Dear friends,

It is not many days left until many of us meet in Trapani to talk amphipods and be together with friends. This deep feeling of friendship and respect is what we feel most defines our little society - the Amphipodologists. We meet in small or large groups for workshops or collection-trips, or we work alone in between the meetings - we might not even go to all the meetings, and all the time the friendship is there helping our science along. Many of the marine amphipodologists have met at the IceAGE workshops in Wilhelmshaven (see AN40) and this April in Spala (p. 51) - and next spring there will be an opportunity for the beach-lovers to meet in Crete (p. 50).

This summer has presented us with the sad loss of our good colleague Murat Sezgin of the Sinop University on the Black Sea coast of Turkey. Living legend Torben Wolff of the Zoological Museum in Copenhagen passed away this spring - 97 years old. Torben, a member of the second ‘Galathea’ expedition and one of the first to document life in the abyssal zones of the sea, worked on his beloved deep-sea critters almost until his last day. We also remember Ed Bousfield.

The bibliography for this Amphipod Newsletter presents 419 papers on amphipods. Publications are on broad biodiversity studies, toxicology, ecology and systematics. Lowry and Myers have again presented us with a benchmark paper on the higher classification of the Amphipoda - which we are sure will be further discussed in Trapani.

Last - but not least - the interview of this Amphipod Newsletter presents our hyperiid colleague Wolfgang Zeidler from the South Australian Museum in Adelaide. The distance - both geographic and working on planktonic amphipods more than freshwater or benthic - might be a reason many might not know Wolfgang, but we hope to rectify that a bit with this interview.

Best wishes from your AN Editors,

Wim, Adam, Miranda and Anne Helene
Interviewing anyone 9 timezones away from you will present a few logistic problems. Thus, most of this was done via email, with a very nice skype-session in the end. That means that Wolfgang has written all his answers, as well as a short introduction. So - from here it is all Wolfgang:

To all my dear gammaridean colleagues: because I study hyperiideans, a pelagic group that spend part or all of their life cycle associated with gelatinous plankton, I have had very little direct contact with you, although I am mainly aware of your good work. Also, because there are very few scientists working with hyperiideans, and most are ecologists, I am almost alone in the world of hyperiidean systematics. But be not alarmed for I am quite happy working on my own, although I do appreciate the few interactions and collaborations that I have had with other colleagues.

When and why did you start studying amphipods?

Amphipods were not my first scientific interest. As a young graduate my ambition was to become a fisheries biologist thus combining my recreational passion for fishing with a career. Unfortunately, then as now, there were few jobs anywhere in Australia in marine science. So, according to the old proverb “those who cant do - teach“ I did my penance teaching biology and science in Adelaide schools for 18 months before moving to Sydney to take up a position as Tutor in biology with the University of New South Wales. After about 18 months in the big smoke I was fortunate enough to obtain a scholarship to study for an MSc in Marine Science at the James Cook University of North Queensland, Townsville. At the time the Australian Government was very concerned about the effects of the apparent plague of ‘Crown of Thorns’ starfish on the Great Barrier Reef and so it was relatively easy to obtain grants for shiptime and associated research. Thus, I was able to access (and collect some myself) plankton samples collected fortnightly between Townsville and the GBR (for starfish larvae), over a period of about two years, to study the distribution and abundance of hyperiidean amphipods (Zeidler 1978, 1984). But first I had to overcome the taxonomic impediment of hyperiidean systematics. The literature was generally poor and sometimes difficult to obtain (no internet in those days). But this challenge ignited my passion for systematics and the detective work required.
with the South Australian Museum as ‘Curator of Marine Invertebrates’. So, back home, the circle completed! I soon discovered that my museum title was a misnomer and that my curatorial responsibilities covered all invertebrates (marine, freshwater & terrestrial) except for insects, arachnids and helminths. Sadly much of the collection was in poor shape, and so it was that I lost my way in science for the first few years while I tried to get the collection in order. Also, marine science was not on any government agenda.

However, the SA Museum had a strong focus on arid-zone research and it was relatively easy to organise field trips to most of Central Australia and beyond. One of the most memorable trips was to the Lake Eyre region when it was flooded in the mid-1970’s. On the way I discovered the fauna of the artesian springs of the Great Artesian Basin, resulting in many future trips to the region and other artesian springs in central and north-eastern Australia. The spring fauna is dominated by several species of molluscs, ostracods, one phreatoicid isopod and of course amphipods. For many years I tried to determine if only one widespread species was present or if, as I suspected, speciation had occured in the different, geographically isolated, spring groups. The springs equating to aquatic islands in a desert. Unfortunately, I was unable to make much progress using morphological techniques and, at the time, molecular technology had not been developed or was too expensive. I described two new species, *Austrochiltonia dalhousiensis* and *Phreatochiltonia anophthalma*, and decided to abandon the project for the time being. It was taken up later by my colleague, Dr Rachael King, who has done a great job so far in describing several new species and genera. By then I had returned to the systematics of hyperiideans to complete the obligatory PhD and had no time for gammarids. Having lost my way with amphipods and published papers on mollusca, echinoderms and other crustaceans I decided it was time to tackle the suborder Hyperiidea as a whole, sorting out minor problems and revising the systematics of the group by superfamily with a view to publishing a guide to the world fauna. To date I have completed most of these revisions and the guide book is already up to 700 pages, excluding figures. Looks like it might have to become two volumes.

Sorry that was a rather long-winded answer to the question but I thought you might be interested in my journey.

What are your favourite amphipod species names?

Most hyperiid names are boring, named after localities, expeditions, friends and enemies (if it is an ugly one with filthy habits). So, maybe one of my own, the phreatic, spring related, gammarid *Phreatochiltonia anophthalma*. The generic name readily identifies it as phreatic and the species name refers to it as

![Backyard fisher: the herons finishing off the fish in the backyard pond.](image)

Photo: private
being without eyes. It is a favourite because I had intended to name it for being blind but a good colleague asked the question “how do you know that it can’t see”. I am now more careful when composing names for new taxa.

What amphipod appendage(s) do you like illustrating the best?

I have always enjoyed art so executing my own scientific illustrations is often the most satisfying part of my research. However, unlike gammarids, the segments and coxae of hyperiids are not always clearly defined and drawing whole animals can be frustrating, especially very small species. Some are mature at 2-3 mm.

![Oxycephalus sp.](Image)

*Oxycephalus* sp. Photo: Karen Osborn

What amphipod appendage(s) do you like illustrating the least?

None of them - I like them all! The act of illustrating is an interesting challenge, and it helps the scientist (who should do the illustrations themselves, by the way, not give away this important job to the artists) to see better. We get to really look for the characters – check that they really are there and are not just something we think we see, and this makes the whole job sounder.

Where is your favourite place to collect amphipods?

Collecting hyperiid amphipods is not an easy task and requires the use of a boat and plankton nets, preferably in oceanic waters. Hence, most of my research time has been spent with collections collected by past expeditions, housed in the major museums in Europe and North America. However, there is nothing more exciting than collecting specimens yourself and no place more magnificent than Antarctica. I was fortunate to be on the maiden voyage of the *Aurora Australis* to Heard Island and the Kerguelens during the mid-winter of 1990 and again to Prydz Bay during the summer of 1991, both courtesy of the Australian Antarctic Division, Hobart.

![At Mawson Base in Antarctica.](Image)

At Mawson Base in Antarctica. Photo: private

![Chonerola spinifera.](Image)

*Chonerola spinifera*. Illustration: W.Z.
Places you wished you never tried to collect amphipods?

I have always enjoyed collecting, regardless of the environment, but perhaps digging up, by hand, the rotting carcass of a cow from an artesian spring to collect animals clinging to it would not be repeated too readily. Still there was great delight and satisfaction when amphipods were found.

Describe/name the most memorable amphipod moment(s)?

There are several, each when discovering, amongst collections, representatives of species previously known only from the unique holotype. Often it is the only other specimen in the world. It’s almost magic! Examples are Lanceola longidactyla Vinogradov, 1964 (one more in Copenhagen); Megalanceola remipes (Barnard, 1932) (several in Smithsonian); Chuneola spinifera Vinogradov, 1960 (one in Copenhagen); Scina chelata Vinogradov, 1970 (one in Smithsonian) and Cheloscina antennula Shih & Hendrycks, 1996 (one in Smithsonian). NOTE to editors: there are some that discourage the description of new species based on a unique holotype. While one should be cautious, not publishing would have meant that the above species would have remained unknown and the fact that there are now at least two specimens of the same species would also remain unknown.

I have also had some memorable moments discovering phreatic amphipods in the springs of northern Australia. For a long time Australia was considered too dry to harbour a significant phreatic fauna. We now know that this is far from the truth, especially in north-western Australia. So, it was very exciting to find these animals in ground water, or springs, in parts of the country that are considered a desert. Of course we now know that if we look hard enough we can find these animals where ever springs are found, even if they are only active after a good season of rain.
Describe/name your most memorable amphipod meeting/s)?

International travel from Australia is relatively expensive and time consuming and this, combined with a lack of funds, and little interest in gammarids, has meant that I have made no effort to attend past amphipod meetings. That does not mean that I don’t like my gammarid colleagues!

We know the work with other scientists can shape your life. We are sure many will like to hear about such your experiences if you’d like to share...

Sorry when it comes to hyperiids I am very much a lone wolf, mainly because when I started my hyperiidean journey I was the only one working on the group, as a whole, outside of Russia. Also, here in Adelaide, I was the only marine invertebrate taxonomist and not much has changed since. Meetings with interstate, or international colleagues, usually involved discussions regarding taxonomy in general, with few opportunities for collaboration. Perhaps my joint fieldwork with Dr Winston Ponder, a malacologist from the Australian Museum (now retired), working on the fauna of the artesian springs, is the most significant for me because he encouraged me to pursue my research work and not to be afraid to publish. We spent many years collecting and studying the fauna of the springs of the GAB, highlighting the significance of these fragile habitats in publications etc., resulting in most of them being preserved in national parks. That was a most gratifying result.

Oh! Then there is the collaborative work I did with Dr Lisa Gershwin on the jellyfish fauna of Australia in the early 2000’s. Yes what was I thinking? Well because hyperiids are associated with gelatinous hosts I started to take in interest in them and kind of got sucked in, helped by Lisa’s enthusiasm for everything jellyfish, and taking the opportunity of her presence in Adelaide. It was a very productive collaboration, we published 9 papers (218 pp) and described 2 families, 3 genera and 31 species new to science. Thankfully I have slowly extricated myself from this project but
any other general thoughts/comments?

while a lot of gammaridean taxonomists have been able to include molecular work in their studies this has been more difficult for me with hyperiids. this is because hyperiids, being pelagic, are collected in plankton samples which are almost always first fixed in formaldehyde to preserve the gelatinous plankton. thus, the only way to obtain specimens for molecular work would be for me, or someone who knows hyperiids, to be present when plankton collections come on board to extract hyperiids before samples are preserved. however, i am making some progress in that direction. currently dr charlotte havermans and i are trying to sort out the mess that is the genus *themisto*. charlotte is in charge of the molecular work and i am doing the morphology. we now have material from most parts of the world but we always welcome more material from anyone who is prepared to send it to us. *themisto* is a very common genus in arctic and antarctic waters, sometimes as abundant as krill and an important food source for birds, penguins and other marine predators. note to ecologists playing with *themisto* watch this space!

one of the reasons i have continued with my research – and the reason i am working on the big book – is to pass on the knowledge i have gathered, rather than just go fishing or travelling to art-galleries overseas. much of the information about the species should also end up in worms.

thank you so much for your interest, if you got this far!

anne helene and wolfgang

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**Compilation of Amphipod relevant literature**

Please tell the AN editors and Olli Coleman about your recent publications on amphipods - and send a pdf of your paper. Olli can include it on the server and the editors can include it in the bibliography....
Prof. Dr. Murat Sezgin in Memoriam

Prof. Dr. Murat Sezgin, a complete, brilliant scientist focused on marine biology and an expert on marine Amphipod species of Turkey, passed away on 28.07.2017 because of a deplorable traffic accident near Sivas province, Turkey.

He was born in 1974 in Malatya Province, Turkey. He completed his PhD thesis in 2003 at Ege University, Izmir on “Sublittoral benthic amphipod species of Aegean coasts of Turkey and their bio-ecological features”. Then, he continued his academic career in Sinop University, Faculty of Fisheries where he established a marine benthology team in a few couple of years and became a professor in 2013.

From the beginning of 2016 Prof. Dr. M. Sezgin served both as the vice chancellor and the Dean of his faculty.

During his short but fruitful academic life, he participated in many international scientific activities and in 2008 he was awarded the “Most Promising Young Scientist in the Field of Marine Ecology” by the Commission on The Protection of the Black Sea Against Pollution (Sofia, Bulgaria). He published 95 papers in peer reviewed journals, mainly on marine biology and ecology of the Black Sea. He was the author of more than 45 communications in national and international conferences. He collaborated in umpteen projects, most of which were international or FP7 projects and became a known and respected scientist abroad, particularly in the other Black Sea countries, mostly in Romania, Bulgaria and Ukraine due to his work-discipline and gentle personality.

In his last years, Prof. Sezgin also became a new light for the study of meiobenthology in Turkey by coordinating two international projects and guiding his team for their training, resulting in two PhD and a MSc thesis and several pioneering papers on marine meiofauna. His name has recently been attributed
to a tardigrade species (*Megastygarctides sezginii*), description of which will be published in the upcoming issue of the journal of Marine Biology Research.

Prof. Dr. Murat Sezgin was married and had a 7 years old son who also passed away because of the accident.

Rest in peace my friend, with your beloved son…

*Murat Özbek, Levent Bat & Derya Turk Urkmez*

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**Edward Lloyd Bousfield (1926-2016)**

*in Memoriam*

*It is with a heavy heart and with great sadness that we inform you of the passing of Dr. Edward Bousfield on September 7, 2016*

He was an exceptional scientist, one of the BIG Guns so to speak, a world authority on the systematics of amphipod crustaceans. He discovered and described over 300 new species and he had 22 named in his honour.

Ed joined the Canadian Museum of Nature (CMN), then the National Museum of Canada in 1950 after obtaining his BA and MA at the University of Toronto (1948 and 1949) and PhD at Harvard University (1954). Following his retirement in 1984, he continued as Research Associate at CMN, the Royal Ontario Museum in Toronto and the Royal British Columbia Museum in Victoria. Ed was pivotal to the 20th century growth of the CMN invertebrate collections through his widespread fieldwork which resulted in the contribution of over a million specimens. In 1995, Dr. Paul LeBlond and Ed formally described the large aquatic mega-serpent *Cadborosaurus willsi* in coastal north Pacific marine waters and large deep water lakes of the boreal northern hemisphere. Ed mentored numerous young scientists throughout his illustrious career (a few are still at the museum today) and influenced science through his key roles in scientific societies. In 1978, Ed was elected Fellow of the Royal Society of Canada and in 1985 he received the Government of Canada’s Outstanding Achievement Award. Even at nearly 90 years of age, Ed continued to travel to the CMN from Toronto to work on amphipods, his love of 66 years! Amphipods were definitely his “way of life”.

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From 1993 to the last published issue in 2004 Ed Bousfield was Managing Editor of Amphipacifica, a journal of aquatic systematic biology. The journal was initiated in the early nineties when he was involved (with colleagues) in production of monographic papers on North American Pacific amphipods that proved overly large for standard scientific journals. Amphipacifica was thus set-up to overcome this problem and between 1994 and 2004, three volumes, each of 4 issues were published. In 2006, through the work of Michel E. Hendrickx and his team, all back issues of Amphipacifica were scanned and PDFs made for distribution on CD ROM so that access was made freely available to all amphipodologists. These PDFs are now available through the Aphia database (see: http://www.marinespecies.org/amphipoda/aphia.php?p=sourcelist&sName=Amphipacifica).

**Ed Hendrycks & Kathy Conlan**

An extensive obituary for Ed was published in *The Canadian Field Naturalist* 130, 359-372.

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HUDEC, I., C. FIŠER & J. DOLANSKY 2017. Niphargus diadematus sp. n. (Crustacea, Amphipoda, Niphargidae), an inhabitant of a shallow subterranean habitat in South Moravia (Czech Republic), ---- Zootaxa 4291, 41-60. (Found in spider traps (!) in the Dyje River watershed, Czech Republic. The paper also includes a table listing 19 morphological traits in 64 Niphargus species in the aquilex aggregate)


IACARELLA, J. C., E. J. HUDGINS, J. T. A. DICK & A. RICCIARDI 2017. Predatory behaviour of an invasive amphipod in response to varying conspecific densities under higher-order predation risk. ---- Canadian Journal of Fisheries and Aquatic Sciences, in press. (Gammarus pulex in Ireland)


IWASA-ARAI, T., A. S. FREIRE, A. C. COLOSIO & C. S. SEREJO 2016. Ontogenetic development and re-description of the whale louse Cyamus boopis Lütken, 1870 (Crustacea: Amphipoda: Cyamidae), ectoparasite of humpback whale Megaptera novaeangliae (Mammalia: Cetacea: Balaenopteridae). ---- Marine Biodiversity, in press (A lectotype is chosen. Table 1 lists the 7 species of whale lice reported from Brazil) (Unsure when this was finally publishes WV)

IWASA-ARAI, T., S. SICILIANO & C. S. SEREJO 2017. Life history told by a whale louse: a possible interaction of a southern right whale Eubalaena australis calf with humpback whale Megaptera novangliae. ---- Helgoland Marine Research 71-6 (Cyamus boopis found on stranded right whale in Brazil.)


JAMIESON, A. J., T. MALKOC, S. B. PIERTNEY, T. FUJII & Z. ZHANG 2017. Bioaccumulation of persistent organic pollutants in the deepest ocean fauna. ---- Nature Ecology and Evolution 1, e0051 (Two Hirondellea spp and Bathycallisoma schellenbergi from 10 000m deep contained PCBs and PBDs!!)

JANIAK, D. S., J. N. ADAMS, B. RUBINOFF & R. W. OSMAN 2016. Predator-prey interactions of the polyclad, Euplona gracilis, and the amphipod, Apocorophium lacustre, in the Chesapeake Bay. ---- Estuaries and Coasts 40, 511-523. (Euplona is a significant predator of Apocorophium.)


 JERMACZ, Ł., A. DZIERŻYŃSKA-BIAŁOŃCZYK & A. KOBAK 2016. Predator diet, origin or both? Factors determining responses of omnivorous amphipods to predation cues. ---- Hydrobiologia in press. DOI:10.1007/s10750-106-2917-1


JUDGE, J. & J. P. BARRY 2016. Macrcoinvertebrate community assembly on deep-sea wood falls in Monterey Bay is strongly influenced by wood type. ---- Ecology 97, 3031-3043. (Bathyceradocus n. sp., Seba bathybia and Paronesimoides voightae.)

Gammaropsis longipropodi, Photis stridulus n. sp. (Maando Island) and Podoceropsis clavapes n. sp. (Somaemuldo Island).


JUNG, T. W., J. G. KIM & S. M. YOON 2016. Two new species of pontogeneiid amphipods (Crustacea, Senticaudata, Calliopiidae) from Korean waters. ---- ZooKeys 635, 53-79. (Deals with Eusiroides pilopalpus n. sp. (Jeju Island) and Paramoera dentipleurae n. sp. (also Jeju Island).)


KAIM-MALKA, R. A., D. BELLAN-SANTINI & J.-C. DAUVIN 2016. On some Haploops species collected in the North Atlantic Ocean with the description of Haploops islandicus n. sp. (Crustacea: Gammaridea: Ampeliscidae) (Contribution to the knowledge of the genus Haploops.8.). ---- Zootaxa 4179, 42-76. (Deals with Haploops carinata Liljeborg, 1855 (revived), H. setosa, H. robusta Sars, 1891 (revived) and H. islandica n. sp. (66*59'N, 8*48'W, 1630m). A synoptic table compares the four species, and an illustrated key to all species in the genus is presented.)

KALINKINA, N., A. SIDOROVA, T. POLYAKOVA, N. BELIKNA, N. BEREZINA & I. LITVINNOVA 2016. (Decline in the deepwater benthic communities abundance in the Onego Lake under multifactor influence.) ---- Principy Ekologii 5, 47-68. (In Russian)


KENCHINGTON, E., I. YASHAYAEV, O. S. TENDAL & H. JØRGENSEN 2017. Water mass characteristics and associated fauna of a recently discovered Lophelia pertusa (Scleractinia: Anthozoa) reef in Greenlandic waters. ---- Polar Biology 40, 321-337. (Caprella rinki, Gitana cf rostrata and a stegocephalid reported)

KHEDHRI, I., H. DJABOU & A. AFLI 2016. Does increased connectivity with the Mediterranean Sea improve the ecological status of the macroinvertebrates in the lagoon of Boughrara (SW Mediterranean)? ---- Community Ecology 17, 156-166.


KOBAK, J., Ł. JERMACZ, D. RUTKOWSKA, P. PAWŁOWSKA, L. WITOWSKA & M. POZNANSKA 2016. Impact of predators and competitors on the depth selection of two invasive gammarids. ---- *Journal of Zoology* 301, 174-183. DOI: 10.1111/jzo.12409 (Behaviour of *Dikerogammarus villosus* and *Pontogammarus robustoides* when exposed to a predator goby.)


KODAMA, M., M N. OHTSUCHI & K. KON 2016. A new species of the genus *Rhinoecetes* Just, 1983 (Crustacea: Amphipoda: Ischyroceridae) from Japan. ---- *Zootaxa* 4169, 133-144. (*Rh. spinicaudus* n. sp. from Nabeta Bay, Shizuoka. A key to all *Rhinoecetes* is provided.)


LABAUDE, S., Y. MORET, F. CÉZILLY, C. REULAND & T. RIGAUD 2017. Variation in the immune state of *Gammarus pulex* (Crustacea, Amphipoda) according to temperature: Are extreme temperatures a stress?. ---- *Developmental and Comparative Immunology* 76, 25-33..

LABAY, V. 2016. Review of amphipods of the Melita-group (Amphipoda: Melitidae) from the coastal waters of Sakhalin Island (Far East Russia). III. Genera Abludomelita Karaman, 1981 and Melita Leach, 1814. — Zootaxa 4156, 1-73. (This important monograph deals in depth with the taxa in the Melita-Abludomelita group. As a result, a number of taxa are transferred to other genera. The genus Nurina is synonymized with Melita, while Paraniphargus is revived. Abludomelita rotundactyla is transferred from Melita, with Megamoera aequidentata as a junior synonym; it, A. kilitini n. sp. (Tatar Strait, SW shelf of Sakhalin Island) and A. okhotensis n. sp. (Kashevarova Shoal, Sea of Okhotsk) are fully described, and a key to Abludomelita is presented. In the genus Melita s.l. M. almagosa, M. annandalei, M. cognata, M. dulcicola, M. latiflagella, M. oba, and M. valesi are transferred to Paraniphargus; Melita awa, M. hainanensis and M. plumulosa to Josephosella; Melita amoena, M. breviarticulata, M. denticulata, M. huanghaiensis, M. japonica, M. machaera, M. mucronata, M. rotundactyla and M. unamoena to Abludomelita; Melita leiotelson, M. reidi and M. shiomodari to Tegano. Here described is Melita shimizui sakhalinensis n. ssp (Aniva Bay, Sea of Okhotsk), and a key to North-Pacific Melita is presented. The genus Paraniphargus is discussed, and 2 new genera erected: Barnardomelita n. gen. for Melita matilda, and Ledoyeromelita n. gen. for Melita excavata and probably also M. festiva.)


LAGRUE, C., K. HEAPHY, B. PRESSWELL & R. POULIN 2016. Strong association between parasitism and phenotypic variation in a supralittoral amphipod. — Marine Ecology Progress Series 553, 111-125. (Transorchestia chiliensis)


increase the impact of an invasive species. ---- *International Journal of Parasitology* 47, 291-296. (*Gammarus pulex* in Ireland)


LEVINGS, C.D. 2016. *Ecology of salmonids in estuaries around the world: adaptations, habitats and conservation*. ---- University of British Columbia Press. 388p (online appendices). (Includes several references and examples of the role of estuarine amphipods in salmonid food webs.)


LISCHKA, S. & W. HAGEN 2016. Seasonal dynamics of mesozooplankton in the Arctic Kongsfjord (Svalbard) during year-round observations from August 1998 to July 1999. ---- *Polar Biology* 39, 1859-1878. (*Themisto abyssorum* and unidentified amphipods in table 1, but mainly copepods discussed)


LOBANOVA, A. I. & Y. A. SHUSTIN 2017. (Characteristic features of nutrition habits inherent to fish species found in the littoral zone of the lake Onego.). ---- ??????, 2(163), 52-56. (In Russian)


LOWRY, J. K. & A. A. MYERS 2016. Zaramillidae, a new amphipod family from the subantarctic Kerguelen Islands (Amphipoda, Senticaudata, Gammaroidea,
Zaramillidae fam. nov.). ---- *Zootaxa* 4169, 387-389. (Contains only *Zaramilla kergueleni*.)

LOWRY, J. K. & A. A. MYERS 2017. A Phylogeny and Classification of the Amphipoda with the establishment of the new order Ingolfiellida (Crustacea: Peracarida). ---- *Zootaxa* 4265, 1-89. (This is a benchmark paper, the final one in the series of monographs on amphipod classification by these authors. The proposed new classification is much too extensive to be repeated here, but the new families are included in the "New Taxa" listings.)

LUARTE, T., C. C. BONTA, E. A. SILVA-RODRIGUEZ, P. A. QUIJON, C. MIRANDA, A. A. FARIAS & C. DUARTE 2016. Light pollution reduces activity, food consumption and growth rates in a sandy beach invertebrate. ---- *Environmental Pollution* 218, 1147-1153. (A Chilean study on *Orchestoidea tuberculata*.)


MALEK-HOSSEINI, M. J. & A. ZAMANI 2017. A checklist of subterranean arthropods in Iran. ---- *Subterranean Biology* 21, 19-46. (10 sp of *Niphargus* (all from caves and karstic springs).)

MANKO, M. J., A. W. SŁOMSKA & K. JAZDZEWSKI 2017. Siphonophora of the Gulf of Aqaba (Red Sea) and their associations with crustaceans. ---- *Marine Biology Research* 11, 460-485. (Hyperiid associates listed in Table 2)


MATSUNO, K., A. YAMAGUCHI, A. FUJIWARA, J. ONODERA, E. WATANABE, N. HARADA & T. KIKUCHI 2016. Seasonal changes in mesozooplankton swimmer community and fecal pellets collected by sediment trap moored at the Northwind Abyssal Plain in the western Arctic Ocean. ****Bulletin of the Fisheries Faculty of Hokkaido University 66, 77- 85. (Themisto libellula)


MESZNER, U. & M. I. ZETTLER 2016. Die aktuelle Verbreitung von Amphipoda (Crustacea) im Verlauf der Oberen Havel. ****Lauterbornia 81, 57-69 (Eight amphipod species in Table 1)


MIKKELSEN, N. & T. PEDERSEN 2017. Invasive red king crabs feed on both spawned-out capelin and their eggs. ---- *Marine Ecology Progress Series* 563, 139-155.


MONROY-VELAZQUEZ, L. V., R. E. RODRIGUEZ-MARTINEZ & F. ALVAREZ 2017. Taxonomic richness and abundance of cryptic peracarid crustaceans in the Puerto Morelos Reef National Park, Mexico. ---- *Peer Journal* 5, e3411. (Amphipods listed in Table 1)


MORAIS, P. & M. REICHARD 2017. Cryptic invasions: A review. ---- *Science of the Total Environment*, in press. (Cryptic invasions are either invasions of non-native lineages within their species native range, or invasions of non-native species, that are not recognized because of misidentification as a a different species.)


MUGGELBERG, L. L., K. E. HUFF HARTZ, S. A. NUTILE, A. D. HARWOOD, J. R. HEIM, A. P. DERBY, D. P. WESTON & M. J. LYDY 2017. Do pyrethroid-resistant *Hyalella azteca* have greater bioaccumulation potential compared to non-resistant populations? Implications for bioaccumulation in fish. ---- *Environmental Pollution* 220, 375-382.


MYERS, A. A. 2016. Amphipoda (Crustacea) from Palau, Micronesia: families Maeridae and Melitidae. ---- *Zootaxa* 4170, 451-474. (Deals with *Elasmopus alalo*, *E. laminischia* n. sp., *Linguimaera siaes* n. sp., *?Parelasmopus setiger* (possibly a species complex), *Quadrimaera pacifica*, *Q. serrata*, and *Dulichiella belun* n. sp.)

MYERS, A. A., J. K. LOWRY & Z. BILLINGHAM 2016. A new family, genus and species of freshwater amphipod *Australomicropotopus megacoxa* gen. nov., sp. nov. (Senticaudata, Corophiidea, Microprotopoidea, Australomicropopidae fam. nov.) from Australia. ---- *Zootaxa* 4161, 412-418. (From freshwater streams in Victoria.)


NARAHARA-NAKANO, Y., T. NAKANO & K. TOMAHARA 2017. Deep-sea amphipod genus *Eurythenes* from Japan, with a description of a new *Eurythenes* species from off Hokkaido (Crustacea: Amphipoda: Lysianassoidea). ---- *Marine Biodiversity*, in press. (Deals with *E. aequilatus* n. sp. (Sea of Okhotsk off Hokkaido, 1580m) and *E. magellanicus*)


ORTIZ, M. & R. LALANA 2016. Estado actual del conocimiento de los anfípodos (Crustacea, Peracarida), de Cuba: Morfología, Historia, Taxonomía y Bibliografía. (Current state of knowledge of the amphipods (Crustacea, Peracarida), of Cuba: Morphology, History, Taxonomy and Bibliography.) ---- Revista Investigaciones Marinas 36(1), 1-19. (In Spanish. Table 1 furnishes data on all species described from Cuban waters.)

ORTIZ, M. & I. WINFIELD 2017. A new species of Nuuanu (Crustacea, Amphipoda, Nuuanidae) from a Caribbean coral reef with identification keys to males and females of Nuuanu species. ---- Zootaxa 4294, 197-208. (N. jaumei n. sp from Puerto Morelos reef, Quintana Roo, Mexico.)


ÖZBEK, M., A. TASDEMOR & S. YILDIZ 2016. (Benthic macroinvertebrates of Adigüzel Reservoir (Denizli, Turkey).) ---- Ege Journal of Fisheries and Aquatic Sciences 33, 259-263. (In Turkish. Pontogammarus robustoides)

and production of macrozoobenthos in the macroalgal belt at Hansneset, Kongsfjorden, after 15 years. ---- *Polar Biology* 39, 2065-2076.


PEART, R. A. 2017. A synopsis and key to the genus *Exampithoe* K.H.Barnard, 1925 (Amphipoda: Senticaudata: Ampithoidae), with descriptions of five new species. ---- *Journal of Crustacean Biology* 37, 63-75. (Deals with *E. ecklonicola* n. sp. ((Cape Banks, NSW), *E. helena* n. sp. (Geraldton, W. Austr.), *E. malus* n. sp. Dampier archipelago, W. Austr.), *E. otway* n. sp. Cape Otway, Vict.) and *E. vasse* n. sp. nr Yallingup, W. Austr.). A key to all *Exampithoe* is provided. *E. waratah* is transferred to *Ampithoe*.


PEREZ-SCHULTHEISS, J. 2016. Synopsis of the superfamily Lysianassoidea (Amphipoda: Gammaridea) in Chile. ---- *Boletin del Museo Nacional de Historia Natural, Chile* 65, 193-246. (A most welcome critical survey of all that is known of Chilean Lysianassoidea. It contains a fair number of large range extensions, as well as some classificatory changes: the new genus *Exuristes* (type *Uristes yamana*) is erected for *E. yamana* and *E. serratus*, both transferred from *Uristes*, while *Tryphosella schellenbergi* and *T. serrata* are transferred to *Uristes*, and the classification of *Uristes paramoi* in this genus is confirmed. Maps showing the Chilean distribution of all species and a key to Chilean Lysianassoidea are also provided.)

PETERS, J. 2017. We’ve polluted hadal zones (deep sea trenches) before we even could explore them. ---- Scienceosaurus, 3 pp


RASTRICK, S. P. S. & N. M. WHITELEY 2017. Comparison of whole animal costs of protein synthesis among polar and temperate populations of the same species of gammarid amphipod. ------ *Comparative Biochemistry and Physiology A* 207, 100-106 (*Gammarus oceanicus*)

RATTANAMA, K., M. S. PATTARATUMRONG, P. TOWATANA & K. WONGKAMHAENG 2016. Three new records of gammarid amphipod in Songkhia Lake, Thailand. ---- *Tropical Life Sciences Research* 27 (Suppl. 1), 53-61 (*Hyale dollfusi, Grandidierella megnae & Hourstonius japonica*).

REMAIL, T. M., S. L. SIMPSON & D. F. JOLLEY 2017. Effects of enhanced bioturbation intensities on the toxicity assessment of legacy-contaminated sediments. ---- *Environmental Pollution* 226, 335-345. (*Victoriopisa australiensis* and *Melita plumulosa* test animals.)


RODRIGUEZ, M., L. C. ARMENDARIZ & A. R. CAPITULO 2017. A new genus and species of Ingolfiellidae (Crustacea, Ingolfiellida) from the hyporheic zone in the Sierra de la Ventana, and its biogeographic relevance. ---- *Zootaxa* 4290, 99-112. (Deals with *Yacana ventania* n. gen. n. sp, a taxon apparently most closely related to the South African ingolfiellids.)


RUFCHAEI, R., S. H. HOSEINIFAR, A. MIRJAZANI & HIEN VAN DOAN 2017. Dietary administration of *Pontogammarus maeoticus* extract affects immune response, stress resistance, feed intake and growth performance of Caspian roach (*Rutilus caspicus*) fingerlings. ---- *Fish & Shellfish Immunology* 63, 196-200

Atlantic sandy beaches of Brazil and Spain. ---- *Marine & Freshwater Research* 67, 1634-1643.


SCHEPIS, W. R., T. V. MEDIEROS, S. A. SILVA & D. M. S. ABESSA 2016. (Acute toxicity and contamination by metals in the sediments of the Rio dos Bugres, Ilha de São Vicente, SP.) ---- *Brazilian Journal of Aquatic Sciences and Technology* 20, 42- ??. (In Portuguese)


SCHRÖTER, F. 2016. Community structure of amphipods from sediment traps in the eastern Fram Strait - interactions with environmental parameters in a changing Arctic. ---- *Master thesis, Eberhard Karls Universität, Tübingen* (not seen)

SEPULVEDA, R. D. & N. VALDIVIA 2017. Macrobenthic community changes of intertidal sandy shores after a mega-disturbance. ---- *Estuaries and Coasts* 40, 493-501 (A Chilean study after earthquake and tsunami)

SEYMOUR, M., K. SEPPÄLÄ, E. MÄCHLER & F. ALTERMATT 2016. Lessons from the macroinvertebrates: species-genetic diversity correlations highlight important dissimilar relationships. ---- *Freshwater Biology* 61, 1819-1829. *(Gammarus fossarum complex)*


SHIMOMURA, M. & K. TOMIKAWA 2016. *Epimeria abyssalis* sp. n. from the Kuril-Kamchatka Trench (Crustacea, Amphipoda, Epimeridae). ---- *ZooKeys* 638,
125-142. (From 5480m deep, the deepest Epimeria ever collected. With a key to N. Pacific Epimeria.)


SIDOROV, D. A., A. D. KATZ, S. J. TAYLOR & M. V. CHERTOPRUD 2016. A reassessment of the phylogenetic utility of genus-level morphological characters in the family Bogidiellidae (Crustacea, Amphipoda), with description of a new species of Eobogidiella Karaman, 1981. ---- ZooKeys 610, 23-79. (Eobogidiella venkataramani n. sp., known from a single specimen from the Shirawati River basin, Western Ghat, India. The authors discuss the value of various morphological characters in the Bogidiellidae, and consider the generic placement of the present new taxon as preliminary.)

SIDOROV, D. A. & G. V. SAMOKHIN 2016. Kruberia abchasica, a new genus and species of troglobiont amphipods (Crustacea: Gammaridae) from Krubera Cave (Western Transcaucasia). ---- Arthropoda Selecta 23, 371-379. (This new genus is close to the typhlogammarids, recently merged with Gammaridae by Sket and Hou)


SORENTOINO, R., J. ALVES, R. JOHNSSON & A. R. SENNA 2016. A new species of Cyphocarididae (Crustacea, Amphipoda, Lysianassoidea) from off the northeastern Brazilian coast. ---- Zootaxa 4161, 345-356. (Cyphocaris pedroi n. sp., from a tuna stomach, St Peter and St Paul Archipelago. With a key to world Cyphocaris species.)


SPIKKELAND, I., K. M. OLSEN, B. KINSTEN & G. KJELLBERG 2016 (The freshwater amphipod Crangonyx pseudogracilis shown to occur in Norway). ---- Fauna, Oslo 69, 24-34. (In Norwegian. Found on Jæren, SW Norway in 2012)

SPILLMONT, N., A. HACHE\nT, M. A. FAASSE, J. JOURDE, C. LUCZAK, L. SCURONT & C. ROLET 2016. First records of Ptilohyale littoralis (Amphipoda: Hyalidae) and Boccardia proboscidea (Polychaeta: Spionidae) from the coast of the English Channel: habitat use and coexistence with other species. ---- Marine Biodiversity, in press. (P. littoralis found in numbers at Wimereux, Pas de Calais) (NB. I can't find it VW)

STAMATAKI, E. & A. PAVLOPOULOS 2016. Non-insect crustacean models in developmental genetics including an encomium to Parhyale hawaiensis. ---- Current Opinion in Genetics and Development 39, 149-156. (Encomium means fulsome, enthusiastic praise; I looked it up)


STRAUB, S., P. E. HIRSCH & P. BURKHARDT-HOLM 2017. Biodegradable and petroleum-based microplastics do not differ in their ingestion and excretion but in their biological effects in a freshwater invertebrate Gammarus fossarum. ---- International Journal of Environmental Research and Public Health14, 774


SUAREZ-JIMENEZ, R., C. D. HEPBURN, G. A. HYNDES, R. J. McLEOD, R. B. TAYLOR & C. L. HURST 2017. Importance of the invasive macroalga Undaria pinnatifida as trophic subsidy for a beach consumer. ---- Marine Biology 164, art. 113 (A New Zealand study, featuring Bellorchestia quoyana.)

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TATO, R. & J. MOREIRA 2017. Two new species of the suborder Senticaudata (Crustacea: Amphipoda) from the upper continental slope off Galicia (NW Iberian Peninsula). ---- *Zootaxa* 4300, 217-237. (Deals with *Pareurystheus vitucoi* n. sp. and *Photis guerrai* n. sp. (both Ferrol Canyon, 43°54´N, 8°57´W, 1005m) Keys to the genera of *Promedieinae*, to *Pareurystheus* species and to Atlantic *Photis* species are provided.)


TOMIKAWA, K., T. NAKANO, A. SATO, Y. ONODERA & A. OHTAKA 2016. A molecular phylogeny of *Pseudocrangonyx* from Japan, including a new subterranean species (Crustacea, Amphipoda, Pseudocrangonyctidae). ---- *Zoosystematics and Evolution* 92, 187-202. (Deals with *Pseudocrangonyx gudariensis* Tomikawa & Sato n. sp. from the Aomori pref., N. Honshu. There are several as yet undescribed species in this genus in Japan.)


VERESHCHAKA, L. & L. L. ANOKHINA 2017. The benthoneuston of the Black Sea: composition and environmental factors influencing its nocturnal dynamic. ---- *Continental Shelf Research* 140, 60-74 (Not seen)


WANG, N., J. L. KUNZ, C. D. IVEY, C. G. INGERSOLL, M. C. BARNHART & E. A. GLIDEWELL 2017. Toxicity of Chromium (VI) to two mussels and an amphipod in water-only exposures with or without a co-stressor of elevated temperature, zinc, or nitrate. ---- *Archives of Environmental Contamination and Toxicology* 72, 449-460 (The amphipod is *Hyalella azteca*).


WINFIELD, I., M. E. HENDRICKX & M. ORTIZ 2017. A new deep-water species of *Trischizostoma* (Crustacea: Amphipoda: Gammaridea: Trischizostomatidae) from western Mexico, NE Pacific Ocean. ---- *Journal of the Marine Biological Association UK* 97, 141-149. (T. *unam* n. sp. from 17°10'N, 101°37'W, c 1400m)


WONGKAMHAENG, K., P. DUMRONGROJWATTANA & M. S. PATTARATUMRONG 2016. Two new species of *Floresorchestia* (Crustacea, Amphipoda, Talitridae) in Thailand. ---- *ZooKeys* 635, 31-51. (Deals with *F. boonyanusithii* n. sp. (Phutsa Reservoir, NE Thailand) and *F. buraphana* n. sp. (Chonburi, E. Thailand).)
YAMAMORI, L. & M. KATO 2017. The macrobenthic community in intertidal sea urchin pits and an obligate inquilinism of a limpet-shaped trochid gastropod in the pits. ---- *Marine Biology* 164, art. 61 (*Aoroides rubellus* and *Maera serratipalma*.)


ZEIDLER, W. 2016. A review of the families and genera of the superfamily Platysceloidea Bowman & Gruner, 1973 (Crustacea: Amphipoda: Hyperiidea), together with keys to the families, genera and species. ---- *Zootaxa* 4192, 1-136. (Another thorough revision of a segment of the Hyperiidea. There are keys to all families, genera and species, as well as extensive descriptions of the type species of all genera. New taxa are the families Eupronoidae, for the genera *Eupronoe* and *Paraprnone*, Amphithryidae, for the genera *Amphithyris*, *Amphithyropsis* n. gen, and *Paralycaea*, and Thamneidae, for the genus *Thamneus.*)


NEW TAXA

New order

Ingolfiellida Hansen, 1903 (new status)

New families and subfamilies

Adeliellidae Lowry & Myers, 2017
Ambasiidae Lowry & Myers, 2017
Amphithyridae Zeidler, 2016
Australomicroprotopidae Myers, Lowry & Billingham, 2016
Derjuginiidae Lowry & Myers, 2017
Eupronoidae Zeidler, 2016
Pakynidae Lowry & Myers, 2017
Parargissidae Lowry & Myers, 2017
Pseudamphilochidae Lowry & Myers, 2017
Thamneidae Zeidler, 2016
Vemanidae Lowry & Myers, 2017
Zaramillidae Lowry & Myers, 2016

New genera and subgenera

Amphityropsis Zeidler, 2016
Australomicroprotopus Myers, Lowry & Billingham, 2016
Barnardomelita Labay, 2016
Eusceliotes Stebbing, 1888 (rev.)
Exuristes Perez-Schultheiss, 2016
Fluviadulzura Myers, Lowry & Billingham, 2017
Kruberia Sidorov & Samokhin, 2016
Ledoyeromelita Labay, 2016
Lowryella Morino & Miyamoto, 2016
Pakynus Lowry & Myers, 2017 (nom. nov)
Persianorchestia Monttazi, Lowry & Hekmatari, 2017
Solitroides Suzuki, Nakano, Nguyen, Nguyen, Morino & Tomikawa, 2017
Yacana Rodriguez, Armentariz & Capitulo, 2017

New species and subspecies:

abchasica Sidorov & Samokhin, 2016 (Kruberia) Gammaridae
abyssalis Shimomura & Tomikawa, 2016 (Epimeria) Epimeriidae
aequilatus Narahara-Nakano, nakano & Tomikawa, 2017 (Eurythenes) Eurytheneidae
alpheus Delic, Svara, Coleman, Trontelj & Fiser, 2017 (Niphargus) Niphargidae
alpinus Alther, Fiser & Altermatt, 2016 (Gammarus) Gammaridae
anchialinus Delic, Svara, Coleman, Trontelj & Fiser, 2017 (Niphargus) Niphargidae
antipodes Delic, Svara, Coleman, Trontelj & Fiser, 2017 (Niphargus) Niphargidae
arethusa Delic, Svara, Coleman, Trontelj & Fiser, 2017 (Niphargus) Niphargidae
arraialensis Ros, Lacerda & Guerra-Garcia, 2017 (Pseudaeuginella) Caprellidae
belun Myers, 2016 (Dulichiella) Melitidae
boonyanusithii Wongkamhaeng, Dumrongrojwattana & Pattaratumrong, 2016 (Floresorchestia) Talitridae
bulbodigitus Jung, Kim, Soh & Yoon, 2016 (Eusirus) Eusiriidae
buraphana Wongkamhaeng, Dumrongrojwattana & Pattaratumrong, 2016 (Floresorchestia) Talitridae
cajasi Alonso & Jaume, 2017 (Hyalella) Hyalellidae
carinata Liljeborg, 1855 (Haploops) (rev.) Ampeliscidae
clavapes Jung, Choi, Kim, Yoon 2017 (Podiceropsis) Photidae

cylindrica Zettler & Glück, 2016 (Wallametopida) Stenothoidae
dentipleurae Jung, Kim & Yoon, 2016 (Paramoera) Pontogeneiidae
diadematus Hudic, Fiser & Dolansky, 2017 (Niphargus) Niphargidae
doli Delic, Svara, Coleman, Trontelj & Fiser, 2017 (Niphargus) Niphargidae
ecklincola Peart, 2017 (Exampithea) Amphipodidae
elegantulus Hou, in Zhang & Hou, 2017 (Pseudocrangonyx) Pseudocrangonyctidae
fjaka Delic, Svara, Coleman, Trontelj & Fiser, 2017 (Niphargus) Niphargidae
gudariensis Tomikawa & Sato in Tomikawa et al., 2016 (Pseudocrangonyx) Pseudocrangonyctidae
guerrai Tato & Moreira 2017 (Photis) Photidae
heike Tomikawa, Tanaka & Nakano, 2016 (Priscomilitaris) Priscomilitaridae
helenae Peart, 2017 (Exampithea) Amphipodidae
hosseiniei Esmaei-Rineh, Sari, Fiser & Bargrizaneh, 2017 (Niphargus) Niphargidae
icapui Andrade & Senna, 2017 (Cymadusa) Amphipodidae
iceage Jimenez Campean & Coleman, 2017 (Sicafodi) Sicafodidae
ilamensis Esmaei-Rineh, Sari, Fiser & Bargrizaneh, 2017 (Niphargus) Niphargidae
incarocai Alves, Jonsson & Senna, 2016 (Elasmopus) Maeridae
islandica Kaim-Malka, Bellan-Santini & Dauvin, 2016 (Haploops) Ampeliscidae
jaumei Ortiz & Winfield, 2017 (Nuuann) Nuuannuidae
klitinii Labay, 2016 (Abudomelita) Melitidae
laminischia Myers, 2016 (Elasmopus) Maeridae
maculanigra Zeina & Asakura, 2017 (Cerapus) Ischyroceridae
magnaoocularis Eun, Hendrycks & Kim (Protohyale) Hyalidae
malus Peart, 2017 (Exampithea) Amphipodidae
megacoxa Myers, Lowry & Billingham, 2016 (Australomicroprotopterus) Australomicroprotoptidae
meufong Hughes & Kaji, 2016 (Quadrisvisio) Maeridae
motokawai Suzuki, Nakano, Nguyen, Nguyen, Morino & Tomikawa, 2017 (Solitroides) Talitridae
muota Fiser, Konec, Alther, Svara & Altermatt, 2016 (Niphargus) Niphargidae
murimali Fiser, Konec, Alther, Svara & Altermatt, 2016 (Niphargus) Niphargidae
nirvana Montazi, Lowry & Hekmatari, 2017 (Persianorches) Talitridae
okhotensis Labay, 2016 (Abudomelita) Melitidae
otway Peart, 2017 (Exampithea) Amphipodidae
pachacutesi Alves, Jonsson & Senna, 2016 (Elasmopus) Maeridae
pedrol Sorrentino, Alves, Jonsson & Senna, 2016 (Cyphocaris) Cyphocarididae
persicus Esmaei-Rineh, Sari, Fiser & Bargrizaneh, 2017 (Niphargus) Niphargidae
piloi Myers, Trivedi, Gorsavi &Vahhrajani, 2017 (Parhyale) Hyalidae
pilopalus Jung, Kim & Yoon, 2016 (Eusiroides) Pontogeneiidae
pincikovae Delic, Svara, Coleman, Trontelj & Fiser, 2017 (Niphargus) Niphargidae
queshmensis Laeyghi & Monttazi, 2017 (Ampithee) Amphipodidae
rasae Andrade & Senna, 2017 (Cymadusa) Amphipodidae
robusta Sars, 1891 (Haploops) (rev.) Ampheliscidae
robustimana Andrade & Senna, 2017 (Ampithee) Amphipodidae
ruffoi Latella & Vonk, in Davolos et al. 2017 (Cryptorchestia) Talitridae
ssp sakhalinensis Labay, 2016 (Melita shimizui) Melitidae
schioettei Just, 2017 (Cephaloecetes) Ischyroceridae
siaes Myers, 2016 (Linguimae) Maeridae
sohrevardensis Esmaei-Rineh, Sari, Fiser & Bargrizaneh, 2017 (Niphargus) Niphargidae
spinicauda Myers, Lowry & Billingham, 2017 (Fluviadulzura) Hadziidae
spinicaudus Kodama, Ohtsuchi & Kon. 2016 (Rhinoecetes) Ischyroceridae
stiridulus Jung, Choi, Kim, Yoon 2017 (Photus) Photidae
styx Fiser, Konec, Alther, Svara & Altermatt, 2016 (Niphargus) Niphargidae
tartarugaes Andrade & Senna, 2017 (Cymadusa) Amphipodidae
telukrimau Lim, Azman, Takeuchi & Othman, 2017 (Pseudaeginella) Caprellidae
trindadensis Andrade & Senna, 2017 (Cymadusa) Amphipodidae
uai Bastos-Pewreira & Ferreira, 2017 (Spelaeogammarus) Artesiidae
unam Winfield, Hendrickx & Ortiz, 2017 (*Trischizostoma*)

*Trischizostomatidae*

vasse Pear, 2017 (*Exampithoe*)

*Ampithoidae*

venkataramani Sidorov, Katz, Taylor & Chertoprud, 2016 (*Eobogidiella*)

*Bogidiellidae*

ventania Rodriguez, Armendariz & Capitulo, 2017 (*Yacana*)

*Hyalellidae*

viricochai Alves, Jonsson & Senna, 2016 (*Elasmopus*)

*Maeridae*

vitukoi Tato & Moreira, 2017 (*Pareurystheus*)

*Protomedeiinae*

wadai Morino & Miyamoto, 2016 (*Lowryella*)

*Talitridae*

yahuarhuaci Alves, Jonsson & Senna, 2016 (*Elasmopus*)

*Maeridae*

ygara Andrade & Senna, 2017 (*Cymadusa*)

*Ampithoidae*

yupanquii Alves, Jonsson & Senna, 2016 (*Elasmopus*)

*Maeridae*

**Taxonomic overview**

*Adeliellidae*

**Ambasiidae**

*Ampeliscaidae*

Haploops *carinata* (rev.), *islandica*, *robusta* (rev.)

*Amphithyridae*

**Amphithyropsis**

*Ampithoidae*

Ampithoe *queshmensis*, *robustimana*

Cymadusa *icapui*, *rasae*, *tartarugae*, *trindadensis*, *ygara*

*Exampithoe* *ecklonicola*, *helenae*, *malus*, *otway*, *vasse*

*Artesiidae*

Spelaeogammarus uai

*Australomicroprotopidae*

**Australomicroprotopus megacoxa**

*Bogidiellidae*

Eobogidiella *venkataramani*

*Caprellidae*

Pseudaeginella *arraialensis*, *telukrimau*

*Cyphocarididae*

Cyphocaris *pedroi*

*Derjugianidae*

*Epimeriidae*

Epimeria *abyssalis*

*Eupronoidae*

Eurytheneidae

Eurythenes *aequilatus*

*Eusiridae*

Eusirus *bulbodigitus*

*Gammaridae*

Gammarus *alpinus*

*Kruberia abchasica*

*Hadziidae*

**Fluvialdulzura spinicauda**

*Hyalidae*

Parhyale *piloi*

Protohyale *magnaocularis*

*Hyalellidae*

Hyalella *cajasi*

*Ingolfiellidae*

Ingolfiellidae

Yacana *ventania*

*Ischyroceridae*
Cephaloecetes schioettei
Cerapus maculanigra
Rhoecetes spinicaudus

Maeridae
Elasmopus incarocai, laminischia, pachacutesi, viracochai, yahuarhuaci, yupanquii
Linguimaera siaes
Quadrivisio meufong

Melitidae
Abludomelita klinii, okhotensis
Barnardomelita
Dulichiella belun
Ledoyeromelita
Melita shimizui sakhalinensis

Niphargidae
Niphargus alpheus, anchialinus, antipodes, arethusa, diadematus, doli, fjakae,
   hosseineiei, ilamensis, muotae, murimali, persicus, pincikovae, sohrevardensis,
   styx

Nuuanuidae
Nanopalpus
Nuuanu jaumei

Pakynidae
Pakynus

Parargissidae
Parascelidae
Eusceliotes (rev.)
Photidae
Photis guerrai, stridulus
Podoceropsis clavapes

Pontogeneiidae
Eusiroides pilopalpus
Paramoera dentopleurae

Priscomilitaridae
Priscomilitaris heike

Protomedeiinae
Pareurystheus vitucoi

Pseudamphilochidae
Pseudocrangonyctidae
Pseudocrangonyx elegantulus, gudariensis

Sicafodiidae
Sicafodia iceage

Stenothoidae
Wallametopa cylindrica

Talitridae
Cryptorchestia ruffoi
Floresorchestia boonyanusithii, buraphana
Lowryella wadai
Persianorchestia nirvana
Solitroides motokawai

Thamneidae
Trischizostomatidae
Trischizostoma unam

Uristidae
Exuristes
Vemanidae
Zaramillidae

Wim
International Sandy Beaches Symposium (ISBS) – 25 to 29 of May 2018

I am pleased to announce that the 8th International Sandy Beaches Symposium will take place from the 25th to the 29th of May at the Institute for Marine Biology Biotechnology and Aquaculture, Hellenic Centre for Marine Research, Crete, Greece. Many advances have been made since the establishment of sandy beach ecology as a discipline (1st symposium, 1983, Port Elizabeth, South Africa). Below, in a nutshell:

A brief summary of the sandy beach research

Morphophysical aspects were firstly approached and interactions of energy and material were identified as responsible of shaping a beach “personality”. With this background central paradigms about features of the resident fauna and biodiversity patterns were developed. Actually most of them are based on amphipods, as they usually dominate (in abundance) temperate and subtropical beaches. The human-environment perspective appeared soon to be a challenge. For a long time the “pristine condition” was the preferred target of sandy beach research, whether as a baseline or as an impact-control study. But it became more and more evident that the human component cannot be escaped. We actually need to know more about the impacts and their dynamics, if we want to get a clear picture of the system. That is why lately the definition of beaches as “social-ecological systems” is getting more and more in use. On the other hand, while the human presence on sandy beaches, is an obvious feature (you can check yourself in the summer), it is not so easy to address and to share research with beach stakeholders. This is definitely an open challenge.

I am then particularly keen to invite to ISBS2018 researchers and students through the amphipod newsletter, as I believe that this lively and interdisciplinary group will strongly contribute to a correct species-environment vision, often biased in favour of general models, loved by managers but perhaps too general to capture the intrinsic diversity of beaches.


Looking forward to meet you in Crete,

Lucia
IceAGE amphipods

Senckenberg am Meer (DZMB) is housing a huge amount of crustacean material collected during two expeditions in 2011 and 2013 via IceAGE (Icelandic marine Animals: Genetics and Ecology). The Icelandic waters are of particular interest for studying patterns of diversity with depth, especially for comparing the diversity profiles with depth for amphipods with various functional properties. The animals are from 56 areas between Greenland and Norway from 150 to 3000 m depth. So far 26 461 of the 58 000 amphipod individuals are sorted by numerous enthusiastic colleagues to 42 (!) families during two amphipod workshops in Wilhelmshaven, Germany (see AN40 and Spala, Poland (photos here); the first workshop was entirely funded by the Volkswagen Stiftung. The second IceAGE amphipod workshop piggybacked on the Synthetis and 7th IceAGE workshop in Spala, April 2017.

Anne-Nina Lörz and Ania Jaźdżewska

Photos by Ania Jaźdżewska and Christian Bomholt
Amphipods take a bite out of the headlines

In August the world became excited about amphipods (as they should!) but for a very different reason than their morphological beauty!

A 16 year boy from Melbourne, Australia was merged in the shallow waters at Dendy Street Beach, Brighton, Australia and when he emerged half an hour later it appeared he had been bitten and ‘attacked’ by some form of tiny marine creatures.

Once out, this story sent the world’s Press into a frenzy! Pictures of the teenager’s feet dripping with blood were sent around the world. Warnings were put on webpages in case the images caused some distress.

In Italy the news had a title like “marine insects attack a tourist!”

In UK one person said “I think the thing I love best about this story is how the dad investigated it especially the part about him trying all the different kinds of food. I’ve never heard of amphipods doing anything like this but I guess the Australian ones are just meaner, like every other animal there.”

What could have bitten him? News reports were everything from piranhas to ‘Sea lice’!

Its amphipods! Through the media the pressure was on to name them and name them now. Alastair Poore said: “Researchers at Museums Victoria have identified the likely suspect of the "sea lice" attack in Melbourne as lysianassid amphipods. Rare to see amphipods in the news like this!”

Jo Taylor and Genefor Walker-Smith responded to the media on pressure surrounding the "sea bug attack" and why it matters to get the fact right: https://www.timeshighereducation.com/blog/australian-sea-bug-attack-scientists-getting-our-facts-right-and-why-it-matters

Meanwhile this news story was the topic of much discussion on the amphipod Facebook page: https://www.facebook.com/groups/238356639577927/

So there we have it, it WAS amphipods but they don’t usually ‘attack’ like this but it did make the headlines and allow us amphipod workers to get people to understand and appreciate our beloved animals more.

Related articles:


Miranda
Jim Lowry and Alan Myers have published their third paper on the higher taxonomic classifications of the Amphipods this year. Here we present figure 11 from the paper, as a summary of their new classifications after removing Ingolfiellidea to a separate order. The remaining order Amphipoda is now divided into six suborders with respective infraorders and parvorders.