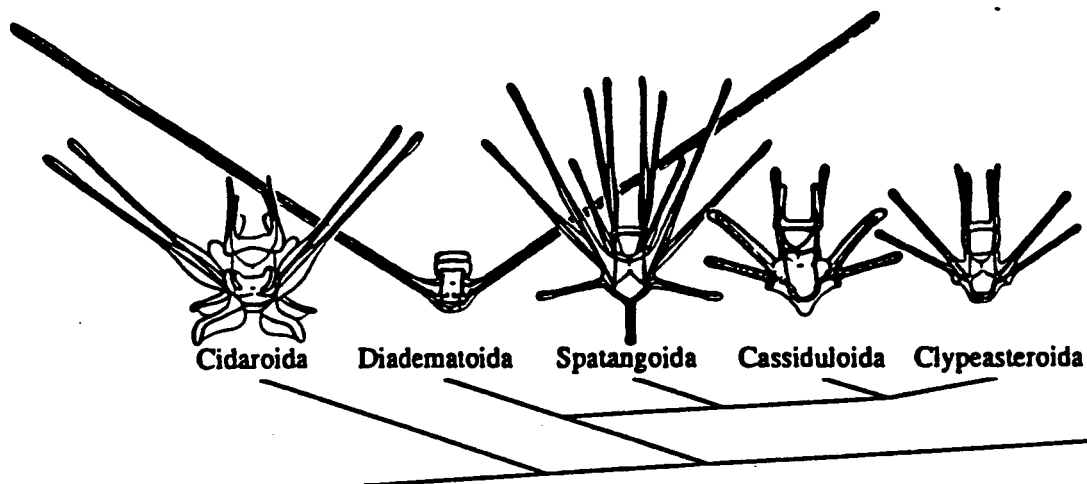


FIGURE 1. Diversity in echinopluteus larvae with planktotrophic development. The overall form of echinoplutei varies considerably between orders and within some orders. Illustrated are representative echinoplutei from several orders and families. Most of these clades themselves contain a significant diversity in echinoplutei. Species shown.

- processes. *Am. Zool.* 32, 123-134.
- Xu, R.A. 1991. Annual changes in the steroid levels in the testis and the pyloric caeca of Sclerastrias mollis (Hutton) (Echinodermata, Asteroidea) during the reproductive cycle. *Invert. Reprod. Dev.* 20, 147-152.
- Yasuhara, T., H. Yokosawa, M. Hoshi, S. Ishii. 1991. Involvement of a sperm aminopeptidase in fertilization of the sea urchin. *Experientia.* 47, 100-103.
- Yokochi, H., S. Veno, M. Ogura, A. Nagai, T. Habe. 1991. Spatial and temporal changes of the distribution of Acanthaster planci (L.) and corals at northwestern Iriomote-jima, Ryukyu Islands, 1983-1988. *J. Fac. Mar. Sci. Tech. Tokai Univ.* (32), 321-242.
- Yoshida, K.-I., Y. Minami, H. Nemoto, K. Numata, E. Yamanaka. 1992. Structure of DHG, a depolymerized glycosaminoglycan from sea cucumber, Stichopus japonicus. *Tetrahedron Lett.* 33, 4959-4962.
- Yoshino, K.-I., N. Suzuki. 1992. Two classes of receptor specific for sperm-activating peptide III in sand dollar spermatozoa. *Eur. J. Biochem.* 206, 887-893.
- Yoshino, K.-I., T. Tako, Y. Shimonishi, N. Suzuki. 1992. Sperm-activating peptide type-V (SAP-V), a fifth member of the sperm-activating peptide family, purified from the egg-conditioned media of the heart urchin Brissus agassizii agassizii. *Comp. Biochem. Physiol.* 102B, 691-700.
- Young, C.M., P.A. Tyler. 1993. Embryos of the deep-sea echinoid Echinus affinis require high pressure for development. *Limnol. Oceanogr.* 38, 178-181.
- Zachos, L.G. 1993. Occurrence of the spatangoid echinoid Maretia arguta (Clark) in the Middle Eocene of Texas. *J. Paleontol.* 67, 148-150.

## THESES

## BELGIUM

Bs. C. Theses

Dominique, F. 1973. Contribution a l'etude du cycle annuel de reproduction de deux especes d'echinoides (echinodermata) des cotes de Bretagne. Univ. Libre de Bruxelles

Mem. Lic. Zool.

Powis de Tendbossche, T. 1978. Comportement alimentaire et structure digestive de Paracentrotus lividus (Lmk.) (Echinodermata: Echinoidea). Univ. Libre de Bruxelles.

## CANADA

Ph.D. Theses

Broom, J.D.G. 1988. Actin gene transcription during spermatogenesis in the sea star Pisaster ochraceus Brandt. Simon Fraser University.

Campbell, S.S. 1990. Morphology and histochemistry of the extracellular matrix of embryos following freeze substitution of the starfish Pisaster ochraceus. Univ. of British Columbia.

Cserjesi, P. 1990. The structure and expression of the metaallothionein genes of the sea urchin Lytechinus pictus. McGill Univ.

Gong, Z. 1987. Regulation of tubulin gene expression in sea urchin embryos. McGill Univ.

Hagen, N.T. 1990. Outbreak dynamics of the green sea urchin Strongylocentrotus droebachiensis O.F. Muller. Dalhousie Univ.

Starr, M. 1990. Mecanismes de coordination entre la ponte de certains invertebres marins et la pousse printaniere du phytoplankton. Universite Laval.

Master's Theses

Cody, L.W. 1973. Aspects of the reproductive biology of Cucumaria frondosa (Gunnerus, 1770) and Psolus fabricii (Duben and Koren, 1846) (Echinodermata: Holothuroidea) in shallow waters of the Avalon Peninsula, Newfoundland. Memorial Univ. Newfoundland.

## CHILE

Licenciado

Arevalo O., J. 1968. Contribucion al conocimiento de la macrofauna de la zona intermareal de la Bahia de Concepcion. Univ. Concepcion. (with keys to echinoderms)

Bay-Schmith B., E. 1975. Aspectos ecologicos de la poblacion de Stichaster stiratus Muller et Troschel, 1840, en la Bahia de Concepcion, Chile (Asteroidea: Echinodermata). Univ. Concepcion.

- Caffi G., M. 1981. Aspectos del ciclo reproductivo de Athyonidium chilensis (Semper, 1868) en Caleta Cocholque, Bahía de Concepción, Chile (Echinodermata, Holothuroidea). Univ. Concepción.
- Castilla, J. 1965. Systemática de los ofiuroideos de la costa Chilena. Univ. Concepción.
- Munoz I., S. Vega J. 1987. Aspectos tróficos de Stichaster striatus (Müller y Troschel, 1840) durante en fenómeno "El Niño" 1982-1983 en la zona de Iquique-Chile. Univ. Arturo Prat.
- Salas, J.C. 1980. Estudio de la biotoxicidad de Athyonidium chilensis (Semper, 1868) (Echinodermata: Holothuroidea: Dendrochirotida). Univ. Concepción
- Troncoso, R.H. 1983. Introducción de Cucumaria godeffroyi Semper, 1868 (Echinodermata: Holothuroidea). Univ. Concepción.
- Urbina, M. 1981. Sinopsis y clave para las especies de holoturoideos (Holothuroidea) de Chile. Univ. Concepción.
- Verdejo, C. 1951. Estudio químico-biomatológico de la estrella de mar (Stichaster aurantiacus F.Y.F. Meyen. Univ. Concepción.
- Werlinger, C.L. 1981. Algunos aspectos sobre de biología de la reproducción de Patiria chilensis (Lutken, 1859) (Echinodermata, Asteroidea). Univ. Concepción.
- Yanez, A. 1971. Estudio prospectivo cuali y cuantitativo de la macrofauna bentónica del sublitoral de Bahía de Concepción, Chile. Univ. Concepción. (asteroids, ophiuroids, echinoids, holothuroids)

## COLUMBIA

### Tesis de Grado

- Pardo Angel, Roberto. 1989. Estudio taxonómico y ecológico de los principales grupos de Equinodermos del Litoral Pacífico Colombiano. Universidad del Valle.

## FRANCE

### Mem. D.E.A.

- Rico, V. 1989. Contribution a l'étude des preferences alimentaires et du comportement moteur de l'oursin régulier Paracentrotus lividus Univ. Aix-Marseille II.
- Frantzis, A. 1988. Relations trophiques entre les oursins Arbacia lixula et Paracentrotus lividus (Echinoidea, Regularia) et le phytobenthos infralittoral superficiel dans la baie de Port-Cros (Var, France). Univ. Aix-Marseille II. (Luminy)

### These 3e Cycle

- Gentil, F. 1976. Distribution des peuplements benthiques en baie de Seine. Univ. Paris VI.

### Docteur de l'Université

- Frantzis, A. 1992. Etude expérimentale des niveaux de consommation et d'utilisation des macrophytes et des détritiques

derives par deux invertébrés benthiques. Univ. Aix-Marseille II. (Luminy). (echinoid)

### Docteur en Sciences

Azzolina, J.F. 1988. Contribution à l'étude de la dynamique des populations de l'oursin comestible Paracentrotus lividus (LMCK). Croissance, recrutement, mortalité, migrations. Univ. Aix-Marseille II. (Luminy)

Davoult, D. 1988. Etude du peuplement des cailloutis à épibiose sessile et de la population d'Ophiothrix fragilis (Abildgaard) du détroit du Pas-de-Calais (France). Univ. de Lille-Flandres-Artois.

Delmas, P. 1992. Etude des populations de Paracentrotus lividus (Lam.) (Echinodermata: Echinoidea) soumises à une pollution complexe en Provence nord-occidentales: densités, structure, processus de detoxication (Zn, Cu, Pb, Cd, Fe). Univ. d'Aix-Marseille III. (St. Jerome)

### GERMANY

Birenheide, R. 1989. Ultrastrukturelle und experimentelle Untersuchungen am Kauapparat und am Darmkanal des Seeigels. Ruhr-Universität Bochum.

### GHANA

#### M.Sc. Theses

Sanusi, S.S. 1980. A study on grazing as a factor influencing the distribution of benthic littoral algae. Univ. of Ghana. (echinoid)

### IRELAND

#### B.Sc. Theses

Grehan, A. 1982. Aspects of the biology and ecology of Amphiura filiformis (O.F. Muller) (Echinodermata: Ophiuroidea). University College, Galway.

### JAMAICA

#### M.Phil. Theses

Gordon, C.M. 1990. Taxonomy and palaeoecology of the echinoids of the late Pleistocene Falmouth Formation of Jamaica. Univ. of the West Indies, Mona

### NORWAY

Fosshagen, A. 1965. Bunnevertebratlarver over et Venus-samfunn i Oresund. Univ. of Bergen.



**PAKISTAN**Ph.D. Theses

Tahera, Qaseem. 1992. Taxonomic studies of northern Arabian Sea echinoderms. Univ. of Karachi.

**PUERTO RICO**MS Thesis

Bissell, J.L. 1978. An investigation of the feeding habits of and population biology of the sea-star Astropecten duplicatus (Gray). Univ. of Puerto Rico.

C6lon-Jones, D.E. 1993. Size (age) specific factors controlling the distribution and population size of the white-spined sea urchin, Tripneustes ventricosus (Lamarck, 1816). Univ. of Puerto Rico.

**REPUBLIC OF CHINA**Master of Science

Chen, B.-Y. 1989. Life history of Patiriella pseudoexigua Dartnall (Echinodermata: Asteroidea). Nat. Sun Yat-sen Univ.

Doctor of Philosophy

Chao, S.-M. 1993. Reproductive biology of sea cucumbers in southern Taiwan (Echinodermata: Holothuroidea). Tunghai Univ.

**RUSSIA**Doctor of Biological Sciences

Ryabushko, Valeri I. Energetic exchange of echinoderms (Type Echinodermata).

**SWEDEN**

Wintzell, J. 1918. Bidrag till de skandinaviska Ophiuridernas biologi och fysiologi. Univ. of Uppsala.

**UNITED KINGDOM**Master's Theses

Taylor, A.M. 1958. Studies on the biology of the offshore species of Manx Ophiuroidea. Univ. of Liverpool.

Ph.D. Theses

Gilliland, P.M. 1990. The skeletal morphology and systematics of Recent and fossil holothurians with particular reference to th

Triassic/Jurassic. Univ. of Exeter.

Hawkins, S.J. 1979. Field studies on Manx rocky shore communities. Univ. of Liverpool.

Lawson, G.W. Studies in the intertidal ecology of rocky shores in West Africa. Univ. of London.

Pitchford, A.J. 1989. The stratigraphy, palaeontology, and palaeoecology of the Campanian Chalk of Norfolk. Univ. Liverpool.

Vost, L.M. 1985. The influence of grazing by the sea urchin Echinus esculentus L. on subtidal algal communities. Univ. of Liverpool.

## UNITED STATES

### Master's Theses

Beddingfield, S.D. 1992. The feeding biology of Astropecten articulatus (Echinodermata: Asteroidea) from the Gulf of Mexico: an evaluation of optimal foraging theory in a soft-bottom predator. Univ. of Alabama/Birmingham.

D'Andrea, A.F. 1993. Sublethal effects of cadmium on arm regeneration in the burrowing brittlestar, Microphiolis gracillima (Stimpson). Univ. of South Carolina.

Fell, F.J. 1971. The Echinoidea of Anton Brunn cruises 12 and 13, 1965-1966, to the southeastern Pacific. Univ. of Maine.

Golde, H. 1992. Respiration rates of regenerating Microphiopholis gracillima (Echinodermata: Ophiuroidea). Univ. of South Carolina.

Hintz, J.L. 1993. The effect of salinity on early development of several species of echinoids and asteroids (Echinodermata). Univ. of South Florida.

Lewis, L.M. 1992. Habitat use, diet, and parasitism of the seastar Rathbunaster californicus Fisher from the Monterey submarine canyon, California. San Jose State Univ.

Mauzey, K.P. 1966. Feeding behavior and reproductive cycles in Pisaster ochraceus. Univ. of Washington.

Smith, S.L. 1991. Investigation of histamine in echinoderms. Univ. of South Florida.

### Ph.D. Theses

Axon, A.G. 1992. Paleocology of a Cincinnatian (Upper Ordovician) crinoid-garden community from southwestern Ohio. Univ. of Kentucky.

Chen, J. 1991. The role of DNA methylation in ectopic MyoD1 expression and sea urchin morphogenesis and gene expression. Univ. of Southern California.

De Vogelaere, A.P. 1991. Disturbance, succession and distribution patterns in rocky intertidal communities of central California. Univ. of California, Santa Cruz.

Eakin, C.M. 1991. The damselfish-algal lawn symbiosis and its influence on the bioerosion of an El Nino impacted coral reef, Uva Island, Pacific Panama. Univ. of Miami.

Hahn, J.-H. 1991. Evolution of structure, expression and regulation of actin gene of the sea urchin Heliocidaris. Indiana University.

Jamison, D.C. 1992. Hatching secretions of the sea urchin, Strongylocentrotus purpuratus. Univ. of Denver.

Kingsley, P.D. 1992. Spatial and temporal patterns of gene expression during sea urchin early development. Univ. of Rochester.

Kozlowski, M.T. 1992. A study of sea urchin USF: a helix loop-helix protein active in embryonic ectoderm cells. Univ. of Houston.

Leonard, A.B.P. 1992. The biomechanics, autecology and behavior of suspension feeding in crinoid echinoderms. Univ. California, San Diego.

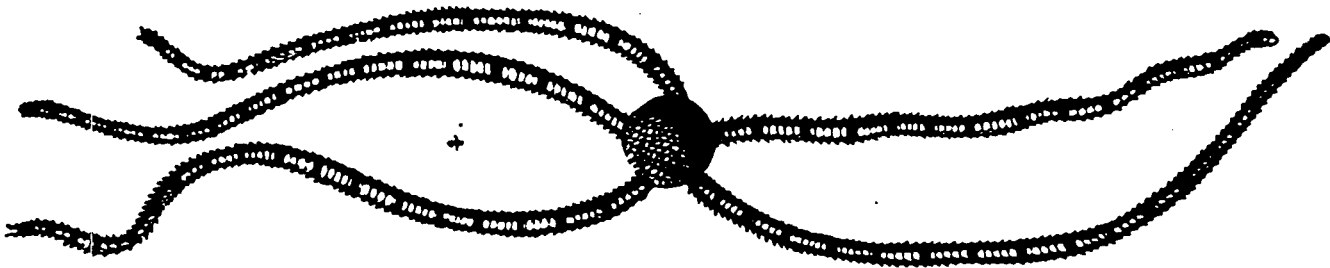
McGlathery, K.J. 1992. Nutrient and herbivore influences on seagrass community dynamics. Cornell Univ. (echinoids)

Reynolds, S.D. 1992. The Strongylocentrotus purpuratus early blastula gene set. Univ. of Rochester.

Wang, C.-C. 1992. Phylogenetic analysis of the clypeasteroid echinoids. Yale Univ.

Wendelburg, B.J. 1991. Determination of the transcription control elements of the sea urchin early U1 snRNA gene promoter. Florida State Univ.

Zhao, Z. 1992. Temporal regulation of the L1 late H2B histone gene during sea urchin embryonic development by an Antemapedial class homeoprotein. Univ. of Southern California.



Kuehler 1905.

## PAPERS PRESENTED AT MEETINGS

## 7th International Coral Reef Symposium

June, 1992, Guam

pages of abstracts volume

- Arizpe, O., H. Reyes, A. Villareal, F. Sinsel. Cabo Pulmo: the northernmost coral reef of the eastern Pacific (Gulf of California, Mexico). (echinoid, asteroid, Acanthaster). p. 4
- Babcock, R.C., C.N. Mundy. Seasonal changes in fertility and fecundity in *Acanthaster planci*. p.6.
- Bak, R.P.M. Echinoids and their impact on coral reef carbonate budgets in past, present and future. p.7.
- Conand, C. Evolution of tropical holothurian fisheries. p. 19.
- De'ath, G., P. Moran. A summary of ecological data on the crown-of-thorns starfish. p. 23.
- Edinger, E.N., M.J. Risk. Bioerosion on modern and fossil reefs: controls, consequences, and corroboration. (echinoids) p. 27.
- Fabricius, K.E. Multispecies-associations of symbionts on the central Great Barrier Reef. p. 28.
- Fagerstrom, J.A. Impact and recovery of reefs from an *Acanthaster planci* outbreak, Moorea, French Polynesia. p. 29.
- Fernandez, L., P.J. Moran, H. Marsh. A system for classifying outbreaks of crown-of-thorns starfish as a basis for management. p. 30.
- Halford, A.R., J.K. Keesing. Large-scale rearing of juvenile crown-of-thorns starfish for ecological experiments. p. 41.
- Keesing, J.K. Role of *Acanthaster planci* in structuring reef communities in Okinawa, Japan. p. 52.
- Lara-P.S., M., A.C. Padilla-Souza, J.J. Espeiel-Montes, C. Garcia-Saez. Coral reefs of Veracruz, Mexico. I. Zonation and community structure. (*Echinometra*). p. 57.
- Lassig, B., W. Gladstone, P. Moran, U. Engelhardt. A crown-of-thorns contingency plan. p. 58.
- Messing, C.G. Diversity and ecology of comatulid crinoids (*Echinodermata*) at Madang, Papua New Guinea. p. 68.
- Mezaki, S. Changing environments of *Acanthaster planci* in the Ryukyu Islands, Japan. p. 68.
- Moran, P., R. Babcock, J. Keesing, B. Lassig. Crown-of-thorns starfish: synthesis of recent research. p. 71.
- Palumbi, S.R. Strong differences between populations of Indo-Pacific urchins revealed by MTDNA sequences. (*Echinometra*) p. 80.
- Scandol, J.P., M.K. James. Comparing a population model of *Acanthaster planci* incorporating hydrodynamics and larval dispersal with results of broad scale surveys on the Great Barrier Reef. p. 93.
- Stump, R.J.W. Age and growth of *Acanthaster planci* (L.) from the GBR, Australia. p. 99.
- Wiedemeyer, W.L. Daily feeding behaviour in two tropical holothurians, *Holothuria scabra* (Jaeger) and *H. atra* (J.). p. 108.

## ZOOLOGICAL SOCIETY OF JAPAN

1992. Sendai

Abstracts published in *Zoological Science* 9 (6). 1992.

- Yamazaki, K., et al. Molecular cloning of Na<sup>+</sup>,K<sup>+</sup>-ATPase alpha-subunit gene in the sea urchin *Hemicentrotus pulcherrimus*. p. 1143.
- Mizoguchi, H. Localization of M phase cells in sea urchin blastulae. p. 1158.
- Kiyomoto, M., H. Shirai. Archenteron-forming ability of vegetal region of starfish egg. p. 1158.
- Kuraishi, R., K. Osanai. Expression of alkaline phosphatase in starfish larvae. p. 1158.
- Shimizu, K., Y. Nakajima, M. Ikeda. Morphogenesis in spicule-removed embryos of the sea urchin. p. 1158.
- Ohta, K., M. Sato, T. Nakazawa. Change in molecular form and activity of acetylcholinesterase during spicule formation in sea urchin embryo. p. 1159.
- Kiyomoto, M., M. Maruoka, J. Tsukahara. Spicule formation-inducing substance in blastocoelic fluid of sea urchin blastula. p. 1159.
- Kinoshita, T., K. Kinoshita. Roles of TGF-beta superfamily genes in differentiation of micromeres of sea urchin embryos. p. 1159.
- Iwase, T., T. Miki-Noumura. Spicule formation of primary mesenchyme cells in the isolated bags from sea urchin embryos. p. 1159.
- Amemiya, S. Metamorphic potency of the aggregates derived from mesomeres isolated from 16-cell-stage embryos of a sand dollar, *Peronella japonica*. p. 1160.
- Chino, Y. et al. Formation of adult rudiment and metamorphosis of sea urchin by thyroid hormones. p. 1160.
- Mita, M. 1-methyladenine production by ovarian follicle cells responsible for oocyte maturation in starfish. p. 1161.
- Kishimoto, K., S. Hisanaga, T. Kishimoto. Starfish oocyte P34cdc2 kinase associated with detergent-resistant cytoskeletons. p. 1161.
- Yoneda, M. Fusion of starfish oocytes with different maturation phases. p. 1163.
- Akasaka, K. et al. Regulatory region of sea urchin aryl-sulfatase gene responsible for spatial expression. p. 1163.
- Yamada, K. et al. Quantitative analysis of the CIS-acting elements regulating the expression of arylsulfatase (ARS) gene in sea urchin embryo. p. 1164.
- Iuchi, Y. et al. Newly detected CIS-acting elements required for increased expression of the arylsulfatase (Ars) gene in the sea urchin (*H. pulcherrimus*) embryo. p. 1164.
- Sakamoto, N. et al. Single and triple strand structures (H-DNA) in regulatory region of arylsulfatase gene of sea urchin embryo. p. 1164.
- Fujita, et al. The EGIP-binding protein in embryos of the sea urchin *Anthocidaris crassispina*. p. 1164.
- Haruguchi, Y. et al. Structural analysis of exogastrula-inducing peptide gene. p. 1165 (echinoid)
- Yamasu, K. et al. Analysis of cDNA fragments for protein tyrosine kinases obtained by RT-PCR method. p. 1165. (echinoid)
- Takeda, S., H. Hayashi. In situ protein phosphorylation in a sea

urchin embryo loaded with radioactive phosphate: analysis by 2D-page. p. 1165.

Okuyama, M. et al. Proteins to be phosphorylated in the reactions catalyzed by CAM kinase, C kinase, A kinase and G kinase in sea urchin embryos. p. 1165.

Kettoku, M. et al. Regulation of early embryonic histone gene expression by  $Ca^{++}$  signals in embryos of the sea urchin, *Hemicentrotus pulcherrimus*. p. 1166.

Furuya, S. et al. (ADP-ribosyl)ation of histones in nuclei isolated from sea urchin embryos. p. 1166.

Kawamoto, M. et al. Partial purification and characterization of a casein-kinase 2-type protein kinase from sea urchin. p. 1166.

Shimizu, T. et al. New histone molecules produced at the blastula stage of starfish embryos. p. 1167.

Nakasone, M. et al. Probable participation of DNA methylation in ectoderm cell differentiation in sea urchin embryos. p. 1167.

Fujiwara, A., T. Nakagawa, I. Yasumasu. Abnormal embryos derived from eggs treated with SCN-. p. 1167.

Nakagawa, T., A. Fujiwara, I. Yasumasu. Extracts of non-dialyzable compounds extracted by SCN- from sea urchin eggs on morphogenesis in embryos. p. 1167.

Kuno, S., T. Nagura, I. Yasumasu. Insulin receptor of the cultured cells derived from micromeres of sea urchin embryos. p. 1168.

Takemoto, K. et al. Exocytosis during fertilization of a sea urchin egg detected by fluorescence dequenching method. p. 1175.

Uto, N. et al. Effects of x ray and/or magnetic fields on sea urchin development. p. 1176.

Ueda, E. et al. The participation of phosphorylated proteins in microtubule nucleating activity of MTOG isolated from sea urchin egg mitotic apparatus. p. 1176.

Hamaguchi, Y., M.S. Hamaguchi, S. Sato. Deformation of the nuclear envelope caused by microtubules during cell division in sea urchin eggs. p. 1177.

Saiki, T., Y. Hamaguchi. Relationship between aster formation and cell division through maturation to cleavage revealed by transplantation of the starfish centrosome. p. 1177.

Washitani-Nemoto, S., S.-I. Nemoto. Behavior of nuclei and chromosomes for parthenogenetic starfish eggs to establish tetraploidy. p. 1177.

Nomura, A., S. Nemoto. DNA replication for parthenogenetic starfish eggs to establish tetraploidy. p. 1178.

Watanabe, K., M.S. Hamaguchi, Y. Hamaguchi. Metaphase is prolonged in sea urchin eggs treated with low pH sea water containing sodium acetate. p. 1178.

Kominami, T. Pseudopodia forming activity during early cleavage stages of sea urchin embryos. p. 1178.

Yokota, Y. et al. Immunological studies on extracellular matrix proteins of sea urchins. p. 1178.

Myotoishi, Y. et al. Fate of an extracellular matrix component of fertilized sea urchin (*Hemicentrotus pulcherrimus*) eggs. p. 1179.

Nakajima, Y., K. Shimizu. Collagenous fibers in the sea urchin *plutei* blastocoel. p. 1179.

Nemoto, S. et al. Activation of starfish eggs by caffeine:  $Ca^{2+}$ -

- release, cortical granule exocytosis and oxygen consumption. p. 1181.
- Kojima, M.K. et al. Effects of H-7 and W-7 on changes in the rate of oxygen consumption and the content of arginine phosphate following treatments with activating reagents in sea urchin eggs. p. 1181.
- Ushiyama, A., K. Chiba, M. Hoshi. Localization for specific receptor for ARIS on spermatozoa of starfish, *Asterias amurensis*. p. 1197.
- Hoshino, K., T. Harumi, N. Suzuki. Characterization of two different protein phosphatases in sperm tails of the sea urchin *Hemicentrotus pulcherrimus*. p. 1205.
- Shimizu, T., Y. Sendai, N. Suzuki. Cloning of cDNAs encoding guanylate cyclase and sperm activating peptide-I receptor expressed in *Hemicentrotus pulcherrimus* sperm. p. 1205.
- Sato, Y. et al. Characterization of wheat germ agglutinin-binding protein in sperm tails of the sea urchin *Hemicentrotus pulcherrimus*. p. 1205.
- Kinoh, H., H. Fujimoto, N. Suzuki. Cloning of cDNAs encoding the putative precursors for sperm-activating peptide (SAPs). p. 1205.
- Osawa, M. et al. Heparin inhibits calcium transients in fertilized sea urchin eggs induced by sperm and its soluble extracts. p. 1206.
- Morimatsu, A., H. Murofushi, I. Mabuchi. TPA-sensitive protein phosphorylation interferes with cortical reaction and induces cytoskeletal changes in the sea urchin egg. p. 1207.
- Saotome, K. et al. Analysis of male chromosomes from the hybrid andromerogones obtained by using cytopreserved sperm of sea urchins. p. 1210.
- Baba, S.A., M. Imagawa, Y. Mogami. Discrete nature of flagellar bending. p. 1219. (starfish)
- Ishijima, S., Y. Hamaguchi. Mechanism of calcium-induced reversal of direction of rotational movement in reactivated sea urchin spermatozoa. p. 1219.
- Shingyoji, C. et al. Effect of ATP concentration on the velocity of microtubule sliding in reactivated sea urchin sperm flagella under imposed head vibration. p. 1219.
- Birenheide, R., T. Motokawa, S. Amemiya. Sponge spicules in the body of spongivorous sea urchins. p. 1251.
- Tominaga, H., M. Komatsu. Aggregation for spawning in the breeding season of the sea star, *Asterina minor*. p. 1292.
- Minokawa, T., S. Amemiya, N. Matsuoka. Genetic variation and differentiation in two local Japanese populations of the sea urchin, *Asthenosoma iijimai*: electrophoretic analysis of allozymes. p. 1299.
- Fujisawa, H. Thermosensitivity of sea urchin embryos is determined maternally. p. 1299.
- Komatsu, M., et al. Development of an apodous holothuroid of the genus, *Labidoplax*. p. 1299.
- Komatsu, M. et al. Development of the sea star, *Luidia maculata* Muller et Troshel. p. 1299.
- Matsuoka, N. et al. Biochemical systematics of the five starfish species of the family Asteriidae from Japanese waters. p. 1300.

**AMERICAN SOCIETY OF ZOOLOGISTS**

1992. Vancouver, British Columbia  
Abstracts published in *American Zoologist*. 32 (5). 1992.

- Hines, S.A., S.A. Watts, J.B. McClintock, K.R. Marion, T.S. Hopkins. A comparison of steroid metabolism in echinoderms of the northern Gulf of Mexico. 23A.
- Shick, J.M., W.C. Dunlap, R.M. Larsen. Accumulation of UV photoprotectants in coral reef holothuroids (Echinodermata). 43A.
- Thurmond, F., J. Trotter. Microfibrils from sea cucumber body wall. 44A.
- Miller, R.L., R.G. Vogt, T. P8unnett. Identification of the sperm attractant from a starfish. 79A
- Oppenheimer, S.B., and 23 co-authors. Mapping developmental changes in sea urchin sugar receptors using lectin beads. 85A.
- Shilling, F.M. Morphological plasticity of echinoid larvae in response to DOM and consequences for nutrient uptake. 86A.
- Bryan, P.J., J.B. McClintock, K. Marion, S.A. Watts, T.S. Hopkins. Feeding deterrence and chemical defense in echinoderm body wall tissues from the northern Gulf of Mexico. 100A.
- Appelmans, N. Evidence for chemically mediated particle selection obtained from video observations of sea urchin larvae. 109A.
- Balser, E.J., E.E. Ruppert, W.B. Jaeckle. Ultrastructure of the axohydrocoel in holothuroid echinoderm larvae. 109A.
- Basch, L.V. Variation in echinoderm larval nutrition: consequences for pre- and early post-settlement life history strategies. 110A
- Frick, J.E., E.E. Ruppert, J.P. Wourms. Nutrition of brooded young in a sea cucumber (*Synapta hydriformis*). 113A.
- George, S.B. Reproductive responses of the seastar *Leptasterias* spp to variations in food supply. 113A.
- Hart, M.W. Larval feeding performance and egg size evolution in echinoids. 114A.
- Hendler, G. 150 million years of swimming brittlestars in brief 114A.
- Herrera, J.C., L.R. McEdward. Body form and skeletal growth in larvae of the sea urchin *Lytechinus variegatus*. 115A.
- Johnsen, S. Can echinoids detect and use polarized light? 115A.
- Levitan, D.R., C.M. Young. The Allee effect in a large free-spawning population. 117A. (Clypeaster)
- McEdward, L.R., D.A. Janies. Evolution of direct development in the velatid starfish *Pteraster tesselatus*. 117A.
- Podolsky, R.D. Separative the effects of temperature and viscosity on activity of echinoderm larvae. 119A.
- Slattery, M., I. Bosch. Costs of brooding and development in an Antarctic sea star. 120A.
- Janies, D.A., L.R. McEdward. Nighly derived coelomogenesis in the velatid starfish *Pteraster tesselatus*: implications for the evolution of the concentricyloid water-vascular system. 125A.
- Shinner, G.O., J.B. McClintock. Structural characteristics of marsupial brood pouches of the Antarctic spatangoids *Abatus nimrodi* and *Abatus shackletoni*. 128A.
- Smith, S.M., J.H. Dearborn. Ophiuroid tube feet: comparative



morphology of species from the NW Atlantic. 129A.

Szulgit, G.K., R.D. Shadwick. The effects of calcium chelation and cell lysis on the mechanical properties of sea urchin ligaments. 129A.

LaCalli 1993

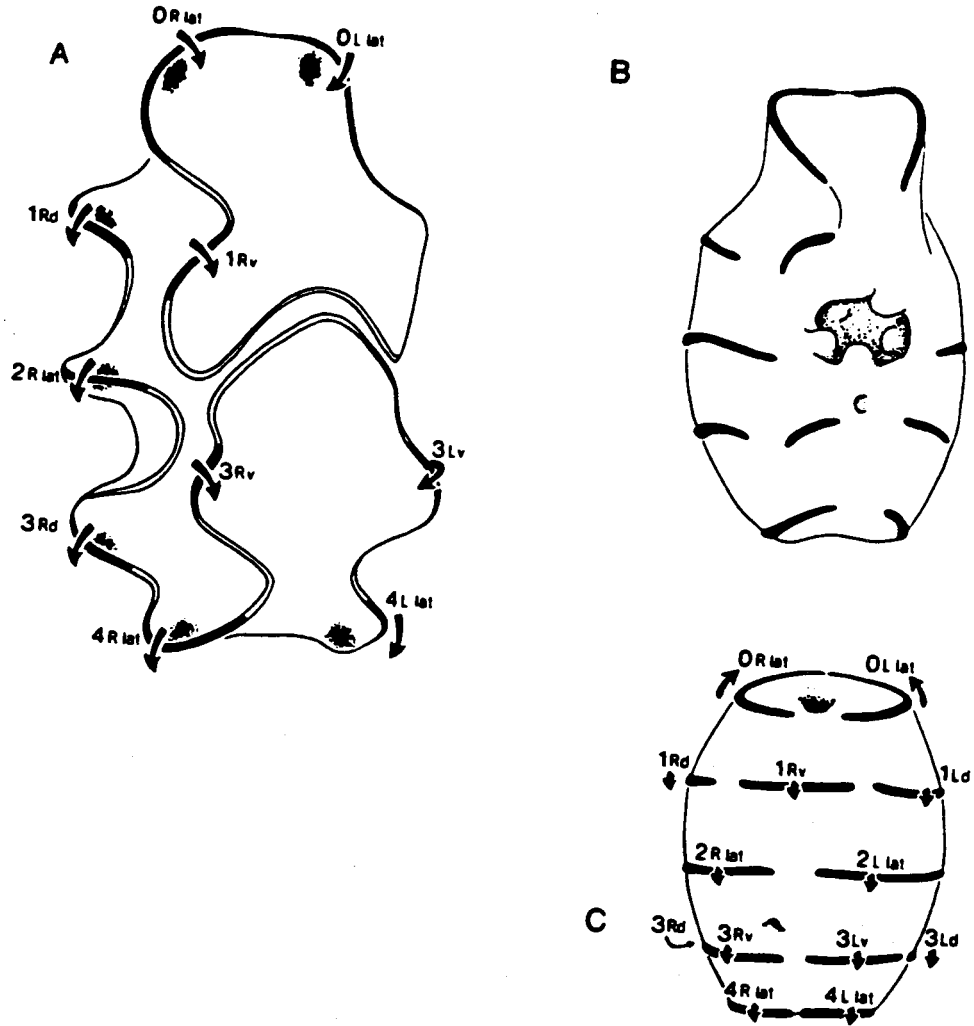


Fig. 2. Summary diagrams of the auricularia-to-doliolaria transition in *S. californicus* showing the origin and positional shifts of the band segments retained by the doliolaria

JOHN  
LAWRENCE

# FRIENDS OF ECHINODERMYS



*Beaman '92*

## 1992

HARBOR BRANCH  
OCEANOGRAPHIC INSTITUTION, INC.

10, 11 July 1992

- Lawrence, J., D. Mahone, and W. Avery. Metals in the body components of *Luidia clathrata* in Tampa Bay and the Gulf of Mexico.
- McHenry, A., and S.E. Stancyk. Cadmium uptake by the burrowing brittlestar *Microphiopholis gracillima*.
- D'Andrea, A.F., and S.E. Stancyk. Effect of Cadmium on arm regeneration of a burrowing brittlestar.
- Burton, M. Indoles in urchins.
- Ferguson, J.C. Fluid volume homeostasis in starfish.
- Telford, M., and O. Ellers. Are echinoids inflated structures?
- Ellers, O. and M. Telford. Laplace's law and peristomially produced pressures in echinoids.
- Adams, J.M., and J. Lawrence. The effect of arm loss on respiration and excretion in *Luidia clathrata* (Asteroidea).
- Lares, M.T., and J. Lawrence. The effect of food quantity on small *Lytechinus variegatus* (Echinodermata: Echinoidea).
- Bishop, C.D., and S.A. Watts. Growth of the digestive system in an echinoid.
- Hines, G.A., and S.A. Watts. Steroid metabolism in echinoids from the Gulf of Mexico.
- Miller, R.L. Identification of a sperm attractant from the starfish *Pycnopodia helianthoides*.
- Hintz, J., and J. Lawrence. Pre-spawning salinity acclimation in gametes of the asteroid *Luidia clathrata*.
- Watts, S.A., J.B. McClintock, T.S. Hopkins, G.O. Schinner and K.R. Marion. Reproductive Biology of *Astropecten articulatus* in the northern Gulf of Mexico.
- McClintock, J.B., S.A. Watts, K. Marion and T.S. Hopkins. Reproductive biology of two deep water sea stars from the northern Gulf of Mexico.
- Slattery, M. and I. Bosch. Possible costs of brooding and embryonic development in an antarctic sea star.

- Pawson, D.L. Incidence and Inheritance of Situs Inversus Viscerum in Echinoids.
- Jaeckle, W.B. Multiple modes of asexual reproduction by oceanic sea star larvae.
- Balser, E.J., E. Ruppert, and W. Jaeckle. Ultrastructure of the hydrocoel of holothuroid auricularia larvae.
- Janies, D. and L. McEdward. Highly derived coelomogenesis in *Pteraster tessellatus*. Implications for the evolution of the concentricycloid water-vascular system.
- McEdward, L., and D. Janies. Life cycle evolution in asteroids: what is a larva?
- Beddingfield, S.D., J.B. McClintock, K. Marion and T.S. Hopkins. The foraging ecology of the armored sea star *Astropecten articulatus*.
- Forcucci, D. Population dynamics and 1991 mortality event of *Diadema antillarum* in the Florida Keys.
- Valentine, J.F., and K.L. Heck Jr. Herbivory and sea grass distributions: evidence for sea urchins controlling turtlegrass distribution and abundance.
- Norris, D.R. Distribution, growth and sediment interactions of an infaunal deposit feeder (*Mareia planulata* Lamarck) in a tropical soft bottom community.
- Stancyk, S.E., W.E. Dobson, and R.B. Aronson. Sublethal tissue loss by *Ophiura sarsi* on the North Carolina Continental Slope.
- Dobson, W.E., and S.E. Stancyk. Use of ophiuroid vertebral ossicle bands as biological markers for population age and sublethal predation studies.
- LeClair, E.E. Brittle-star baffles: flow patterns in dense arrays of model ophiuroid arms and their consequences for suspension feeding communities.
- Fox, D.J. Comparative morphology of the central arm ossicles of *Ophiocoma wendti* and *O. echinata*.
- Shlepr, M.G., and R.L. Turner. Early ossicle growth in a brittle star: a re-examination of the syncytial requirement.
- Lawrence, J., B. Robbins and S. Bell. A comparison of the sizes of the pieces of the Aristotle's lantern in five species of *Strongylocentrotus*.

- Polson, E.S., L.L. Robbins, J. Lawrence. Shell matrix proteins- a potential tool for investigating phylogenetic relationships in Echinodermata.
- Pomory, C.M. Echinoderms from the shallow waters of Texas.
- Messing, C.G. Diversity and ecology of comatulid crinoids (Echinodermata) at Madang, Papua New Guinea.
- Heyman, R.M., and R.L. Turner. North Pacific ophiurans in the North Atlantic.
- Stickle, W.B. Towards a better understanding of six rayed sea stars of the genus *Leptasterias* from the west coast of North America.
- Mooi, R. and B. David. Deep history-phylogenetics of deep-sea urchinid echinoids.
- Hendler, G. 150 million years of swimming brittlestars, in brief.
- Hotchkiss, F.H.C. Footnotes on Loven's Law.
- Blake, D.B. Evolutionary relationships of stelleroids.
- Bryan, P.J. and J.B. McClintock. Population structure and organ indices in *Lytechinus variegatus* from contrasting habitats within St. Joseph's Bay, Florida: a preliminary survey.
- Hamel, J.-F. Relation between reproductive cycle of the sea cucumber *Psolus fabricii* (Duben and Koren) and environmental conditions.
- Komatsu, M., C. Oguro, and J. Lawrence. Development of the sea star *Echinaster spirulosus*.
- Lee, K.J., C.D. Bishop and S.A. Watts. Na<sup>+</sup> K<sup>+</sup> ATPase activity in the gut of the regular echinoid, *Lytechinus variegatus*.
- Messing, C.G. Postural, Distributional and Taphonomic Variations Relative to Current Flow and Topography in an Assemblage of Living Stalked Crinoids.
- McClintock, J.B., S.A. Watts, K. Marion, T.S. Hopkins and G. Schinner. Reproduction in a deep water ophiuroid from the Gulf of Mexico.
- Schinner, G.O. and J.B. McClintock. Functional morphology of petaloid brood pouches of the antarctic sea urchins *Abarus nimrodi* and *A. shackletoni* (Echinoidea: Spatangoida).

1991 ANNUAL MEETING OF THE GEOLOGICAL SOCIETY OF AMERICA -- SAN DIEGO, CALIFORNIA, October 21 to October 24, 1991. Geological Society of America Abstracts with Program 23(7). (communicated by W.I. Ausich).

- Ausich, W.I., and T.K. Baumiller. Did muscular articulations have muscles in advanced cladid crinoids?
- Beadle, S.C. Heterochrony and eccentricity: A new model for the oral surface morphology of dendrasterid sand dollars.
- Baumiller, T.K. Importance of hydrodynamic lift to crinoid autecology, or could crinoids function as kites?
- Llewellyn, G., and C.G. Messing. Local variations in modern crinoid-rich carbonate bank-margin sediments.
- Messing, C.G. Variations in posture, morphology & distribution relative to current flow and topography in an assemblage of living stalked crinoids.
- Sprinkle, J. Origin of echinoderms in the Paleozoic evolutionary fauna: New data from the Early Ordovician of Utah and Nevada.
- Sumrall, C.D. Plate morphology in stalker edrioasteroids.
- Taylor, W.L., and C.E. Brett. Silurian crinoid Lagerstätten: Taphonomic and ecologic windows.
- Watkins, R. Tiering and guild structure in a high-diversity Silurian marine community.

199 NORTHEASTERN SOUTHEASTERN SECTION OF THE GEOLOGICAL SOCIETY OF AMERICA HARRISBURG, PENNSYLVANIA, March 26-28, 1992. Geological Society of America Abstracts with Program 24(3). (communicated by William I. Ausich)

- Brower, J.C. Growth and functional morphology of Euptychocrinus skopaioi, a dwarf camerate crinoid from the Ordovician.
- Greenstein, B.J. A temporal gradient of taphonomic overprint and the diversity history of the family Cidaridae (Echinodermata: Echinoidea).

1992 NORTH-CENTRAL SECTION MEETING OF THE GEOLOGICAL SOCIETY OF AMERICA IOWA CITY, IOWA, April 30 to May 1, Geological Society of America Abstracts with Program 24(4). (communicated by William I. Ausich)

Blake, D.B., and T.E. Guensburg. Caught in the act: A Late Ordovician asteroid and its pelecypod prey.

Meyer, D.L., and T. Oji. Experimental taphonomy of a recent stalked crinoid: implications for the crinoid fossil record.

Schumacher, G.A., and M.R. Caudill. Combined flow storm-generated crinoid taphonomy: an example from the Upper Ordovician, Cincinnati Series in northeastern Kentucky.

Terry, R.E. Echinoderm taphofacies within the Banff Formation (Lower Carboniferous; Alberta): Western versus eastern facies belts.

FIFTH NORTH AMERICAN PALEONTOLOGICAL CONVENTION CHICAGO, ILLINOIS, June 28 to July 1, 1992, Fifth North American Paleontological Convention Abstracts and Program, The Paleontological Society Special Publication 6. (communicated by William I. Ausich)

The Cretaceous crinoid Uintacrinus was part of the logo for the meeting, appearing on advertisements, the abstracts volume, and tee shirts.

Baumiller, T.K. The energetics of passive suspension feeding: ecological and evolutionary consequences for crinoids.

Brower, J.C. Ontogeny and functional morphology of Eoparisocrinus crossmani, a cladid crinoid from the Middle Ordovician

Donovan, S.K., and C.J. Veltkamp. A Rhuddanian (Silurian: Lower Llandovery) echinoderm fauna from Haverfordwest, southwest Wales

Foote, M. Early morphological diversity in blastozoan echinoderms.

Guensburg, T.E., and J. Sprinkle. Environmental controls of rapidly diversifying echinoderms during the Early Paleozoic.

Holterhoff, P.F. Ecophenotypic variation and phylogeny within the Erisocrinaceae (Crinoidea): linkage of morphology, ecology, and sea-level in the Late Paleozoic.

Kammer, T.W., and W.I. Ausich. Demise of the middle Paleozoic crinoid fauna: gradual or mass extinction?

Maples, C.G., J.A. Waters, N.G. Lane, and H.-f. Hou. Paleobiogeographic significance of Famennian echinoderm faunas from northwestern China.

Nebelsick, J.H. Actupaleontological investigations of shallow water Red Sea echinoids.

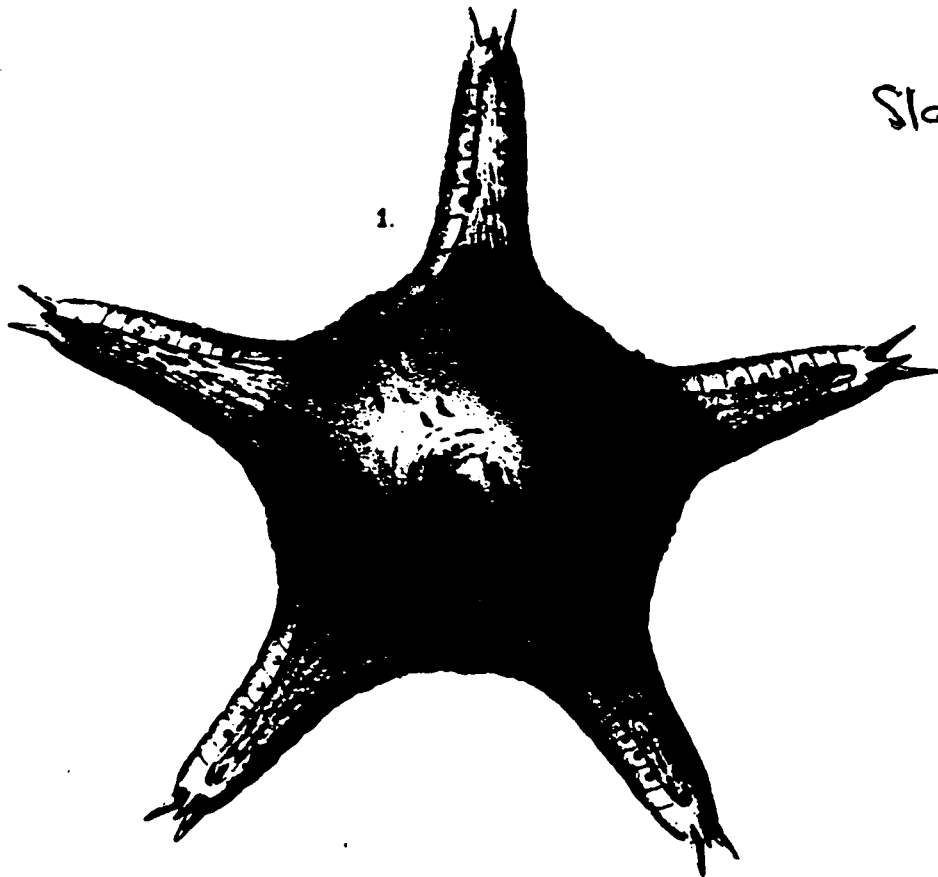
Oji, T., and S. Amemiya. Long survival of stalk pieces of Metacrinus rotundus Carpenter, a modern stalked crinoid, in an aquarium.

Polson, E.S., J. Lawrence, and L.L. Robbins. Shell matrix proteins -- a potential tool for investigating the phylogenetic relationships of Echinodermata.

Smith, A.B., and R. Christen. Morphological and molecular rates of evolution in post-Paleozoic echinoids.

Sumrall, C.D., and J. Sprinkle. Could edrioasteroids move?

Terry, R.E., and D.L. Meyer. A unique taphonomic profile for Lower Carboniferous crinoids of western Canada and possible Recent analogs.





## AILSA'S SECTION

### Echinoderms in culture

Reinhard, J. 1992. Sacred peaks of the Andes. National Geographic. 181 (3). p. 93: "The Cuzco market sells plenty of the small sacrificial bundles called *despachos*, like the ones the yatiris had taken to Illimani. These may contain items such as starfish, cookies, minerals, miniature metal figurines, seashells, incense, llama fat, and cocoa leaves and are remarkably like the offerings I have dug up from centuries-old Inca ceremonial platforms on the peaks."

### Echinoderms in literature

Durrell, L. 1990. Caesar's vast ghost. Arcade Publishing, N.Y. "The virtue of such a beginning will become obvious when one wants to extend one's travels, for Arles is like a starfish in a central position, extending its arms in all directions."

### Echinoderms in art

William Hogarth (British, 1697-1764): "The Mackinnen Children, 1747" Elizabeth is holding a fold in her skirt that contains various shells and, prominently, the clean test of a sea urchin. As the sea urchin has an oblong shape, it is almost certainly *Echninometra*. (contributed by Fred Hotchkiss)

### Echinoderms in poetry (?)

(communicated by Penny Barents)

#### SONG/ODE TO A SEA-DAISY

Daisy, Daisy, give me your answer, do.  
I'm half crazy over the Phylogeny of you.  
You won't make a stylish Cladogram.  
With your shape why should you give a damn.  
But we might have known,  
With that sperm of your own,  
You'd out-class your relatives too.

# 4) DEUX RECETTES POUR PREPARER LE RORI

TAHITI

## RORI A LA CHINOISE

Recette du Jade Palace.

Les «Rori U» sont achetés déjà séchés. C'est l'opération que nous avons montré hier. Ils sont dans un premier temps cuits à l'eau pendant trois heures, puis trempés dans l'eau froide pendant 18 heures.

Ensuite on remet à bouillir 3 heures et de nouveau tremper 18 heures dans l'eau froide, puis rebouillir jusqu'à trois ou quatre fois.

On laisse sécher et l'on découpe en morceaux les «Rori U».

Les morceaux sont ensuite sautés à la poêle, puis l'on mélange les morceaux avec de la sauce soja, de l'huile de sésame, du jus de gingembre et du rhum.

On mélange le tout que l'on fait cuire 3 à 5 minutes à l'étouffée. C'est à ce moment que l'on peut ajouter des champignons ou des ablonas ; dernière cuisson de 3 mn à l'étouffée. Servez c'est prêt à manger.

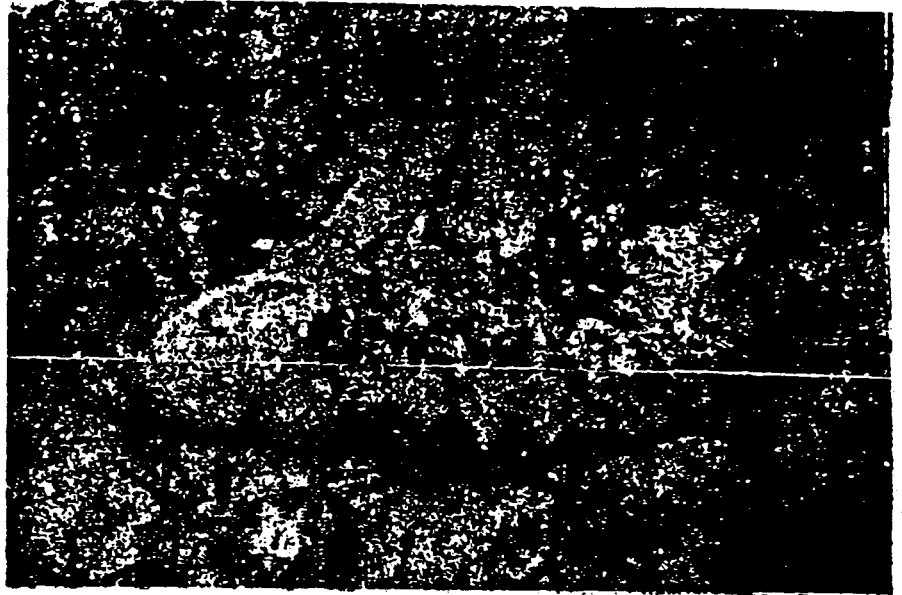
## RORI A LA TAHITIENNE

C'est le plat «Rori Taioro», que l'on peut trouver le dimanche matin tout préparé au marché.

Les espèces d'holothuries utilisées ne sont pas les «Rori U» mais celles plus faciles d'accès qui peuplent nos lagons à faible profondeur.

Laver le «rori», enlever les viscères et les gonades, ôter la peau, découper le rori en morceaux.

Faire bouillir à feu tahitien



(jusqu'à 4 heures) de manière à ce que les morceaux soient bien tendres.

Le taioro est du coco râpé très fin, auquel on mélange le jus exprimé de 2 à 3 têtes de chevret-

tes écrasées dans un linge. Laisser fermenter une nuit dans un récipient couvert de feuilles de burau.

On mange les morceaux de «rori» accompagnés de cette sauce.



### Consumer usage

The cook's preparation for beche-de-mer takes on some of the aspects of persuading a sponge to take up a lot of liquid. After all the care that the processor takes to dry it for storage, the cook must take equal care to reconstitute the product to obtain a high quality dish.

There are thousands of cookbooks but few present information on such an exotic collection of foodstuffs as "Unmentionable Cuisine" by Calvin W Schwabe. (published by the University Press of Virginia, Charlottesville, Virginia, 22903). Even if you have no intention of cooking, you would enjoy this book. Dr Schwabe gives the following directions for the preparation of sea cucumbers:

### Recipe

#### Basic preparation

Dried sea cucumbers, if not purchased pre-soaked, must be treated by a fairly involved process. First, soak them for four hours in cold water and then scrub them with a brush. Place them again in cold water, bring to boil, cook for five minutes, and allow to cool in the water again. Repeat this process ten times. The meat then is swollen and soft and ready to use in the following recipe:

#### Stewed sea cucumbers (Dun hoi sun)/China

Place eight prepared sea cucumbers in cold water. Bring to boil, simmer five minutes and drain. Then simmer them for about 20 minutes in about three cups chicken stock and cut the drained meat into large pieces. Discard the stick. Stir-fry the meat for about two minutes with a little light soy sauce in some hot oil in a wok. Remove the meat. To another wok add some fresh oil, heat it, add two green onions cut into 1½ inch length, about six thin slices of ginger, two tablespoons sherry, and the sea cucumbers. Stir-fry a bit and add three tablespoons light soy sauce, dashes of pepper and Ajinomoto, ½ teaspoon sugar, ½ tablespoon dark soy sauce and ½ cup fresh chicken stock. Cook on high heat for two minutes. Add one tablespoon cornstarch dissolved in two tablespoons cold water and ½ tablespoon sesame oil. Stir well and serve.

Mrs Bruce, a freelance writer, researched and wrote several articles on fish species and seafood for INFOFISH during a recent prolonged stay in South East Asia.

N° 6/83

Communicated by  
Chantal Couand



92

1972, t. 33

MARIA ELENA CASO

Rev. Soc. Mexicana  
de Historia Natural

coeficiente de 0.65; es decir, presenta casi los dos tercios de su valor alimenticio.

El producto de la región indopacífica contiene 15 a 30% de cenizas, 35 a 52% de proteínas y de 21 a 23% de agua; los carbohidratos faltan, pero existe una pequeña cantidad de grasa (Greshoff y Sack, 1900; Greshoff y van Eck, 1901).

El producto del Mediterráneo parece ser aún más nutritivo, contiene de 56 a 65% de proteínas; 13 a 24% de cenizas, cerca de 0.7% de grasas y 10 a 11% de agua (Sella, 1940). Según Frankel y Jellinek (1927), la proteína, que existe en el *trepang*, es completamente soluble en pepsina, por lo que este producto parece ser muy fácilmente digerible. En algunos lugares de la región indopacífica, los holoturoideos son irritados hasta provocar la evisceración y los tubos de Cuvier y las gónadas, son entonces comidas en crudo.

El material ya seco es cortado en pequeñas piezas las que son usadas para condimentar las sopas o estofados y se dice que imparten un delicioso aroma. Cuando el *trepang* es cocinado, cada pedazo se hincha y adquiere un aspecto gelatinoso.

#### RECETA DE UNA SOPA HECHA CON TREPANG

Esta receta ha sido tomada del libro de Cherbonnier G., 1954, págs. 111, 112. "Se remojan a las holoturias en agua fría durante 4 horas y se les quita la delgada piel que las cubre, la cual se separa de ellas con relativa facilidad. Para 500 gramos de holoturias, se requeman 3 centilitros de aceite, en el que se freirán hasta dorarse, 20 gramos de ajos y 200 gramos de carne de cerdo, la cual se habrá cocido con anterioridad y rebanado en rodajas delgadas. Cuando está todo junto, se añaden las holoturias y se les deja cocer a fuego lento por espacio de 5 minutos. A todo lo anterior, se añade un cuarto de litro de consomé de pollo, una cucharada de sopa de soya y se deja cocer todo ello, a fuego lento, durante unos 15 minutos.

Lo expuesto anteriormente, no pretende dar una explicación detallada, de lo que hasta ahora se conoce con el nombre de *trepang*, ni mucho menos. Tan sólo se ha tratado de reunir algunas de las referencias más interesantes, que tuvieran algún interés, desde un punto de vista de divulgación general, con el fin de dar a conocer este interesante y peculiar producto marino.

76

*Verdura cinese o cavolo di Tientsin, con trepang.*

In un po' d'olio si frigge la verdura (cruda). Salare. Un momento prima di servire si aggiungerà il trepang.

*Costole di maiale (ossa lievemente ricoperte di carne) con trepang.*

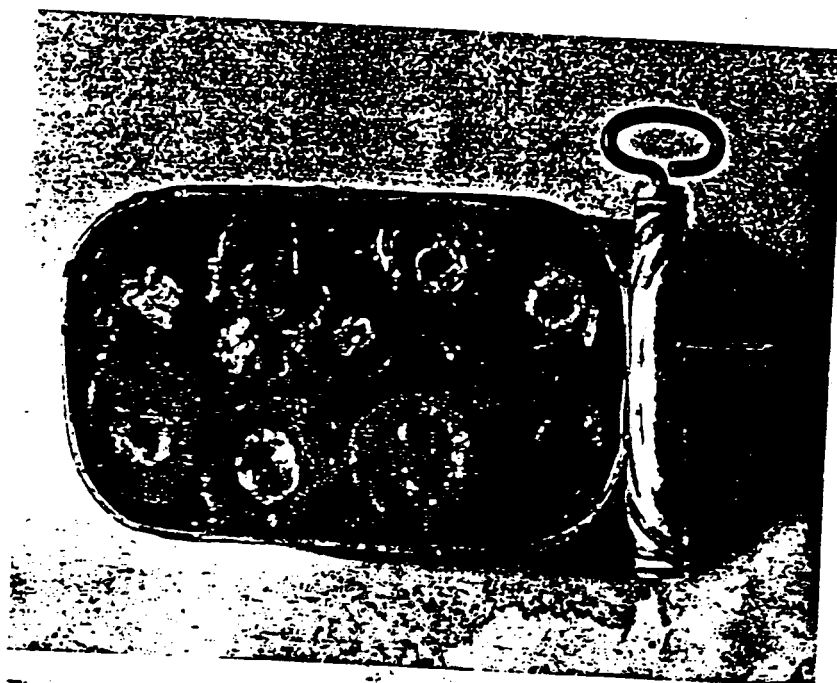
Si friggono ben bene le ossa di maiale, spruzzandole con un po' di salsa di soya, dopo di che s'aggiungerà un pizzico di farina ed un po' di brodo. Sale, pepe. Si lascia cuocere per molto tempo. Un momento prima di servire si aggiungerà il trepang.

Per secoli e secoli, fatto curioso, il trepang è rimasto un cibo esclusivamente cinese, quasi un ghiotto attributo di quella razza e le altre popolazioni marinare, pur avendo a portata di mano oloturie a dozzina, non ne hanno imitato l'esempio. Chi di essi ha ragione? Probabilmente hanno ragione i cinesi e il loro segreto deve consistere in una certa delicatezza, non facilmente imitabile, nel modo di preparare la vivanda, se è vero questo aforisma di una graziosa giapponesina: « Gli europei cucinano per il cervello, i giapponesi per gli occhi ed i cinesi per il palato ». Ma nei tempi odierni, molti cambiamenti che non si sono prodotti durante secoli possono verificarsi in pochi anni.

Le oloturie seccate e indurite, scrive Seale, sembrano imman- giabili e solo quando siano state trasformate in una zuppa deliziosa dalle abili mani di un cuoco cinese, si comprende il reale pregio di questi frutti del mare. I quali vivono nei giardini di alghe sulla bianca sabbia e fra i coralli, nutrendosi di animalletti marini e di vegetali, e non v'è perciò ragione che non debbano appartenere alla classe dei cibi squisiti e non entrino nell'uso comune degli europei e degli americani ».



Fig. 7.

Fig. 8 - Anelli di *H. tubulosa* fresca spellata, in salsa di pomodoro piccante.

Carbajal. 1900. Zoological Record. Fuegians eat sea-urchins and starfish in fabulous quantities, til their stomachs are tight as drums, not withstanding continued indigestion due to economical methods of cooking.

in : Sella & Sella 1940

14

L'industria del Trepang  
Communicated by Charles Conrad

Lo « Hai-shen » (trepang) viene immerso nell'acqua per qualche giorno prima di usarlo, quindi viene bollito fino a che sia diventato morbido. Solamente dopo questo processo le olturie vengono tagliate trasversalmente in tanti pezzi della lunghezza di un pollice (non risulta che venga mai grattugiato) ed aggiunto alle pietanze sempre un momento prima di servirle. Lo « Hai-shen » per se stesso non ha nessun sapore e può essere aggiunto a qualsiasi pietanza, sia carne (di pollo o di maiale) o siano verdure. Non ha la proprietà di rendere le pietanze gelatinose. Sul mercato può essere acquistato già rammollito. I ristoratori cinesi lo cucinano soltanto quando viene ordinato appositamente.

RICETTE

*Brodo di trepang.*

Nel brodo di pollo aggiungere il trepang tagliato a pezzi. Salare se vi è bisogno e servire caldo.

*Trepang in salsa bianca.*

A una certa quantità di brodo di pollo, aggiungere un po' di salsa di soya, un po' di farina, un pizzico di pepe ed unirvi il trepang tagliato a pezzi; lasciar cuocere fino a che quest'ultimo abbia preso bene la salsa. Servire caldo.

*Trepang in salsa di pomodoro.*

Nel brodo di pollo sciogliere la salsa di pomodoro. Sale e pepe ed un pizzico di farina. Mentre il tutto bolle aggiungervi il trepang tagliato a pezzi e lasciar cuocere. Servire.

*Trepang con polpette di carne di maiale.*

Si tritura la carne di maiale alla quale si aggiungerà sale, pepe ed un pochetto di zenzero, un po' di cipolline novelle tritate, alcune castagne d'acqua pure tritate finemente ed un giallo d'uovo. Farne delle polpette e friggerle. Allorchè saranno ben fritte, si ritireranno dalla padella e si terranno al forno ad una temperatura mediocre. Nel sugo che sarà rimasto nella padella mettere mezzo cucchiaino di farina, alcuni funghi di bambù (rammolliti), un po' di brodo ed un poco di salsa di soya. Dopo un momento di cottura, vi si rimetteranno le polpette ed il trepang. Si lascerà cuocere il tutto un momento ancora e si servirà.

*Pollo e castagne con trepang.*

Si frigge il pollo tagliato a pezzetti. Si aggiungono le castagne crude, alcuni funghi di bambù (rammolliti), mezzo cucchiaino di farina, un po' di salsa di soya, brodo, sale e pepe. Si lascia cuocere il tutto ed in ultimo, un momento prima di servire, vi si getterà il trepang.

*Rognoni di maiale con trepang.*

Dopo aver preparato i rognoni di maiale per la cottura, si taglieranno per metà. Fare alcuni intagli sulla superficie, friggere un momento, aggiungervi un bicchierino di cognac, un poco di salsa di soya, ed in ultimo il trepang. Servire caldo.



- 1) Make a thick sauce with white sauce, cream, and smashed anchovies. Add salt, nutmeg or the spices you like.
- 2) but recently made pancakes into thin ribbons like spaghetti, mix with sauce. Serve hot with very cold Chilean wine.

Have luck!

Laura Post

(Alberto Lanain's mother)

CONCEPCION, CHILE 20 March 1993

13 JUNE 1992

PUESTO DE MARISCOS Y PESCADOS "APA"<sup>S</sup>



Centollas - Ostras - Erizos  
 Jaibas - Picorocos - Almejas  
 Cholgas - Potache - Curanto  
 Cancato - Sopa Marina  
 Pescado Frito - Moches - Camarones

ATENDIDO POR SUS DUEÑOS  
 Daniel Adonay Pino Hernández  
 y Sra.

LOCAL 37 - ANGELMO - PUERTO MONTT - CHILE

# Pate de Erizos



yadran

Fabricado en  
Panamericana Norte 482 - Santiago - Chile  
Teléfono: 361011 Telex: 47-57 PYSOL CL  
REGISTRO S.M.A.D. EN CHILE N. 3111 31.03.81  
PRODUCTO DE CHILE

ERIZO  
NATURAL

yadran

yadran

# ERIZO NATURAL

ERIZO  
NATURAL

CHILE



Ingredientes: Erizo, Azúcar, Sal, Levadura.  
Características: Erizo, Azúcar, Sal, Levadura.  
Peso Neto: 250 g.  
Puede ser congelado.  
Duración: 4 años

## **Recipes for sea cucumbers - a little bit different**

(from K.Y. Arakawa: A Handbook on the Japanese sea cucumber: Its Biology, Propagation and Utilization (In Japanese), Midorishogo, Tokyo)

### **1) Kodatami**

.....one of the eldest sea cucumber recipes in Japan

- cut a raw sea cucumber in pieces or slices and put for some time in sake (jap. rice wine)
- drain the sea cucumber pieces and season with a mixture of dashi (jap. soup stock), mirin (sweet sake) and salt
- serve with grated wasabi (jap. hot green radish)

### **2) Kaki-kodatami**

like above recipe but add also some oysters to the sea cucumber pieces and season in the same way

### **3) Koubashira and konawata**

- wash koubashira (adductor muscle of small scallops) in lightly salted water and drain
- mix 100 g of konowata (salted entrails of sea cucumbers) with two egg white, put into a vessel and steam. Drain.
- heat the konowata in a pot until water is evaporated
- mix with koubashira

### **4) Koubashira, iriko and cucumber in vinegar dressing**

- wash iriko (dried sea cucumber, also known as trepang) in water
- put into water with charcoal ash added and simmer on low heat for 3 days (!)
- cut the now soft iriko into halves, wash the inside thoroughly and cut into small pieces
- fry with some salad oil and salt lightly
- wash koubashira and drain
- cut cucumber into small pieces
- mix iriko, koubashira and cucumber and season with white vinegar

**5) sea cucumber and vinegar**

.....one of the traditional recipes from the Hiroshima region

- cut sea cucumber in pieces or slices, mix with cucumber and fried anago (conger) and season with sambaizu (mixture of soy sauce, vinegar and sugar).
  - serve with grated daikon (jap. white reddish)
- (the fried conger can be replaced with other fried fish)

**6) Namako-tempura (deep-fried sea cucumber)**

.....another highlight from the Hiroshima region

- cut big sea cucumber in rather thick pieces
- put in boiling water, boil quickly and drain
- cover pieces with flour and deep-fry
- eat while still hot

**7) Yaki-namako (fried sea cucumber)**

.....surprises you with the simple beauty of the countryside

- wash a clean sea cucumber in salt water and put it as a whole onto a charcoal grill
- when done eat without any seasoning

**8) Shaburi-namako (shaked sea cucumber)**

.....soft and good to keep

- put sea cucumber in a sieve and add 10 round pebbles and some salt
- shake for 20 - 30 minutes
- remove entrails, put sea cucumber into warm bancha (green tea) and let get cool.
- cut the considerably softened sea cucumber into pieces and season with sambaizu (see recipe 5); keep in refrigerator
- serve with fried conger and cucumber on a small plate and season with ponzu (mixture of bitter orange juice, soy sauce and vinegar); also add some grated daikon (white reddish) and chili pepper

# なまご謠本

つたてられた生物等口傳の種および利用

著者 荒川 好徳

なまごの謠本

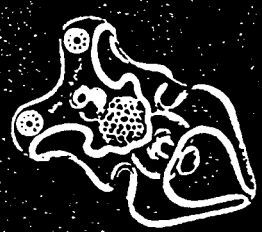
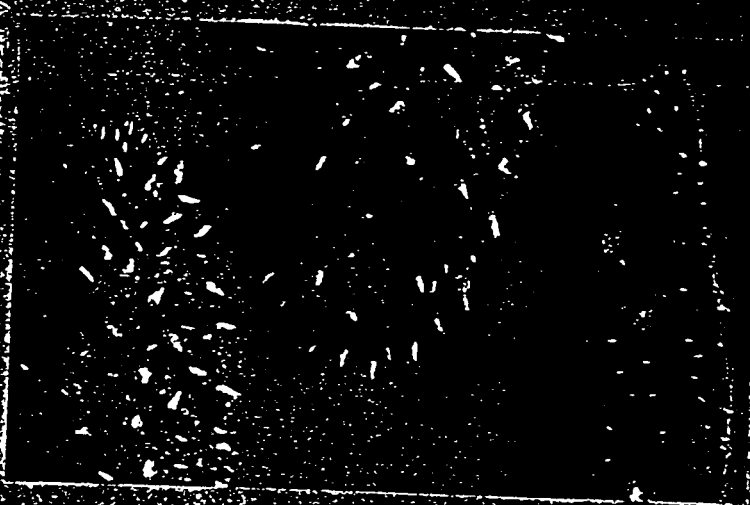
荒川 好徳 著

圖書房

ISBN 4-89531-409-X 02049 P1650E

定価 1650円 (税別 1602円)

A Handbook on the Japanese Sea Guppies  
(Burger's Guppies) *Amphiprion*  
by  
Kohzo ARAKAWA, D.S.S.



ナマコ切手エックリスト

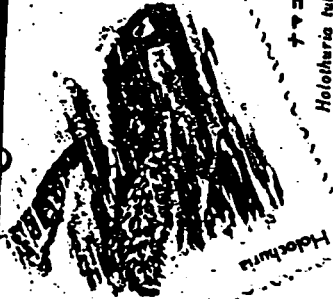
(1989年11月現在)

ハーブグダ1987年発行  
 棘皮動物3種セットの1  
 ナマコの1種  
*Holothuria* sp.

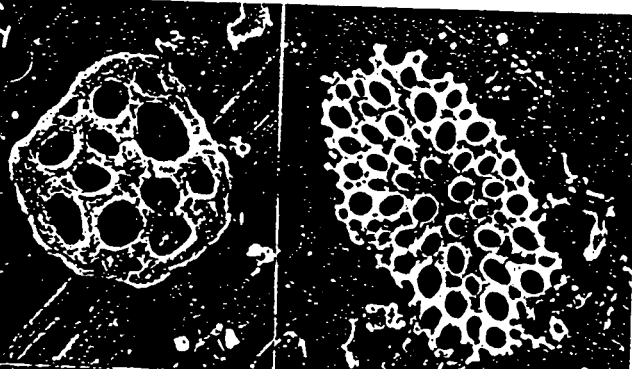


ナマコの産卵

アルバニア1966年発行  
 ナマコの1種  
*Holothuria tuberosa*



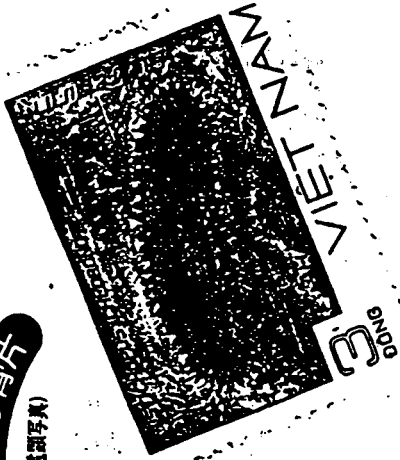
ナマコの骨片  
 (走査電顕写真)



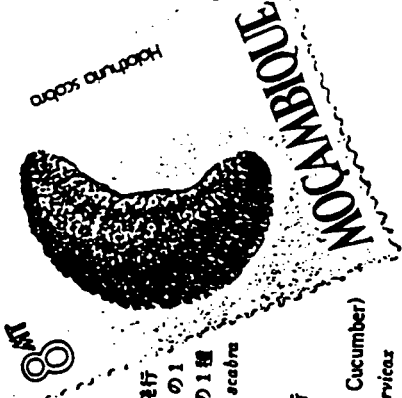
ブエトナム1985年発行  
 棘皮動物7種セットの2種  
 ナマコの1種  
*Holothuria monocaria*



ナマコの1種  
 (An Aspadochirotid Sea Cucumber)  
*Stichopus chloronotus*



モザンビーク1983年発行  
 棘皮動物6種セットの1  
 ナマコの1種  
*Holothuria scabra*



パラオ1989年発行  
 ナマコの1種  
 (Dappled Sea Cucumber)  
*Holothuria parvicar*

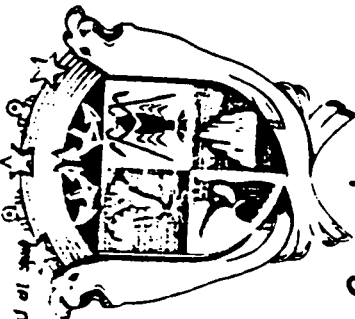


TERRES AUSTRALES  
ET ANTARCTIQUES FRANCAISES

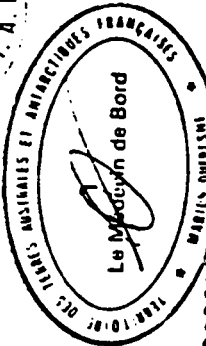
OP 93-3.1

Courrier postal à bord

Posté au bord

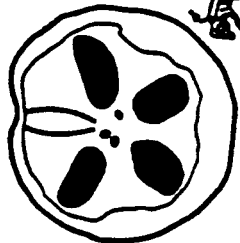


Le Commandant  
Jean-Paul AUFRANT



BIODIVERSITE

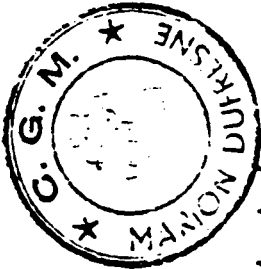
GENETIQUE



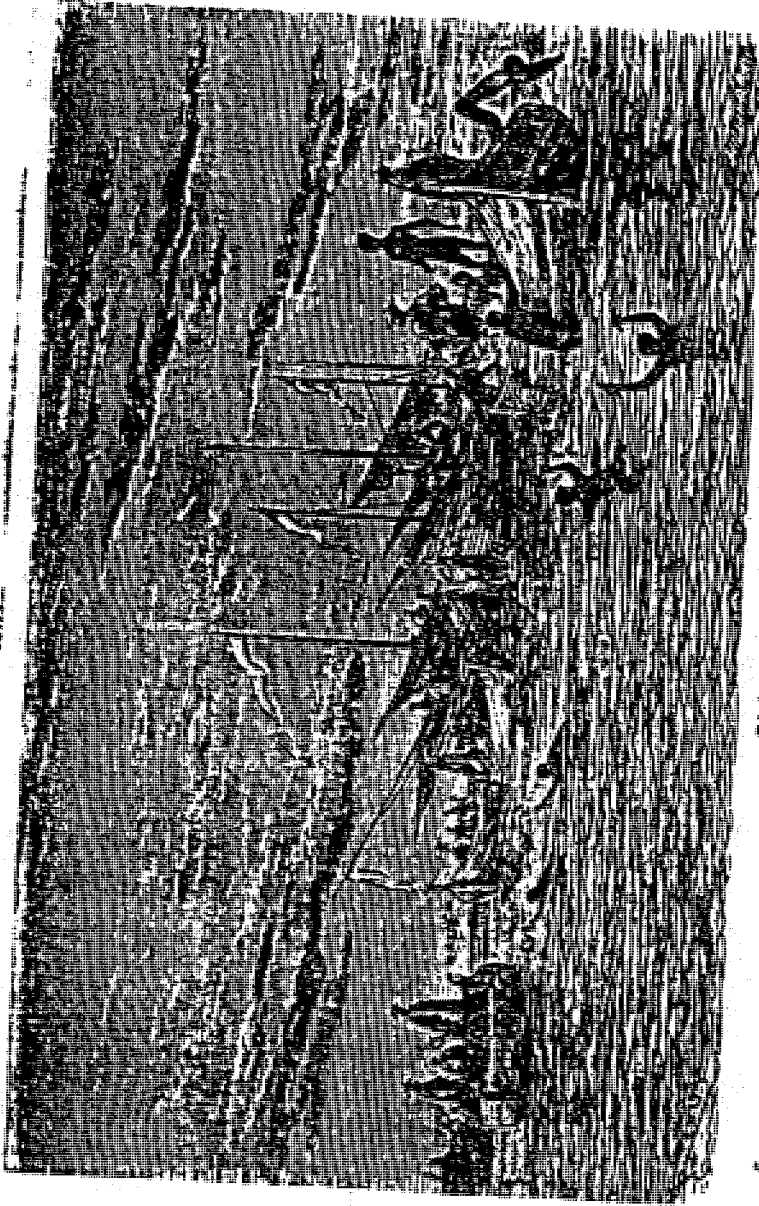
KERGUELEN

1992-1993

EVOLUTION  
RAPporteur Spécial du Sénat  
HENRI GUETSCHY EN MISSION  
AUX TERRES AUSTRALES



John L. Lawrence  
Department Biology  
University South Florida  
Tampa  
Florida 33620 U.S.A.



Pêche de l'holothurie.

(3)

Bibliothèque des Merveilles - Les grandes pêches - Mounier-Huchette 1871

Chantal Comand



in: MULVANEY - The prehistory of  
Australia

(3)

Plate 7 A scene purporting to be Port Essington, but with an East Indian terrain, around 1843-5, by Harden S. Melville, draftsman on the surveying voyages of H.M.S. Fly. Despite the unreal background and the brace of wildfowl (who fly across several of Melville's drawings) the scene contributes details of the trepanning industry. Note the lines of flat iron cauldrons, pottery, ladles and smoke houses. The Aborigines, bottom left, wear arm bands and shell ornaments of a characteristic form.



Communicated by  
Oscar Conrad

# Pacific Daily News

©1992 Guam Publications, Inc.

VCL 23 NO. 135 AGANA, GUAM, AUGUST 4, 1992

Hafa Adai, it's Tuesday

## Biba balaté

**Rare find:** Colorful sea cucumber found near Guam thrills UOG scientists

By LINDA AUSTIN  
Daily News Staff

A brilliant red and white candy-striped sea cucumber discovered last weekend at Glass Breakwater is wowing marine scientists who have never seen its kind in Guam waters.

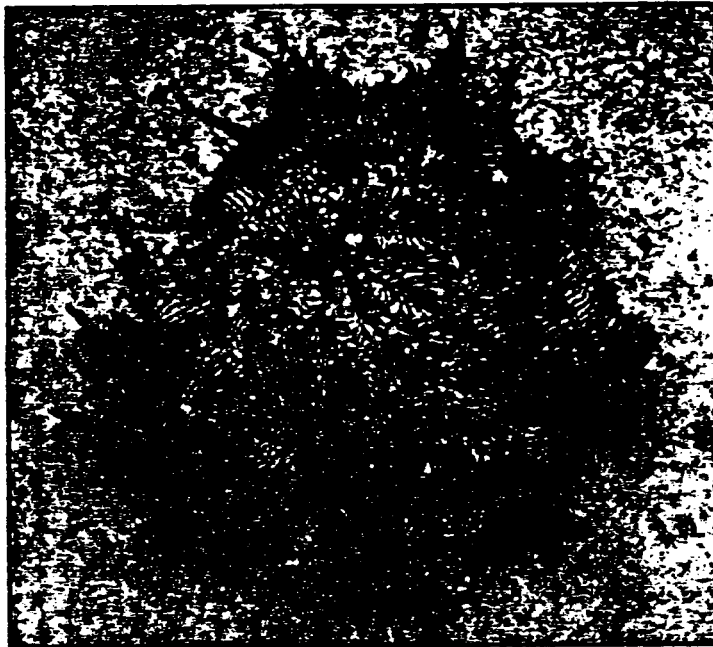
"It's a beautiful animal ... just spectacular," University of Guam's sea-cucumber expert, Gustav Pauley, said about the two-foot-long creature, scientifically called *Thelelenota rubralineata*.

"That means the red-striped spiny backed one in Latin," quipped graduate student Alex Kerr.

Kerr's diving partner, Peter Schupp, discovered the harmless animal in about 200 feet of water while the pair were searching for deep-water fish, they said.

The species is a new face on the international scene, discovered less than a year ago in the shallow waters near Papua New Guinea and Indonesia. The discovery of a Guam version snuggled in deep waters suggests the critters are more common and more widely dispersed than anticipated, Kerr said.

Sea cucumbers, known as balaté in Chamorro, are humble simple creatures with "not a whole lot of gray matter (brain) accumulation," Kerr said. "They pretty much just process sediment" and keep the reef face clean.



Cheryl Mitchell/Daily News Staff

A diver discovered this conspicuous candy-striped sea cucumber last weekend at Glass Breakwater in about 200 feet of water. See related photo/Page 4.

Still, they can perform amazing feats, especially when fleeing their main predator, sea snails. In the art of escape, balatés have been known to leap-frog to freedom or break off pieces of skin and spines to distract the gastropods while the main body flees, Kerr said.

balatés also come in male and female varieties, but because their reproductive organs are located on their heads, sometimes determining who's what can be tricky, Kerr admitted yesterday while tipping the striped one end over end looking for a clue.

For a while, Guam's latest celebrity will enjoy carefree days lounging at UOG's marine lab while Kerr incorporates this new discovery in his research papers.

Then, he or she will be "pickled — but not to eat," Schupp said. Such specimens are needed as scientific evidence and for future research, the pair said. Interest in the soon-to-be specimen has come as far away as the Smithsonian Institute but the balaté probably will remain in UOG's marine archives, they added.



*In communities there are little players  
and big players, and the biggest players  
of all are the keystone species. As the  
name implies, the removal of a keystone  
species causes a substantial part of the  
community to change drastically.*

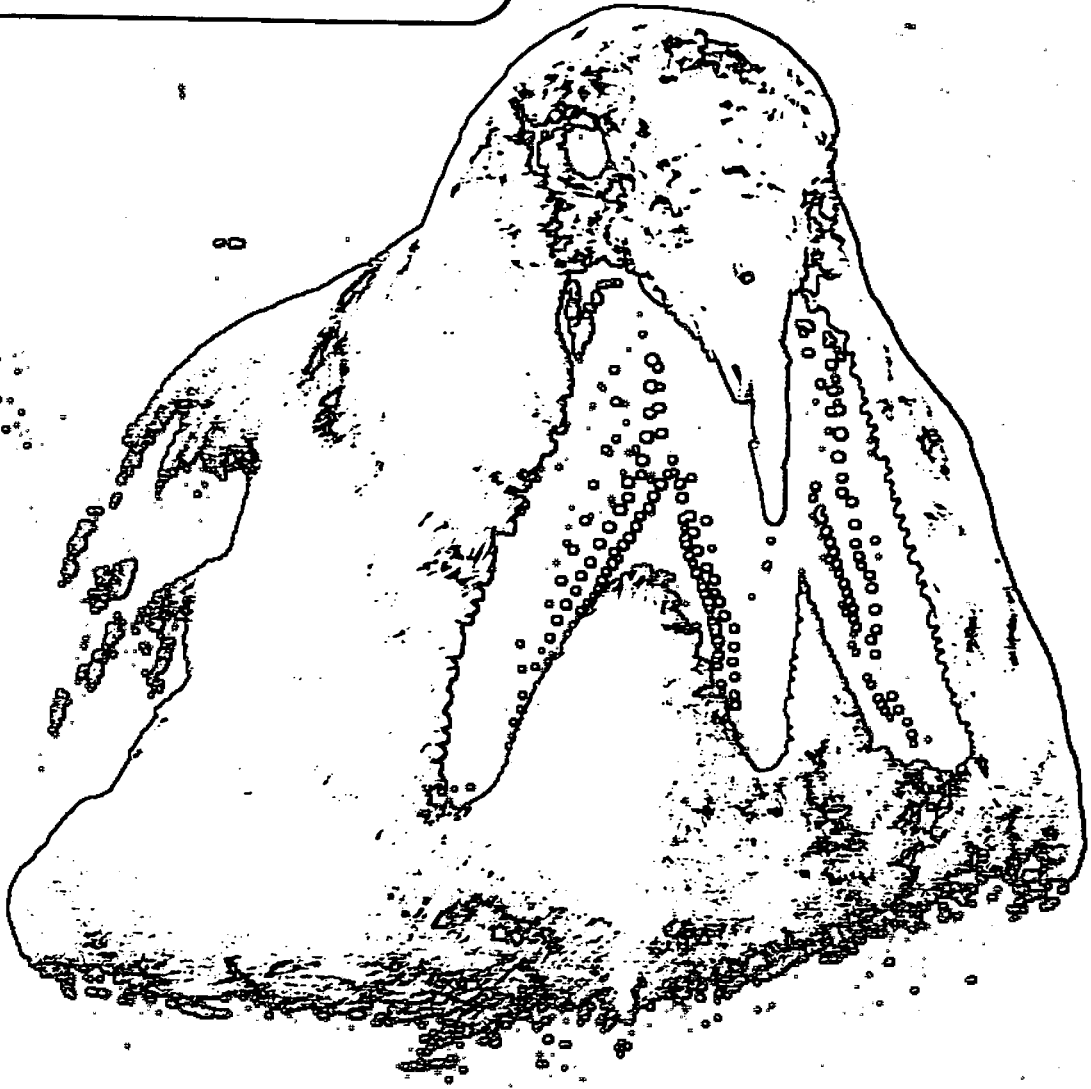
G.O. Wilson 1992.  
The diversity of  
life.  
Belknap /  
Harvard

130

\*\*\*\*\* 5-DIGIT 33613  
N12 MEMBERS 117843444 EXPL9310  
ADAM P SUPPERS  
2011 GREGORY DR  
TAMPA, FL 33613-2542

MSV1

00826





DIVISION OF INVERTEBRATE ZOOLOGY  
AMERICAN SOCIETY OF ZOOLOGISTS

NEWSLETTER

FALL 1992

GREAT INVERTEBRATE ZOOLOGISTS:  
ALEXANDER EMMANUEL RODOLPHE AGASSIZ (1835-1910)

(by John P. Wourms)

Although a legend in his own time, Alexander Agassiz now is only vaguely remembered as a pioneer oceanographer. Yet, the enigmatic figure of the "less celebrated Agassiz" is worth closer study for he greatly influenced the conduct and infrastructure of zoology in 19th century America. Not only was he a prime mover in oceanography, but he played a pivotal role in the emergence of embryological research in America, and was deeply involved in the systematics of echinoderms, especially the echinoids. The latter stages of his career were devoted to the study of coral reefs. He brought the nascent Museum of Comparative Zoology (MCZ) at Harvard to fruition and established the Newport Marine Zoological Laboratory. The latter was the first permanent marine research laboratory in the US. Here, I can do little more than highlight some aspects of Agassiz's life and career.

Alexander Agassiz, the only son of Louis Agassiz, was born in Neuchatel, Switzerland, on December 17, 1835. His mother, Cecile Braun, was an artist of exceptional ability from whom he acquired his own skill as an illustrator. By 1846, the elder Agassiz's finances had collapsed. Cecile, ill and exasperated by the conduct of some of Agassiz's collaborators, separated from him. She returned to Freiburg to be near her brother, Alexander Braun, the distinguished German botanist. Louis Agassiz also left Neuchatel and eventually made his way to the United States. In Freiburg, Alexander cared for his invalid mother and two sisters. He was introduced to botany by his uncle and to zoology by von Siebold. His mother died in 1848 after which he joined his father in the United States. Louis' marriage to Elizabeth Cary in 1850 restored stability to the Agassiz family.

Alexander was introduced to the family business of natural history and zoology as a boy. He accompanied his father on a cruise of the *Bibb* and aided in a survey of the Florida reefs. Graduated from Harvard in 1855, he took a degree in engineering from the Lawrence Scientific School. A position with the US Coast Survey enabled him to travel to the coast of North America by way of Panama. Research that he carried out on jellyfish and viviparous surfperches apparently settled him on a career in zoology. Returning home, he took a second degree in natural history from the Lawrence School. Although he received many honorary degrees, he did not have an earned doctorate.

In 1860, he joined the newly established MCZ as an "agent", supervising the Radiate Collection (cnidarians, echinoderms, etc.) and assuming charge of the business affairs of the MCZ. In the same year, Alexander married Anna Russell, a member of a prominent Boston family. Summers were spent at Nahant where a marine lab had been fitted out in the Agassiz cottage. His first major scientific paper, on viviparous

surperches, appeared in 1861 and was followed by over 250 more. Most of the early papers dealt with marine organisms, especially their development. Among the highlights of this period were the *Embryology of the Starfish* (1864 and 1877), a handsomely illustrated monograph that provided a definitive account of development from fertilization through post-metamorphic juveniles, and the *North American Acalephae* (1865), a study of the medusoid jellyfishes of the Atlantic and Pacific coasts illustrated with 360 figures that Agassiz drew from life. In 1866, he was elected to the National Academy of Sciences.

His own impecunious state, the financial debacles of his father, and the recurrent fiscal problems of the MCZ caused Alexander to seek his fortune in mining. During 1866-68 he became supervisor of the Calumet and Hecla copper mine in northern Michigan. Agassiz transformed a failing enterprise into a fabulously prosperous mine and secured his own fortune. He labored under incredible conditions on what was then a rough and tumble frontier. To recoup his health, impaired by the Calumet ordeal, he spent 1869 and 1870 in Europe. There he visited museums and made "the acquaintance of nearly every working naturalist in Great Britain, Scandinavia, Italy, Germany, and France", among them Wyville-Thomson, with whom he discussed the emerging field of deep sea research. Alexander's primary objective was to examine collections of echinoids. On his return, he published his massive (770 page) *Revision of Echini* (1872-1874).

The first session of the Penikese school was held in the summer of 1873. Alexander taught embryology. The following year, he organized the second and last session, but did not participate in it. Late in 1873, Louis Agassiz died, followed shortly thereafter by Alexander's wife, Anna, who had been nursing the elder Agassiz. Their deaths dealt him a shattering blow from which he never fully recovered. Although still capable of warm friendship and great enthusiasm, he became a very private person, projecting an austere public image. It is interesting to speculate on the extent to which Alexander blamed his deceased father for the death of his wife and, earlier, for his mother's death. Following this tragedy, Alexander sought refuge in his work. In 1875, he became curator of the MCZ and launched his first major expedition, traveling to Lake Titicaca and coastal Peru.

To digress briefly, the first marine "lab" in the United States was part of the Agassiz cottage at Nahant, Massachusetts. Here from about 1850-1870, the elder Agassiz, his collaborators, and Alexander studied invertebrate biology and development. Alexander's research and field observations led to the publication in 1865 of *Seaside Studies in Natural History* which he co-authored with his stepmother, Elizabeth Cary Agassiz. This book contains a wealth of information on the ecology, biology, and development of the intertidal and inshore fauna of northeastern North America. It is the first such American guide to coastal marine life. The history of marine labs moves from Nahant to Newport by way of Penikese. The Penikese school was a glorious experiment in instruction carried out under quite primitive conditions. Attempts to move the Penikese operation to Woods Hole failed. Because his memories of Nahant were too distressing, Alexander built a home at Newport, Rhode Island. He had a room fitted out as a laboratory in which W.K. Brooks carried out his study on *Salpa*. In 1877, Agassiz constructed a separate building which became the Newport Marine Laboratory. He did not intend to replace Penikese but rather to move in a different direction, providing facilities for research. The original research emphasis was on the development of fishes and marine invertebrates. After its enlargement in 1891, the emphasis shifted to deep sea and coral reef biology. The laboratory was superbly designed and appointed. Illustrated accounts appear in *Nature* 19: 371-319; *The Century Magazine* 26(5): 728-734; and *Annual Reports of the Museum of Comparative Zoology* for 1877-78 and 1891-92. In architecture and design, the Newport Lab was distinctly American and differed from European labs such as the Stazione Zoologica at Naples. It served as a physical and intellectual model for other American marine labs. It incorporated many features that are now common, such as: (1) a well-designed flowing sea water system with aeration; (2) access to a rich local fauna; (3) facilities and boats for both in-shore and off-shore collecting; (4) custom glassware and aquaria; (5) reference library; (6) experienced assistants; (7) facilities for photography; (8) good research microscopes; and (9) knowledge of the local fauna and practical experience in its study using techniques such as artificial fertilization and experimental hybridization. The Newport Lab was the premier American research lab from the time of its inception until the US Fish Commission Laboratory and the MBL attained maturity in the 1890's. Among those who did research at Newport were A. Agassiz, W.K. Brooks, C.B. Davenport, W. A. Faxon, J.W. Fewkes, E.L. Mark, A.G. Mayer, E.A. Nunn (Mrs.

C.O. Whitman), G.H. Parker, W.E. Ritter, C.O. Whitman, E.B. Wilson. Detailed information about the lab seems to be fragmentary and scattered, but it might prove interesting to try to draw it together.

Alexander Agassiz was considered one of the leading American embryologists in the period 1860-1885. In 1879, he was awarded the Prix Serres for his research on the development of cnidarians, ctenophores, annelids, echinoderms, enteropneusts, and fishes. He was the first non-Frenchman so honored. Alexander was part of what should be considered the Boston-Cambridge school of embryology. This group was established by the elder Agassiz, a student of Döllinger. Initially, it was comprised of students and associates, such as Alexander Agassiz, Clark, Morse, Packard, Wyman and, subsequently, their students and associates such as Brooks and Minot. They operated within the framework of the MCZ, Boston Society of Natural History, and local colleges and institutes. Alexander espoused an integrative, organismal approach to development within the conceptual framework of what is now known as the life history model. He helped establish a tradition of painstakingly detailed observation of living embryos, larvae, and post-metamorphic juveniles combined with state-of-the-art microscopy. This approach later was to become associated with Brooks, Whitman and their students and culminated in the Hopkins-MBL school.

Under Agassiz's direction and with his financial assistance, the MCZ became the first American center for embryological research and the hub of the Boston-Cambridge school. He established an embryological research agenda and was aided in this by E.L. Mark who joined the Harvard faculty in 1877. Mark, the first American student to obtain a doctorate from Rudolf Leuckart (1876), brought to bear a cytological and histological approach. Mark, together with Minot and Whitman, was responsible for the introduction of advanced European microscopic technique. Whitman, while working as Agassiz's research associate on teleost development, wrote his *Methods in Microscopical Anatomy and Embryology*, the first American work. To further embryological research, Agassiz (1) insured access to research material via the Newport lab; (2) acquired microscopes and apparatus needed for advanced microscopy; (3) assembled a research library; and (4) fostered an in-house publication series for well illustrated embryological monographs. To provide easy access to the scarce European literature, Agassiz, Mark, Faxon, and Fewkes published a series of selected illustrations from embryological monographs, i.e., "normal stages of development" with accompanying bibliographies. In addition, E.L. Mark translated several German reference works, notably, Korschelt and Heider's four-volume *Textbook of Embryology*. Unfortunately, Agassiz's vision of a center of excellence was thwarted by President Eliot's refusal to hire additional faculty, such as C.O. Whitman. Harvard's loss was Chicago's gain!

Agassiz's period of embryological research activities extended from about 1860 through 1890 during which he explored a number of topics, among which are the following:

- (1) Cnidarians: alternation of generations, embryonic and larval development, tentacle differentiation, morphological variation, and growth rate of corals.
- (2) Ctenophores: elegant comparative studies that accurately depicted unilateral (heart-shaped) cleavage and the segregation of the comb-forming cortical cytoplasm in micromeres by unequal cell division. He advocated splitting the ctenophores off from the cnidarians.
- (3) Annelids: Alternation of generations, embryonic development, and metamorphosis.
- (4) Echinoderms: an enormous body of work on echinoderm development that involved artificial fertilization and experimental hybridization. Description of starfish development from fertilization through post-metamorphic juveniles with elegant illustrations from living material. Viviparity and brooding. Homology of pedicellariae and simple spines. Post-metamorphic studies of growth and development carried out on a comparative basis.
- (5) Enteropneusts: He described the metamorphosis of the tornaria larva and identified it as a stage in the development of *Balanoglossus*.
- (6) Fishes: Cellular analysis of early teleost development (with C.O. Whitman), comparative studies of the development and metamorphosis of pelagic and demersal fish larvae, metamorphosis of flounder larvae and



different modes of eye migration to the dorsal surface, first observations on the development of the garpike, *Lepisosteus*, a primitive bony fish.

(7) Theoretical Studies: Agassiz criticized Haeckle's theories both in terms of factual evidence and its interpretation. Unlike his father, Alexander Agassiz accepted the evolutionary paradigm. In 1880, he presented a remarkable address to the annual AAAS meeting, entitled, "Paleontological and Embryological Development" in which he presaged the role of developmental constraints in evolution, questioned the constancy of evolutionary rates, and took the first steps toward a cladistic approach to taxonomy.

Having accomplished many of his goals in embryology, Agassiz moved on to deep sea biology and coral reef studies. Between 1877 and 1880, Agassiz directed the three cruises of the *Blake* in the Gulf Stream and West Indies. On these expeditions, he drew on his experience as a mining engineer to introduce wire cable as a replacement for rope, thus significantly improving sampling capabilities. Assessing the expedition, A.G. Mayer stated that, "due to the explorations of the *Blake* under Alexander Agassiz, we know more of the topography and the animals of the Gulf Stream and West Indian region than of any submarine area of equal extent in the world". In 1888, Agassiz published a unique two volume work entitled *Three Cruises of the Blake*. Not only did he provide an account of the scientific work of the expedition, but he also provided an overview of physical and biological oceanography, an analysis of the West Indian marine fauna, and a lengthy account of deep sea organisms. In actuality, it was the first oceanography textbook. In 1891, Agassiz undertook the first of three expeditions in the US Fish Commission's research vessel, *Albatross*. He explored vast areas of the Pacific. The first voyage traversed portions of the eastern Pacific. Many new deep-sea organisms were collected. Agassiz ascertained that the deep-sea fauna on the west coast of Central America had a close affinity to the Caribbean fauna confirming their continuity prior to the uplifting of the Isthmus of Panama. In 1899-1900, he took the *Albatross* from San Francisco to the Marquesas and across the tropical Pacific and its fabled islands and, thence, north to Japan. He found that the pelagic and bottom fauna in many areas was impoverished. On the third voyage in 1904, the *Albatross* returned to the eastern Pacific and conducted extensive studies on the rich pelagic fauna of the Humboldt Current and the correspondingly rich bottom fauna. Agassiz also explored the coral reefs of the Hawaiian Islands, Fiji, the Great Barrier Reef of Australia, the Maldives, Florida, and the Bahamas. He believed that Darwin's theory of coral reef formation was an oversimplification and that reefs could be formed in a variety of ways. Although he was working on a fourth version of a book to synthesize his years of coral reef research, he did not complete it before his death.

Agassiz resigned his curatorship at the MCZ in 1898 but retained control of its activities. His last years were interspersed with the *Albatross* and coral reef expeditions, the publication of expedition reports, and the study of echinoderms that were collected, e.g. "Panamic Deep Sea Echini" (1904). He died at sea while returning from Europe in 1910. With the deaths of W.K. Brooks and C.O. Whitman that same year, the era of 19th century American zoology was brought to a close. If you seek his monument, look not only to the men but also to the institutions. Singular tributes were paid to this "Prince of Oceanographers" by Henry Adams, A.G. Mayer, John Murray, and George Parker. As unofficial mentor and role model, Alexander Agassiz influenced many about him, amongst whom I would single out W.K. Brooks, H.P. Bigelow, A.G. Mayer, W. Ritter, T.W. Vaughn, and C.O. Whitman. Brooks founded the Chesapeake Lab and was mentor to Conklin, Harrison, Morgan and Wilson. Whitman nurtured the MBL through its early perilous years, created Chicago's center of excellence, and was mentor to the Lillies. Ritter founded Scripps and was succeeded by Vaughn. Mayer founded Carnegie's Tortugas Lab. Bigelow founded WHOI.

I acknowledge with thanks the comments and suggestions of Michael Ghiselin and Edward Ruppert.

#### Selected References

1. Agassiz, G.R. 1913. *Letters and Recollections of Alexander Agassiz*. Houghton Mifflin Co., Boston.
2. Goodale, G.L. 1912. Alexander Agassiz, 1835-1910. *Biogr. Mem. Nat. Acad. Sci. (US)* 7:291-305 (contains a bibliography of Agassiz's papers).
3. Mayer, A.G. 1911. Alexander Agassiz, 1835-1910. *Ann. Rept. Smithsonian Inst.* 1910, pp. 447-472, 1 plate. Also *Popular Science Monthly* 76(5) (Nov.).
4. Murray, J. 1911. Alexander Agassiz: his life and scientific work. *Bull. Mus. Comp. Zool.* 54(3):138-158.
5. Winsor, M.P. 1991. *Reading the Shape of Nature*. Univ. of Chicago Press, Chicago.



### How I began to study echinoderms... Part 3.

The section of the Echinoderm Newsletter containing personal accounts of how individuals began their studies on echinoderms has proved to be extremely interesting and popular. I have found them fascinating.

**Anderson, John M.** (Cornell University). The year was 1948. Returning to Academia from four and a half years' service as a naval officer in WWII, I had finished my graduate training in insect physiology under Prof. Daniel Ludwig at New York University and managed to produce an acceptable dissertation. Settling into an assistant professorship at Brown University as a fresh-caught PhD (by now with a wife and two small sons), I found myself for the next couple of years casting about for a stimulating field of research for the long haul. Histological and histochemical studies on the male reproductive system of the Japanese Beetle (the animal whose metamorphic biochemistry I had worked on as a graduate student, some intriguing problems in the reproduction of freshwater planarians, and even an investigation of the biological effects of ultrasonic radiation (on years, of all things!) occupied my questing attention. At one point my newly assigned research assistant at Brown (and I as well) needed training and experience in histological and histochemical techniques (the latter coming increasingly into vogue at the time), and in seeking not-too-difficult invertebrate practice materials we began to play around with such things as crayfish hepatopancreas and the pyloric caeca of sea-stars (then commonly referred to as starfish). Brown University is located in Providence, RI, not far from Narragansett Bay, and sufficient numbers of starfish could be had by special request from dragger operating out of nearby Bristol. Seeking to understand what we were seeing in our practice slides of the starfish material, it was natural to check into the existing literature - which revealed interesting gaps, inconsistencies, and obvious inaccuracies in previous descriptions. This, plus the fascinatingly different nature of the asteroid digestive system in general, led us to focus on the starfish sections to the exclusion of other materials - and thus began my career-long obsession with gross and microscopic details of digestive systems in various species of sea-stars.

In 1952 the offer of a tenured appointment in the Department of Zoology at Cornell University lured me away from the seashore, but summertime association with the Marine Biological Laboratory at Woods Hole, beginning in 1953, made it possible to continue studies on Asterias forbesi for the next several years. The shallow-water asteroid fauna of the region south of Cape Cod is relative depauperate, however, and it was not until 1958 that a sabbatic leave from Cornell, together with a Guggenheim fellowship, enabled me to spend several months at the Hopkins Marine Station at Pacific Grove, California, studying species of sea-stars I'd never seen before. This experience opened my eyes to the variety of ways in which different asteroid families had adapted, in a functional-anatomical sense, to different food habits and feeding methods.

Following up aspects of these diverse adaptations occupied my research interest during the remainder of my active professional career.

**Birkeland, Charles** (University of Guam). I have always been interested in animals, but as a boy I found terrestrial animals difficult to work with. They were either speedy, cryptic, at the tops of the trees, or hidden in the shrubbery. I enjoyed searching and getting glimpses, and frequently even good views. But they were frustrating for getting serious work accomplished. I thought birds, fishes, mammals and insects make good hobbies, but I would rather do serious work with large, slow animals.

In high school I had a summer job with the Illinois Natural History Survey, sampling forage-crop insects. We would take sweep samples in the open alfalfa fields, then dump several sweep-net loads into the cab of the pickup truck, hop in, close the door, and sample the galaxy of insects for particular species of economic interest as they spread themselves out on the windshield, attracted to the sunlight. In the Illinois summer, the temperature and humidity were usually about 100 (degrees F and %, respectively). As I worked in the hot, stuffy cab, searching and collecting selected species with an aspirator, the many insects would stick to me and drift into my eyes while they crawled in the sweat that poured down my face and neck. During those times, I thought that I would like to be a marine biologist, feel clean and never be hot and sticky.

Like many other marine biologists, I got imprinted on my first field trip in graduate school. On my first dive in Puget Sound, I encountered several species of starfish eating Ptilosarcus in a vast bed of sea pens. It was a spectacular and interesting sight. The starfish were large, surreal, bizarre, and beautiful, as were their prey. These echinoderms preying on anthozoans became the subject of my thesis right then, and for the last 22 years, it has been a major focus of my experiences on coral reefs. They have never become routine or boring.

**Conand, Chantal** (Universite de Britannia Occidentale). When I arrived from France and Senegal in New Caledonia in 1979, I wanted to continue studying fish biology. As in other tropical islands, the fishery in New Caledonia was considered at that time as not important enough to require scientific management. The New Caledonia economy was based on nickel and due to the nickel crisis the authorities wanted to diversify the sources of income and the artisanal fisheries. One of them, the beche-de-mer, was said to have been prosperous once but there was no information! I was asked by ORSTOM (Institut Francais de Recherches pour le Developpement en Cooperation) to undertake studies to determine which holothurian species are valuable, where they live, how they grow, reproduce, die, how much could be fished... I started sampling on reef flats and diving in the lagoons with admiration. It has been the start of my association with these wonderful creatures, although mostly depreciated by "non connaisseurs"! I soon met my new echinoderm colleagues at the Seminaire organized in Paris by Alain Guille and they became friends. (John, my English is

too poor to make a joke with nickel - "mining" - and nickel - "money" - with beche-de-mer!)

Farmanfarmaian, Allahverdi Abdul-Hossieh (Rutgers University). Toward the end of WW II, my guardian put me on a train in Teheran and, along with several of my brothers, sent me off to the American University of Beirut (AUB) Prep School (IC) "...so that I might become civilized and educated and leave my saintly mother to have some peace", his words. I was 13 and had never seen an echinoderm though I had collected shells on the beautiful shores of the Caspian Sea. Zenkevitch does not report any echinoderms in the Caspian (average salinity 12-13 ppt). To save fuel, the IC boarding masters dictated cold showers from April to October, and to make sure we "savages" took our showers, they herded us off to the beautiful AUB plage to swim in the Mediterranean. Mr. Damous, our athletics director, kept order with his piercing police whistle and giant torso. In spite of his severe injunctions and my bad right ear drum, I could not wait to explore the sea bottom. I had no diving implements other than my brief Bikini - the recyclable standard issue of the IC athletic department. It was there on an algae-covered rock, about 14 feet below, that I saw a purplish greenish spiny fuzz-ball - larger than a golfball - moving on that rock. I took a breath and returned to the same rock, and the creature was there and moving. I looked and found no head, no eyes, no antennae, no legs, arms, fins or claws. A little afraid, but mostly curious, I went to my brothers who were attending Mr. Damous' high diving lessons near the diving board. They would not believe my story and laughed at the idea of the moving creature without a head, eyes, legs, and we almost came to blows when my brother Tari said, "Oh, yeh, we have several of those in our class and one is named Verdi". Mr. Damous broke up the fight and ordered us to the showers. He told us to get dressed and report to him just outside the University gates where the vendor Hatab sold different kinds of foods and sandwiches to students. We dutifully turned up at the appointed time and place. Mr. Damous said something in Lebanese Arabic to Hatab. We Persians did not understand. Hatab went behind his cart and came back with a bucket of chilled sea-water, a plate, a loaf of French bread, a couple of fresh lemons and a peculiar knife. He then reached into the bucket and pulled out my creature - the Mediterranean Echinus of Aristotelian fame. I started to scream with laughter at my brothers who were looking for cracks to crawl into. Hatab deftly cut the creature at the ambitus into clean halves and placed it on the plate. Mr. Damous showed us how to pick the yellowish gonads with a small fork, place it on a piece of bread, squeeze a few drops of lemon juice and eat it. Tari did not like it but the rest of us enjoyed several of these sea urchins. Mr. Damous paid Hatab 80 Lebanese piasters and dismissed us in his usual military manner. For years thereafter I looked for these creatures without heads, tails, right or left, front or back on shores, in museums, and in exhibitions with great curiosity. As a sophomore in my first invertebrate course, and later as a graduate student in D.P. Abbott's Marine Invertebrates Course and A.C. Giese's Comparative and Ecological Physiology Course at Stanford's Hopkins Marine

Station, I studied a variety of echinoderms. Scuba diving around Monterey, Catalina, Florida, Cayman, Woods Hole and Palau, I observed and collected many echinoderms and never ceased to marvel at their beauty and behavior. I did research on more than 30 species, representing all five living classes and ranging from the depth of the Monterey Canyon to the shores of Cape Cod, the sea tables of Naples, and the rocky beaches of Hormuz in the Persian Gulf. I read Cuenot and Hyman cover to cover twice and in part several times and consulted tons of other literature in several languages including German and Japanese. I published and presented many papers on the respiratory, circulatory, digestive, absorptive temperature tolerance, osmoregulatory, and reproductive physiology of echinoderms. For several years I taught a highly specialized course in echinoderm biology at Woods Hole. Finally, though I have worked on other animals, I have never stopped puzzling about echinoderms for the last 50 years (13-63). The puzzle never ends. I admire all who continue the struggle to unravel these great secrets.

**Fenau, Lucienne** (Station Zoologique, Villefranche-sur-mer). After having obtained my Licence en Sciences Naturelles (Academie de Paris, 1958), I had two possibilities: commencing a third cycle of biological oceanography with the obligation of returning to my country Haiti after finishing my studies, or having a post as technician in a laboratory of plant physiology at the University of Paris. I liked the second possibility better as my future advisor, Dr. Camus, gave me the opportunity of beginning a doctoral program under his direction. Then I went to Villefranche to see Robert Fenau, my future husband, who introduced me to Mr. Roger Lallier, a scientist with CNRS. Mr. Lallier spoke to me of his studies on embryonic determination and showed me the effects of lithium and zinc on the embryonic development of urchins. This was a blow to the heart! The material was fantastic: it was easy to observe under the stereomicroscope and the changes induced by lithium and zinc were spectacular. Now I knew that I wanted to work on echinoderm larvae but how to choose in a field so vast? Professeur Paul Bougis, director of the Station Zoologique of Villefranche-sur-mer, received me and spoke to me about larval nutrition that was still poorly known, of the experiments that could be done on larval growth raised with particles encountered naturally in the marine environment. This time my research project took form. But it was first necessary to know well larval ecology in situ, the gametogenic cycle of adults, the different changes in form the larvae took during the course of gametogenesis. The adults I could collect at Villefranche still had an unknown larval development. The literature of Muller, Mortensen, Giese and his collaborators, and those of other authors revealed by reading the book edited by Boolootian "Physiology of Echinodermata" became familiar to me... And this is how I presented a thesis titled "Aspects ecologiques de la reproduction des Echinides et Ophiurides de Villefranche-sur-Mer". Later I discovered that Richard Strathmann had the same passion...

**Ferguson, John Caruthers** (Eckerd College). Before deciding to be

an echinoderm biologist, I had to elect not to follow my parents into medicine and biomedical research. This departure occurred as an undergraduate at Duke, after reading N.J. Berrill's The Living Tide, with its description of marine life in the Dry Tortugas. A life studying sea animals pictured as much more rewarding than one emersed in human blood and guts. Knut Schmidt-Nielsen and his Comparative Physiology course further fascinated me with the beautiful diversity of animal functions. While enquiring into graduate schools to pursue this new interest, John Anderson intercepted my letter and invited me to focus my attention on invertebrates with him at Cornell. It as a big step to give up animals with backbones, but there was a lot to discover in these "other" animals, and they could be a good excuse to spend a lot of time at marine labs. Arriving at Cornell, I was surprised to find John Anderson off on sabbatical, leaving a note telling me, in effect, to choose an animal and get to work on something - anything! Thus, I had complete freedom on choose any animal group for my career. After considering all the alternatives, the answer was clear: starfish (and echinoderms were weird, puzzling, casually fascinating, and definitely marine - what else could be more interesting! Further, since Anderson also worked on them, he could provide me unique support and guidance. When Anderson finally did return, I found in him a true mentor who never farmed his work out on me, but provided a free, intellectual environment that fully encouraged my own creativity, subject only to the rigors of his constructive criticism. Being fortunate to find employment on a sea-front campus in Florida, I have been able to investigate through the years a number of basic properties of echinoderm biology. Each January I lead a sailing and study expedition with students to the Dry Tortugas, and try to pass on to them some of the spirit I inherited from Berrill (whom I have never met), Schmidt-Nielsen, and John Anderson.

Fischelson, Lev (Tel Aviv University). It was just at the beginning of my studies in the Gulf of Aqaba in 1951 when I became fascinated by the changeover from day-active to nocturnal animals. The most impressive was the appearance of the crinoids Lamprometra klunzingeri and Capillaster multiradiata that at this time covered all the coral forereef like a long-haired carpet. To observe this we used a normal flashlight inside a glass jar. This was before underwater lights became common.

The obstacle to overcome in observing crinoids was that one had to "fight" one's way through and between the most frightening long spines of the common Diadema setosum. Getting stung here and there on the stomach was worth it, however, and so one night I went out without a wetsuit, manoeuvring between the hostile spines. And there it was, on one site, that I lost my balance and fell over on my behind, landing of course on dozens of spines. It was very painful and I got angry and decided to crush this Diadema. Turning around, I pulled it out of me and discovered that it was not a Diadema, despite its black appearance. This was my first encounter with Echnothrix calarmaris, then a rare animal indeed on the Eilat reefs. From this time on I decided to study echinoderms and the rest is written history.

Holland, Nicholas Drew (Biological Oceanography, Scripps Institution of Oceanography, La Jolla). Growing up in southern Florida, I spent much of my time combing beaches for sea shells. I even took my specimens to the public library to match them up with the pictures in shell books to learn the scientific names. Although I was not adverse to including dried remains of starfishes and sea urchins in my collection, I was definitely not an echinoderm biologist in those years. Even so, whenever a grown-up would inquire as to my plans for a future career, I would always tell them I would be a marine biologist. Neither they nor I really knew what marine biologists really did for a living, but my answer never failed to satisfy everyone concerned. After leaving all oceans far behind, I majored in biology at a very small college in Minnesota. There, I stayed interested in the invertebrates in spite of a lean offering of living material (hydras, planarians, earthworms, and crayfishes). My differentiation into an echinoderm biologist took place at the MBL at Woods Hole in the summer of 1960 between my undergraduate and graduate school. I took the MBL invertebrate course (which long ago vanished beneath the rising tide of reductionism), which was taught by a series of instructors, each entrusted with a phylum or two. As a result, the quality of the teaching fluctuated wildly from the ridiculous to the sublime. Certainly, the best lecture series was given by Verdi Farmanfarmanian, who covered the echinoderms. That lecture series played a big part in bringing me into the echinoderm fold-- but it was not quite the whole story. The critical moment occurred in the middle of Verdi's performance, when the student next to me blurted out: "This sure is interesting, it's too bad you can't make a living doing it." I looked her in the eye (she's a college administrator now, and a very good one and I said: "We'll see about that!") The rest is history.

Jangoux, Michel (Universite Libre de Bruxelles). Until the age of 18 I was not particularly interested in biology, nor anything else except reading novels of all kinds, walks in the woods, and going out with friends. I registered in the school of medicine at the University of Brussels...as that pleased my parents, especially my mother. One of the required courses was zoology. The course was remarkably taught by a true zoologist (a specialist in fresh-water sponges) and with such enthusiasm that I decided to abandon the medical direction and registered in the section of biology of the faculty of science. In reality, all aspects of fundamental biology interested me and when after three years it was necessary to choose between botany, zoology, and molecular biology, it was a true rending. If I finally chose zoology, it was because it was the only department where research could be done in the marine environment. It was thus quite natural that, a year later, I went to the director of the laboratory of marine biology, Prof. Jean Bouillon, a specialist of hydrozoans, to ask him to accept me as a master's student. He very cordially welcomed me and asked me which group of animals I wished to study. I was not at all prepared for this question, being convinced that he was going to propose a study on hydropolyps or hydromedusae to me! He explained to me that he found it much more rewarding to have a student work on subjects of

which he was not a specialist. He then gave me two books (one of Monton on molluscs, and the other edited by Boolootian on echinoderms), and asked me to read them and return to see him two months later...this was in August 1968. The echinoderms pleased me more than the molluscs, both for aesthetic reasons and as I found them very mysterious. I was especially interested in reading the chapter by J.M. Anderson ("Aspects of nutritional physiology") and very intrigued by a small paragraph reporting the existence, in starfish, of small digestive organs of unknown function: the rectal caeca. I thus proposed to Prof. Bouillon to make this the subject of my master's thesis...and it was thus my researches on echinoderms began.

Johnson, Craig (University of Queensland, Brisbane). My story is simple. I had good lecturers as an undergraduate at the University of Tasmania so from the beginning found echinoderms both interesting and aesthetic creatures. I was further motivated by Ken Mann when he visited the University on a lecture tour as a Senior Queens Fellow in my third year as an undergraduate. Ken gave a tremendous talk about the Nova Scotia "urchin situation". I was fascinated to hear of the outbreaks and destruction of kelp, and of the best pieces of Breen's, Fong's, Guerinot's and others of his students' work. Some good slides certainly helped stimulate my interest. Subsequently I wrote to Ken about the possibility of pursuing a PhD under his supervision in Halifax, and the rest, as they say, is history.

Levitan, Donald (University of California, Davis). In the summer of 1983 I went to the U.S. Virgin Islands to teach a course, and start my dissertation research on overgrowth interactions between encrusting taxa. When I got to the shore, I quickly realized that entering the water was impossible; from the intertidal down to the sand halo, the entire reef was covered spine tip to spine tip with the poisonous sea urchin Diadema antillarum. In fact, from a distance, the reef had a black hue caused by the huge numbers of Diadema. When I finally managed to get into the water (by jumping off a dock), I noticed that the surface the sea urchins were feeding on were grazed clean. I began to wonder how individuals could survive and how populations could persist under these apparently food-limited conditions. This changed the direction of my research and started my interest in echinoderm biology.

Manchenko, Gennady (Institute of Marine Biology, Vladivostock). I am not an echinoderm biologist in the strict sense of the term. However my entering biological science was tightly connected with echinoderms. When I was still a school boy I, as many children in the former Soviet Union, dreamt to become a military pilot. In 1964 my dream almost became true after my entering the military aviation high school. However after only one year of studies at the school I drastically changed my mind and left the school. This mistake in my life-strategy cost me four years of life which I had to spend surviving in the Soviet Army. Nevertheless, I am very thankful to this period of my life as it stimulated me to think much more deeply about my being. As a result, I firmly decided to

enter the Division of Natural Sciences of the State University of Novosibirsk. I entered Novosibirsk University in 1968 and was graduated in 1973, majoring in "cytology and genetics". About one month before graduation from the University, I met Dr. Alexey V. Zirmunsky, the Director of the Institute of Marine Biology at Vladivostok. He invited me to work at the Genetics Laboratory headed by Alexander Pudovkin. The Institute was very young (it was founded in 1971) and no zoologists were very familiar with the great variety of invertebrate species inhabiting the Sea of Japan. Echinoderms and molluscs were perhaps the only invertebrate groups represented by species with large, attractive, and well-diagnosed individuals inhabiting the sublittoral zone and thus easily available for collection and identification. Fortunately, the most beautiful sea animals, the sea stars, were almost completely unstudied at that time in respect to the level of intraspecific genetic variability. So I began to study allozymic variation in sea stars using the enzyme electrophoresis technique. Since that time I studied isozymes in sea stars, sea urchins, sea cucumbers, and sea lilies. In 1981 I was invited by John Lawrence to take part in the International Echinoderm Conference at Tampa Bay and to prepare a report on genetic variation in echinoderms. However, my attempts to visit Tampa failed because of the well-known difficulties concerning travel from the U.S.S.R. at that time. My report was also not submitted to the Conference because of some unexplained delay of Soviet authorities in supplying me with the necessary permission to send the manuscript abroad. Although I am rather an isozymologist than an echinoderm biologist, I love echinoderms as they are beautiful and studying them is associated with a substantial part of my scientific life. I believe my love is not yet finished.

**McNamara, K.J.** (Palaeontology, Western Australian Museum, Perth): The lure of fossil echinoids has been a pervasive influence on my palaeontological career, even from the earliest days when I first started collecting fossils. Having been brought up on the chalk downland in Sussex in England, it is not surprising that when I first got bitten by the fossil bug when I was 9 years old that echinoids should have been one of my sought after treasures. While many Museum fossil collections in England would seem to indicate that the English Chalk is riddled with echinoids, this was certainly not the case in Sussex, and I spent many hours searching for those elusive urchins. My university training took me away from fossil echinoids, and following 4 years in the Precambrian at Aberdeen University my PhD at Cambridge was on Ordovician trilobites.

My first employment after completing my doctorate was at the University of Queensland in Australia. There I dabbled in trilobites and ammonites. After two years the job folded, and it was a case of returning to the UK or finding another in Australia. Scanning the local newspapers I found a 3 or 4 line advert that was to affect my future career quite dramatically. Graeme Philip, then Edgeworth David Professor at the University of Sydney, wanted a research assistant to work on Tertiary irregular echinoids with



him. Not being able to resist the lure of echinoids, I applied and got the job. As my research background had precluded any studies of echinoids I asked and got the job. As my research background had precluded any studies of echinoids, I asked Graeme what papers I should read. "Don't you dare read anything", he threatened. "I don't want you to be influenced by what other people have said. Work it all out for yourself!" This, curiously, turned out to be really quite good advice and perhaps was one of the reasons why I was able to combine my interest in heterochrony with the echinoid studies. Southern Australia has a magnificent Tertiary echinoid fauna, much of which had been hardly worked on. Not only did I have access to magnificent collections made by Bob Foster and his mother, but I was able to draw on many of the untouched collections in the Museums of Victoria and South Australia.

Not longer after starting in Sydney the job as Curator at the Western Australian Museum was advertised. With some reluctance I applied, having just got into the echinoid studies with Graeme, but when offered the job I could hardly refuse. But this opened up the even more untapped echinoid treasures of Western Australia and a lifetime's work.

Mooi, Rich (California Academy of Sciences). I was fortunate in being born to two extraordinary parents who fostered my interest in the natural world from a very early age. This was reinforced by a synergy with my younger brother. I remember sitting in the forests of local conservation areas around Toronto, Canada, drawing plants and animals from life, and spending hours with identification guides, learning the rudiments of evolutionary biology and systematics. I also recall watching every nature program that we could get on TV, and through this window on the planet, I discovered an inner drive to work in the ocean. At the age of 10, I decided that marine biology would be my calling, and almost every move I made in subsequent years was aimed at fulfilling this goal, in spite of my landlocked situation in the middle of Ontario. Camping trips to both coasts of Canada enhanced my desire to study marine organisms, and I practically lived in the pages of Rachel Carson's books.

In my first year at the University of Toronto, I had the audacity to think that I could just march into the office of the campus' resident marine biologist, and demand information and ideas for how to pursue my calling. Luckily, the man occupying the office was Malcolm Telford, and he indulged my brash enthusiasm to the point where my energies could be focused on a library project that searched databases for papers on deep-sea ecology. Little did I know that Malcolm was to become not only the most important academic influence on my career, but one of my closest friends as well. Naturally I took his marine biology field course, and the invertebrate zoology course, too. I really enjoyed all the stuff about worms, molluscs, and crustaceans, but it wasn't until he introduced the echinoderms as a weird bunch of mutated space garbage left over from an extraterrestrial's picnic that I had my niche. By this time, my early experience with drawing had developed into a real interest in wildlife painting and technical illustration, and it seemed as though the Echinodermata were the

perfect blend of artistic, aesthetic, and scientific wonder. I became a graduate student of Malcolm's who insisted that I was not working for him, but with him, an important distinction that emphasized to me the value of collaboration.

My master's degree dealt with tube-foot morphology of sand dollars, and I soon realized that as good as functional morphology was as a topic, it seemed to obtain much greater value as a comparative study. Which led me to phylogenetics, which in turn led me to Rick Winterbottom, a Royal Ontario Museum ichthyologist who introduced me to cladistics, and made me realize that phylogenetic analysis should be the basis of all comparative biology. My doctoral thesis put this idea to work, using sand dollars as a model for some of my wilder thoughts about miniaturization, and heterochrony within the Echinoidea. I obtained a post-doc with Dave Pawson at the National Museum in Washington, where I was fortunate in having access to the famous collections, as well as a conduit to collaborations with many international figures in echinology. My interests have consequently expanded to include deep-sea echinoids, and the evolution of the phylum as a whole - projects that I am fostering with Bruno David in Dijon, France. I have tried as best I can to maintain these contacts and am now pleased to have the opportunity to do so from a position at the California Academy. They seem happy to suffer my eccentricities for echinoderms, and take delight in pointing out to me that the value of the echinoid type of dollar is higher here than in Canada. Dendraster does seem to be larger in California.

**Pentreath, Victor W.** (University of Salford). A required component of my first degree in Zoology at St. Andrews University, Scotland, was the completion of a research project. My supervisor, Glen Cottrell in the Gatty Marine Laboratory, who was studying neurotransmitters in a range of invertebrate nervous systems, suggested I investigate the cholinergic system in starfish nerve. This proved an immediate fruitful source of the cholinergic triad, thus allowing me to extend the early observations of Bacq, and also to make some functional interpretations. However, the main growing awareness for me then was how little was known about the echinoderm nervous system and how few biologists had made a serious study of it.

Cold but happy hours spent on the windy Scottish shoreline, collecting various Asterias, Ophiothrix and Antedon with my friend and colleague Jim Cobb, also at the Gatty Marine Laboratory, led to a series of valuable collaborative investigations. Although it is now almost impossible to obtain funding for studies of echinoderm neurobiology, my deep interest remains.

**Propp, Michael** (Institute of Marine Biology, Vladivostock). I remember my first encounter with sea urchins perfectly well. From childhood I was fascinated by books of Jules Verne and about the sea, and dreamed myself as an explorer of the Ocean. So, when SCUBA appeared in the fifties, it was my chance. But Leningrad where I began to dive in 1957, is on the practically fresh Baltic

and Black Seas and nearly devoid of echinoderms. But in August 1959 I stood on the rocky shore of the Barents Sea in very cumbersome, home-made SCUBA ready to become the first man to dive in the Russian part of this sea. I stepped into the water - and this was my first contact with the wonder of Strongylocentrotus droebachiensis - as all the needles pricked my heel followed by trickles of ice-cold water. I slipped and bumped my bottom on the rock and - O! O! O! -the urchins were everywhere. I grabbed stones to regain my balance -and they were all covered by these wonderful animals. So a little later, having obtained the proper skill to extract crushed needles from my own skin, I simply could not make them the main point of my Ph.D.

Later I chose to supervise the research of my disciples and many urchins were sacrificed to measure respiration rates, dry weights and many other things. So I had my vengeance, but in due turn the urchins had theirs when I advised my last disciple to begin work with Diadema - he fled to Sebastopol on the Black Sea where he was unable to find any living echinoderms. This, unfortunately, happened to be the end for me of researching these fascinating animals.

**Scheibling, Robert** (McGill University). I suppose I became interested in echinoderms in a rather round-about way. When I enrolled in graduate school at McGill University in 1973, I was undecided about whether I wanted to study marine biology or animal behaviour. My supervisor, Carol Lalli was a malacologist and I thought I could indulge both of my interests, as well as my desire to work in some exotic place by undertaking a field study of predatory behaviour and territoriality in tropical octopuses. Somewhere in the Caribbean would be perfect, and the fact that McGill had a research station in Barbados fit well with my plan. So I diligently devoted myself to learning all I could about octopuses, and went to Barbados that first winter to begin some pilot studies. As it turned out, Barbados was no place to study octopuses (only because there were very few of them about), although I knew the Caribbean was right for me, maan! While on the rebound to find a new tropical animal to study, another student of Dr. Lalli's suggested I work on Oreaster reticulatus. He told me these sea stars occurred in large numbers in the Grenadines, where he had observed them feeding on urchins in seagrass beds. At the time, I was taken by Bob Paine's work on sea stars as keystone predators, and I thought I might be able to find a tropical analogue in Oreaster. In any event, I was interested in predator-prey interactions, and even if sea stars were brainless, it was still the Caribbean. When I searched the literature on Oreaster, I found that virtually nothing was known about its biology and ecology. It appeared that, while this species used to be widespread throughout the Caribbean, it had long been collected as a curio due to its handsome appearance and was now rare among the more populated islands like Barbados. I happened to see my first specimen in a shop window in downtown Montreal. It was part of a clothing display but I convinced the shop owner to sell it to me, and so the love affair began. The next thing I knew, I was on Carriacou, a remote desert island in the Grenadines, where I set out to learn what I could about this curious beast, armed only with a diving mask, snorkel and fins, and the companionship of a good woman. What more could one ask for? I suppose I learned more about life and about myself on that isolated and wondrous island than I did about Oreaster, but that experience paved the way to further adventures and discoveries, and ultimately decided my career. By the way, I soon found that the shallow lagoons off Carriacou were full of octopuses that no one had ever studied, but once I had stars in my eyes I never looked back. Those years in the Caribbean, studying the life of a sea star, were among the best years of my life.

**Sibuet, Myriam** (Institut francais de recherche pour l'exploitation de la mer, Brest). I was struck, during my first voyage in 1969 for three months aboard the ship Jean Charcot, by the abundance of echinoderms in the trawls and especially their unusual, nearly elegant forms. It was of course the genus Deima that attracted me! A long time the photo of this species was at my door and the figure of Herrouard in 1902 illustrated the first page of my state thesis! This typically marine group has seemed to me from the beginning of my research on the abyssal biology a group well adapted to the conditions of extreme life in the depths of the abyss. The

morphological characters are very peculiar and curious in certain cases and answer to a form of adaptation. I have sought to interpret the mode of life and the adaptation to the great depths in considering the echinoderms and especially the holothuroids as a model zoological group. Later the studies made in collaboration on the nutrition of holothuroids have led me again to interpret the detritivore system in using the holothuroid species as a model.

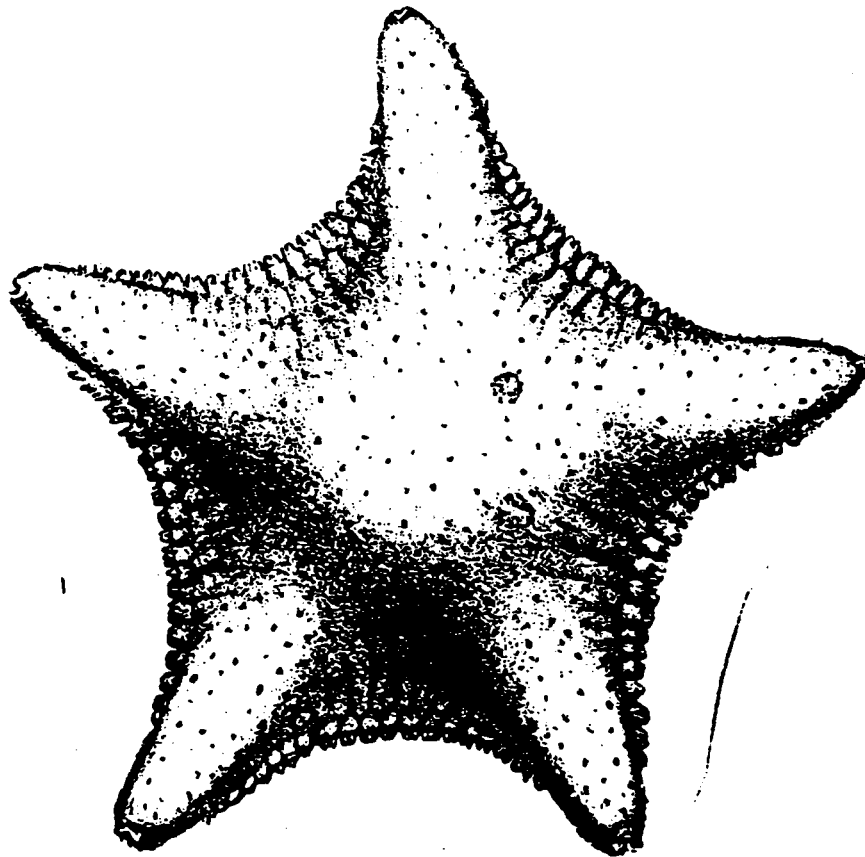
Woodley, Jeremy (Discovery Bay Marine Laboratory). When I was an undergraduate in the Zoology Department in Oxford, I attended a field course at the Millport marine Station. It was in the Easter vacation of, I think, 1959 and was attended by students from several other universities. We learned a lot and had a good time! Our visit overlapped with April Fool's Day when, I remember, the front gates of all the houses around Kames Bay were mysteriously interchanged. It was then that I learned how to bolt a door from the outside (on the inside) with a piece of string. One of my colleagues, excluded thus from his bedroom, borrowed. I regret to report that, when he took the ladder back, I was secretly watching and slipped in and did it again!

Echinoderms? Oh, yes: we each had to choose a group of organisms to sort from communal collections, identify and display to the others. I chose brittle-stars because I thought they were pretty. Also something of a challenge because some of them looked rather similar. So it was that I first met those fascinating creatures, the burrowing amphiurids, dredged from the glutinous mud of the Firth of Clyde.

Later in my undergraduate career, I had a term of tutorials in vertebrate biology from David Nichols. Not surprisingly, we talked about echinoderms quite often, and he suggested that I stay on after graduation to work with him. I had been much influenced by Arthur Cain and the school of evolutionary ecology at Oxford, and my first thought was to find out more about the adaptive radiation of British brittle-stars. But that meant, not only finding out what they do, but how they do it, and one day David asked "How do brittle-star tube-feet work, anyway?" We soon realized, not only that no-one knew the answer, but that the question had never really been asked. I resolved to try and answer it. Even before I graduated, I was back at Millport watching tube-feet and preserving brittle-stars for histology.

Even so, my career could have been in quite another field. I went on a University Expedition to British Guiana (as it then was), studying frogs and lizards in the rain forest of the Potaro River. I enjoyed working in the tropics, became fascinated by frogs, and actually secured a Brazilian Government Scholarship to continue working with them, though it would have been for only one year. Had it been possible, in those distant days, to get support for a postgraduate project with fieldwork overseas, I would now be a herpetologist! Funding was available to support projects at home, and I remember saying to myself that I would just knock off this little project on brittle-star tube-feet, then go back to my real love, the study of tropical frogs. People do change, don't they?! Nonetheless, when I took my first real job, I moved to the

University of the West Indies in Jamaica because it offered research opportunities, not only on marine invertebrates, but on tropical frogs! As it turned out, they remained a hobby, and I moved deeper and deeper into the sea.



Bell

**OBITUARIES**

Smith, James Eric. (an obituary for Dr. Smith by David Nichols appeared in the 1992 Echinoderm Newsletter. An extensive one by Q. Bone and D. Nichols appeared in Biographical Memoires of the Royal Society. 38, 325-343, 1992. The section on his echinoderm work follows)



*J.E. Smith*

### HONOURS

Eric was awarded a C.B.E. in 1972 and was knighted in 1977; among other honours, he received the gold medal of the Linnean Society in 1971, the Frink medal of the Zoological Society in 1982, and was elected a Fellow of King's College (1964) and Queen Mary College (1967). He received an honorary D.Sc. from Exeter (1968), and was one of the first Fellows of the then Plymouth Polytechnic (now the University of Plymouth).

### SCIENTIFIC WORK

Smith began his scientific life as a Student Probationer at the Laboratory of the Marine Biological Association at Plymouth. His first paper (1)\* in 1932 was the result of this two-year studentship, and described the physical nature and fauna of the Eddystone Shell-Gravels. He concluded that 'The distribution of species over the whole area is such as to suggest that any one of the three lamellibranchs, *Glycymeris glycymeris*, *Chione* (= *Venus*) *fasciata*, and *Astarte triangularis*, associated with *Spatangus purpureus* - which, although few in numbers, is characteristic of the ground - is sufficient to characterise the community'. Interestingly, he noted a significant decline in numbers of the small bivalve *Gouldia minima* since a previous survey ten years before, and Holme's later extensive surveys of the Channel benthos showed that this species has declined even more steeply since Smith's work (Holme, 1946). During his work on the shell-gravel, Smith started to collect embryonic stages of the nemertine worm *Cephalothrix rufifrons*. When he moved to an Assistant Lectureship at the University of Manchester, he filled in the missing developmental stages on return visits to Plymouth. His paper (2) describes the early development, of interest because the worm is a representative of the group of nemertines which exhibit direct development.

Smith's echinoderm work began with a short letter to *Nature* (5) on the occurrence of immature individuals within the genital bursae of *Ophiothrix fragilis* anticipating his 1940 paper (6) on the reproductive system of this brittle-star, a diversion from his principal work at that time. Smith notes the occurrence of young in the genital bursae of males as well as females. The habit is almost certainly a chance entering of the bursa by a young individual seeking a small crevice for safety during a vulnerable stage of the life-cycle: like spine-clinging, it does not denote vivipary. He was later to comment (6) that the term 'genital bursa' is unfortunate, as in so many ophiuroids, including *O. fragilis*, the oviducts do not open into these sacs, and their use as havens for the young is far from universal.

In this later paper (6) Smith describes for the first time, the gross stages in gametogenesis. As there is no evidence of proliferation of germ-cells from the gonadial epithelium itself, Smith concludes that the primary germ-cells present in the gonads have migrated there from the rachis: 'The gonad is not the place of origin of the gametocytes, but within it maturation and growth of germ-cells takes place.'

\* Numbers in this form refer to entries in the bibliography at the end of the text.



### *The neurobiology of asteroids*

While at Manchester, Smith embarked on the monumental work for which he became principally known: the neurobiology of the starfish. When he began, knowledge of the echinoderm nervous system was rudimentary. It was known that the neurons were very small, that spines, pedicellariae and tube feet could function autonomously, and that there was a motor system innervating effector organs. There was, however, still little unanimity of view regarding the subdivisions of the nervous system, and connections of neurons within it were largely unknown. It was in this historical context that Smith began the daunting task of investigating the nervous system of a group of animals that did not lend themselves readily to specific nerve stains, and whose nerves were difficult to identify (or even to see) by routine methods. He said later in life that his colleague at Cambridge, Carl Pantin, had tried in vain to warn him off the echinoderms, reminding him that their cells were far smaller than those of most other animals; that nobody could get very far with the neuroanatomy of animals so generously endowed with endoskeleton; and that these animals did not lend themselves to specific nerve stains. A later worker in echinoderm neurobiology has commented that 'There is less known about the organization of the echinoderm nervous system than that of any other phyla of metazoan animals. The reason for this is not that they are obscure or that they are not intriguing. It is simply that they are very difficult to work with technically (Cobb 1987). Electron microscopical techniques, which were later to enable great strides to be made in neural mapping, particularly at nerve-muscle junctions, were not yet available. Recently, too, new staining techniques, such as Lucifer yellow introduced iontophoretically into neurons, have enabled pathways to be traced with greater accuracy. But even without these later techniques Smith's work took the subject of echinoderm neurobiology forward with an impressive set of papers.

In the first of a series of three major papers in the *Philosophical Transactions* (3), Smith began to extend understanding of the nervous system by painstaking sectioning in several planes, followed by observations at the limits of optical microscopy. In this 1936 paper, in which his preparations were stained by conventional histological techniques, particularly Mallory's trichrome, Smith made enormous advances in understanding the neural basis for starfish activity. The paper has been extensively quoted subsequently, and no fewer than ten of Smith's figures from it were reproduced by Hyman in the echinoderm volume of her classic series *The Invertebrata*, a book that begins with the now-famous affirmation: 'I here salute the echinoderms as a noble group especially designed to puzzle the zoologist.'

The second paper (8) on the mechanics and innervation of the starfish tube-foot-ampulla system greatly advanced the study of the neural pathways involved in locomotion and other activities. As Smith remarked in his introduction, 'The tube feet of a starfish are concerned in almost every action which the animal as a whole is called upon to perform. It is, moreover, the *coordinated* movement of the podia which gives direction, purpose and rhythm to the action.'

To map neurons and their axons for this paper, Smith used for the first time the Unna-MacConnell methylene-blue leucobase technique for *intravital* staining of nervous tissue in fairly large chunks of starfish arm, subsequently fixed in ammonium molybdate and thick-sectioned. This variant of Ehrlich's technique had been used successfully in

demonstrating the nerve net of the coelenterate *Hydra* about ten years previously, and although it was to prove invaluable in tracing some nervous pathways in the starfish, it was also to be instrumental in misleading him in the structure of certain elements.

In his third paper on starfish neurobiology, published in 1950 (10), Smith describes in detail the innervation of the arm, ampullar and tube-foot musculature in *Astropecten*. He states that the axons innervating the tube-foot musculature 'may best be likened to ribbons many times broader than the thread-like fibres of the nerve chain. They are applied along the length of the muscles, many fibres of which are served by collateral branches of the ribbon axons.' A principal difficulty in this work was the accurate identification of the form of nerve-fibre terminals, as Smith recognized, and it later became clear that the 'ribbon axons' are in fact modified muscle cells. Dr J.L.S. Cobb writes: 'Smith produced many useful images (by his methylene-blue staining) but regrettably the advent of the electron microscope showed: (a) that muscle cells had also stained; and, (b) that long processes arise from muscles with a striking convergent similarity to axons.'

#### *Coordination of movement in the starfish*

Smith's first review (7) was written during the highly productive period of laboratory work he undertook at Cambridge. The principal theme was the way arm and tube-foot movements are coordinated. Movement in echinoderms, he suggests, is worthy of special study because these are animals in which radial symmetry extends in varying degrees to almost all organ systems of the body, and so the nature of their movement has no counterpart in the activities of bilaterally symmetrical animals. The nervous system exhibits different powers of integration in different parts of the body: spines and pedicellariae are virtually independent effectors, whereas tube feet and arms are subject to a rigid coordination in the interests of the whole animal.

The question whether there are special coordinating centres had much exercised echinoderm biologists, and in this review (7) Smith remarked that 'One might suggest that the nerve ring includes a nerve centre at the base of each arm, and that the neurones of these centres can heighten the general level of excitation within the radial nerve cord of their respective arms'. He reiterates the classic experiment in which starfish arms are autotomized or severed with or without a part of the adjacent nerve ring, experiments that had so influenced earlier thinking. With part of the nerve ring intact, the arm is said to act as a leading arm, with its tip foremost, whereas an arm severed distal to its junction with the ring will move off with its proximal (open) end leading. This experiment does not always produce cut-and-dried results, and after a time the arm with part of the nerve ring attached will often revert to behave like the arm lacking part of the ring. As Smith said, 'there are indications that the general level of activity of the neurons of these supposed centres is not always constant'.

In a synthesis of the relation between structure and function in echinoderms (11) he showed how his work on starfish sensory and motor systems and the distribution and fine anatomy of nerve tracts connecting them enabled a more meaningful examination of behavioral aspects of nervous control. He pointed out that it is not possible to study locomotory coordination of arms and tube feet by simply suspending a starfish in water: the

arms of such a preparation will twist and bend incessantly and the tube-foot will attempt to regain contact with a hard surface. But an inverted starfish placed over a suitably sized cylinder will permit observation of an actively stepping experimental animal.

When such an animal is gently touched on its dorsal surface by a probe, the spread of excitation from the stimulated spot can be followed.

The sequence of responses evoked lead to the conclusion that there is a dual-control mechanism in place, the one peripheral and reflex, the other central and generalized, and this pattern is of significance in the integration of locomotory stepping.

Because the sequence of tube-foot movements is not dependent on a centrally determined rhythm, the integration of stepping activity must be by cycles of activity within the foot itself. Smith therefore advocated in this paper that there should be subsequent study of the effects of localized and graded stimuli applied to a tube foot at different phases of its stepping cycle, and a survey of the different kinds of step that may be associated with various stimuli. Such a study has yet to be done.

The current view on the neural basis of echinoderm behaviour is that coordination is effected by the series of segmental ganglia composing the radial nerve-cord along each arm, and that the circum-oral nerve ring serves mainly to connect the nerve cords one to another. The layout of the nervous system is thus radial rather than central, and 'the dominance that coordinates whole-animal behaviour can shift from one part of the radial nervous system to another' (Cobb 1990). Such an approach helps to explain not only the observed differences in tube-foot response down the length of an arm that Smith and others had observed, but also the progressive change in tube-foot response in severed arms. It is also becoming increasingly evident that responses and hierarchical interactions are dependent on the physiological state of the animal, such as its nutritional condition or the level of illumination in which it has been living before observation.

Smith's achievement was to provide, for the first time, considerable anatomical detail of sensory and motor pathways, giving to subsequent echinoderm workers a framework for further studies using more recent ultrastructural and recording techniques.

Some 15 years after completing experimental work on the echinoderm nervous system, and shortly before leaving Queen Mary College London, Smith published two further reviews on the current status of echinoderm neurobiology. In both (15, 16) he suggested the occurrence of a giant-fibre system in ophiuroids. More recent work by Brehm (1977) has indeed described giant fibres in the ophiuroid arm, from which single-unit action potentials could be recorded. Since then, these large neurons of brittle stars foreseen by Smith have enabled an appreciable increase in knowledge of the echinoderm nervous system at the cellular level.